

Changes of chloroplast structure in lichenized algae



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Introduction

Lichens are symbiotic organisms composed of a fungal partner (mycobiont) and photosynthetic partner represented by green alga or cyanobacteria (photobiont). Both symbionts undergo a variety of structural, physiological and biochemical modifications as a result of lichenization (Galun, 1988). Photobionts are more or less changed by the influence of the fungal partner. The most frequent changes are reduction of cell size, changes of reproductive strategies and various modifications of some organelles and structures on subcellular level (chloroplast, thylakoids, pyrenoid, pyrenoglobuli, dictyosomes, etc.) (e.g. Bubrick, 1988; Ahmadjian, 1992; Tschermak-Woess, 1995). However, the changes of the algal cells are not permanent and when the cells are freed from the fungal hyphae, they resume after several divisions their original size and characteristics (Ahmadjian, 1992).

The most frequently occuring phycobiont in lichens is Trebouxia (Asterochloris resp.). A basic character in systematic of trebouxioid algae represents chloroplast morphology. However, in different ontogenetic, physiological and ecological stages, chloroplast may markedly vary.

Objectives

Cladoniaceae

 Isolation and identification of photobionts from four lichens with different type of thalli and systematic position (collected on a single locality)
Detailed investigation of chloroplast morphology in different ecological and physiological conditions, made by laser scanning confocal microscopy.

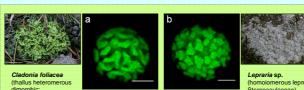
Results

- Identity of photobionts:
- · Only two species of symbiotic algae were distinguished in four lichens based on morphology and ITS sequences: Asterochloris sp. (C. foliacea, Lepraria sp.) and Trebouxia incrustata Ahmadjian ex Gärtner (L. fuscoatra, X. conspersa).

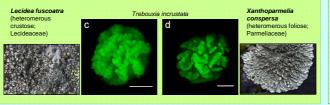
Chloroplast structure

- Confocal microscopy revealed obvious differences between chloroplast structure of algae within dessicated and hydrated thalli and between lichenized and freeliving (cultured) form of photobionts.
- During the liberation of photobiont from the lichen thallus, chloroplast changes significantly in its shape and structure. The intensity of these changes was more expressive in Asterochloris sp. compared with Trebouxia incrustata.
- The pairs of genetically identical strains show certain differences in the morphology and ontogeny of chloroplasts (Fig. 1: compare a with b, c with d).

Asterochloris sp.



One locality - four lichens - two photobionts



Effect of hydration to chloroplast structure

In a dessicated lichen thallus (herbarium specimen), chloroplast is strongly deformed (often "star-like" shaped), distinctly compressed within dehydrated protoplast.

Immediately after hydration of dry thallus, chloroplast is changing very fast to globular compact form without lobes, pyrenoid is well observable thylakoids.

Within a wet (living) lichen thallus, chloroplast occurs in several morfological stages with different degree of lobation. Pyrenoid is distinctly penetrated by thylakoids

(all pictures: Asterochloris sp. from C.

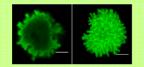
Trebouxia incrustata



Changes of chloroplast structure during the liberation of photobiont from the lichen thallus

Fresh wet lichen thali

Active photobiont cells contain compact to simple lobed chloroplast. Pyrenoid is penetrated by one or more thylakoids and surrounded by starch sheath.



7th-11th day after inoculation of thallus fragments to agar slants Strongly lobate chloroplast is formed. Algae adapted themselfs to new light and hygric conditions (= lag phase) and decreasing influence of mycobiont, which withers away due to unsuitable conditions (continual moisture within P dish). Chloroplast contains a large volume of starch (dark area around the black pyrenoid on the left picture). within Petri

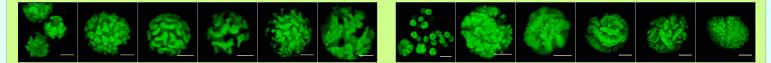


20th day

The phase of intesive division, characterized by the dominance of autospore (Trebouxia) or aplanospore (Asterochloris) packages

30th and 40th day

Mature cells of the new generation are fully liberated from mycobiont influence. Several ontogenetic stages with different morphology of chloroplast occur in the cultures, reproduction mode is changed compared with lichenized generation (Asterochloris sp. forms aplanospores and zoospores, T. incrustata autospores, aplanospores and zoospores)



Conclusions

Chloroplast morphology is dependent mainly on physiological status (dryness/dampness), degree of lichenization (lichenized/free-living) and the species level (Asterochloris sp./Trebouxia incrustata). Ontogenetical changes are obvious especially in cultured (free-living) form of photobiont.

Very closely related algae (on species as well as population level) may show number of individual differences in their behaviour. We observed distinct differences in the chloroplast structure and reproduction mode between closely related species of Asterochloris and Trebouxia. Moreover, even genetically identical strains isolated from different lichen species exhibited different morphology and ontogenetical development of chloroplast.

Confocal microscopy constitutes a convenient technique to demonstrate details in chloroplast structure of green algae.

Methods

Isolation and cultivation of the photobionts: Photobionts were isolated by the thallus fragment method (Ahmadjian, 1993). All lichens were collected from the single rocky steppe slope, at the "Máslovická stráň" protected area in Central Bohemia, Czech Republic. Algae were cultivated on agar slants of Bold's basal medium modified by adding more nitrogen (BBM 3N) at 18 °C, under an illumination of 20-30 mmol m 2 s⁻¹ and 16:8 h lightcark cycle.

to on migricular cycle: Microscopical observation: The pure algal samples and fragments of lichen thalli were investigated by laser scanning confocal microscope Leica TCS SP2 equipped with an Argon-Krypton laser using the 488 nm metodoptical observations: me port and/a data to integration to one of the control index interception of the integration of the control of th alli (fresh thalli incubated 3 davs Molecular methodology: ITS rDNA sequences were obtained from four samples. Homology between sequences and those currently available in the NCBI database was assessed using BLAST (National Center for Biotechnology Information, NIH, http://www.ncbi.nlm.nih.gov/BLAST) score system.

The study was supported by the grants 136/2006/B-BIO/PrF of the Grant Agency of the Charles University in Prague and VaVSM/ 2990/05 of the Ministry of Environment of the Czech Pomblio

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Scale bar in all pictures indicate 5 µm.

