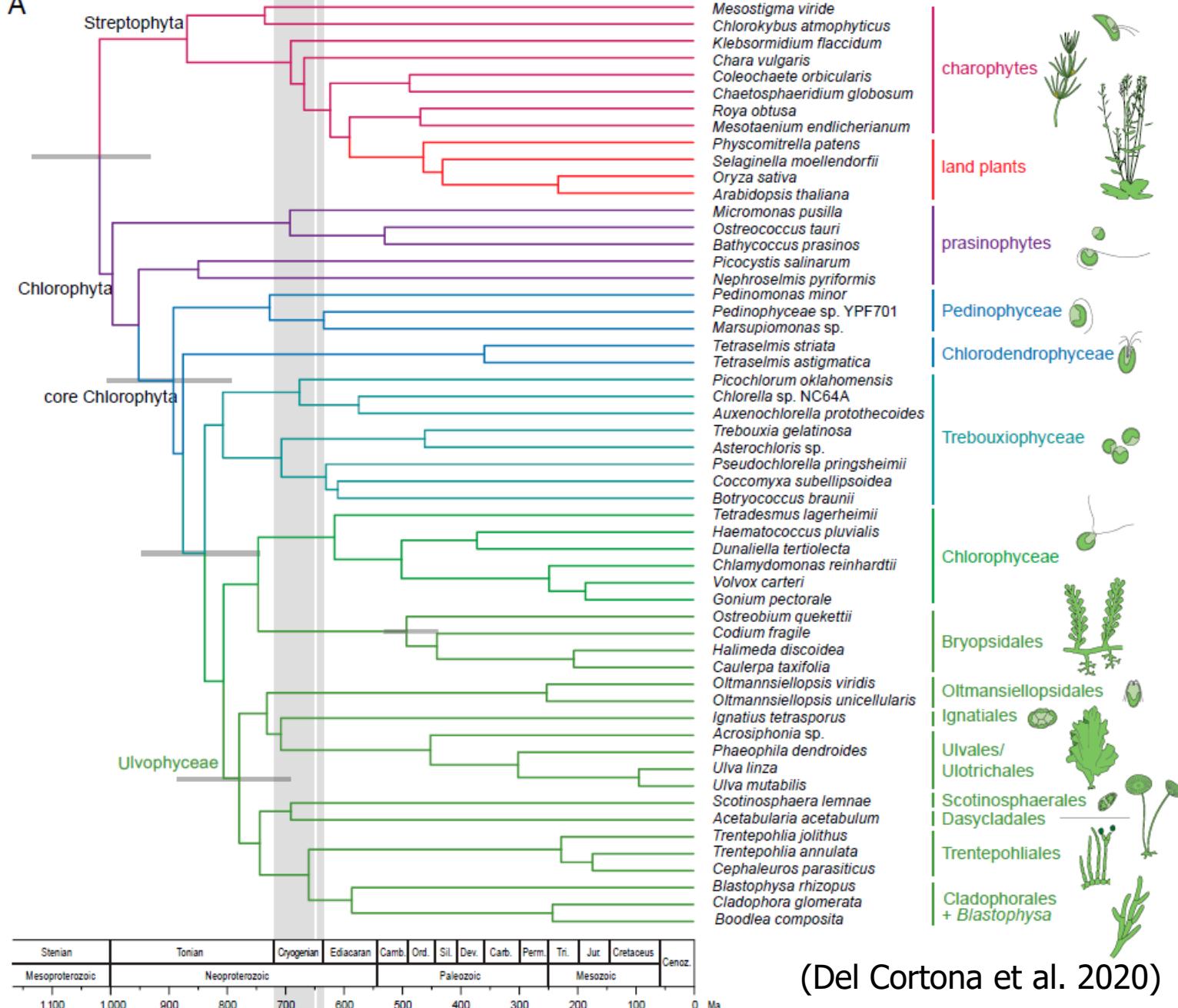


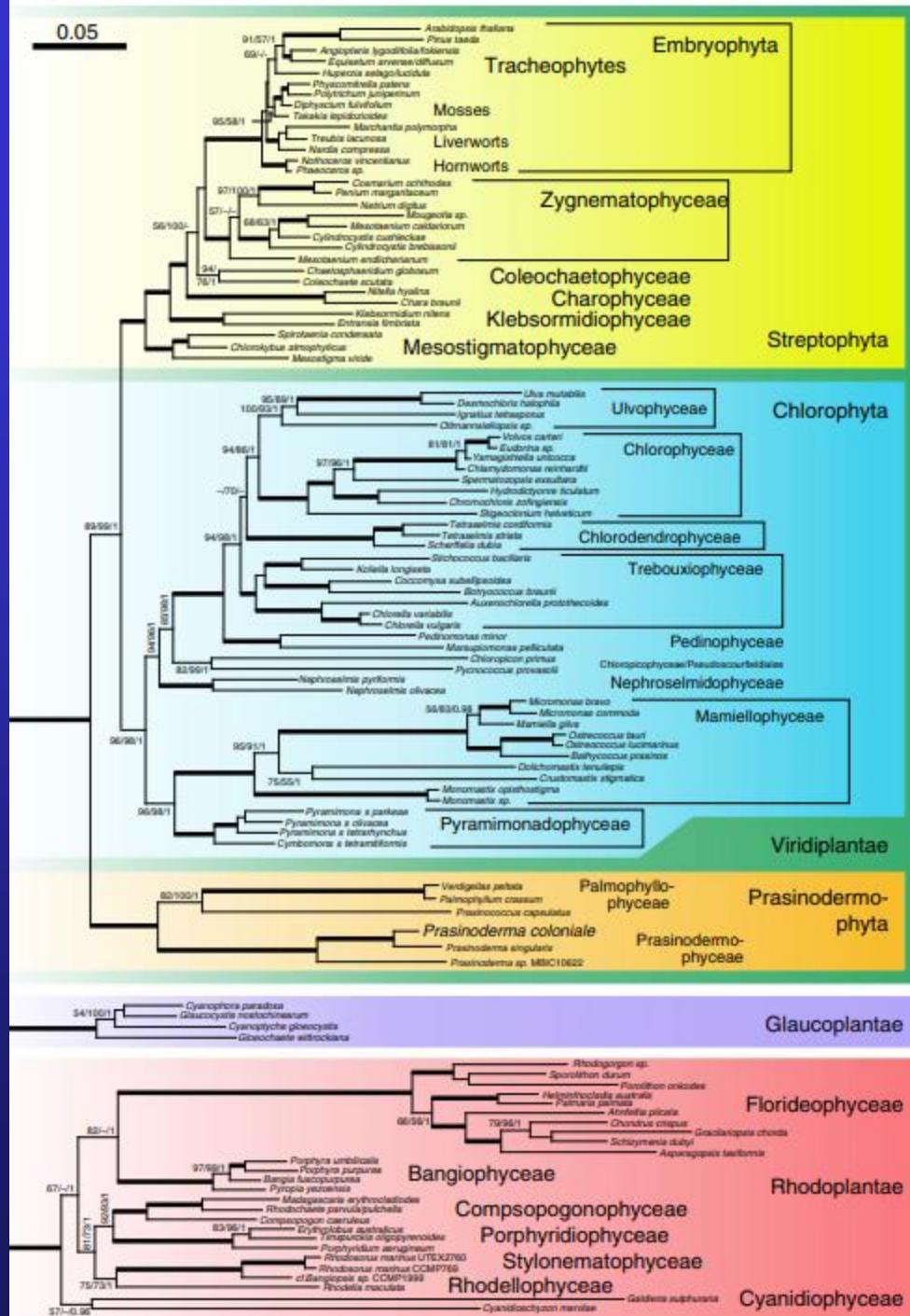
Chlorophyta

Prasinophyta

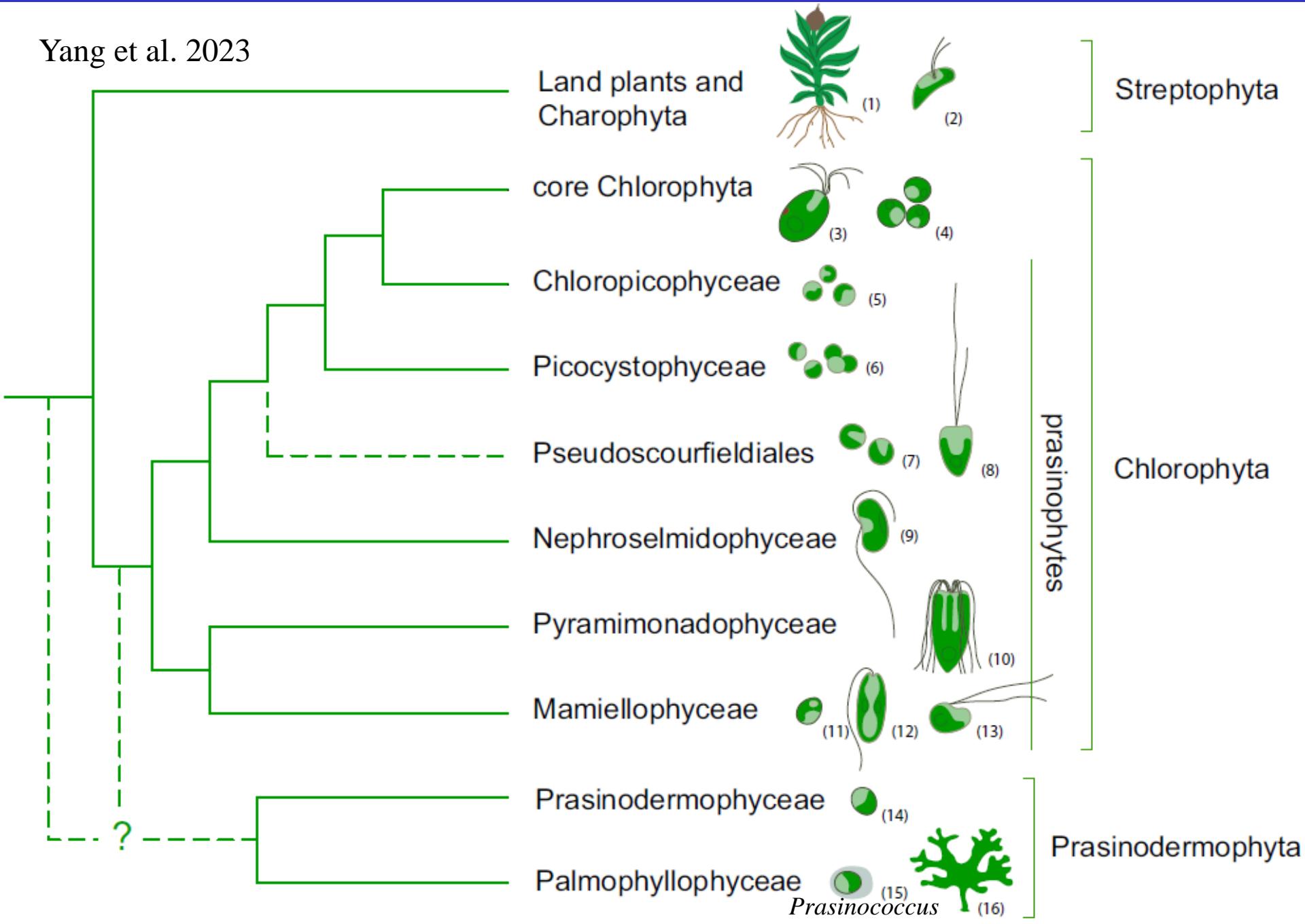
Ulvophyceae

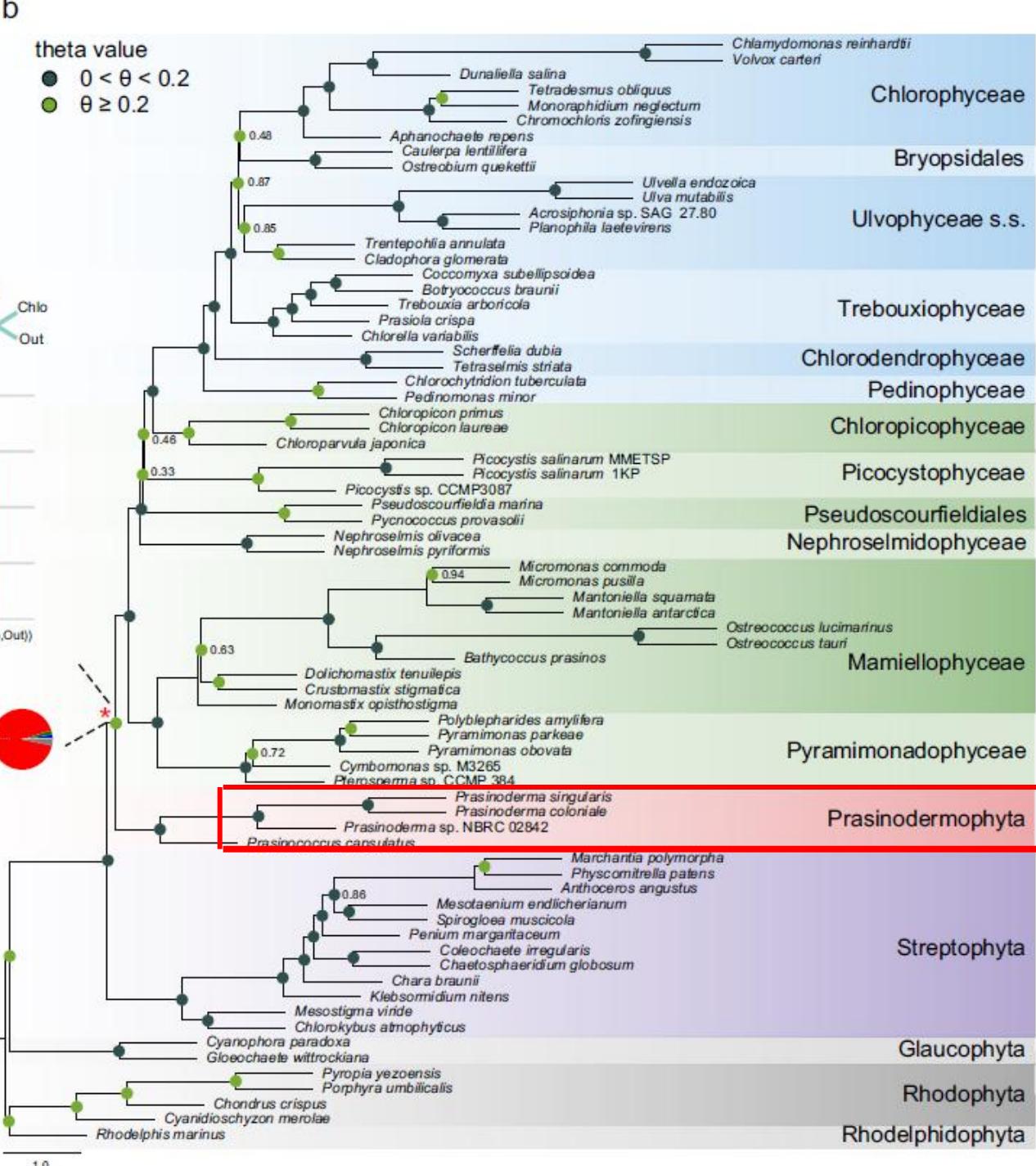


Time-calibrated phylogeny of the green algae. (A) The topology of the tree is based on the ML analysis inferred from a concatenated amino acid alignment of 539 nuclear genes



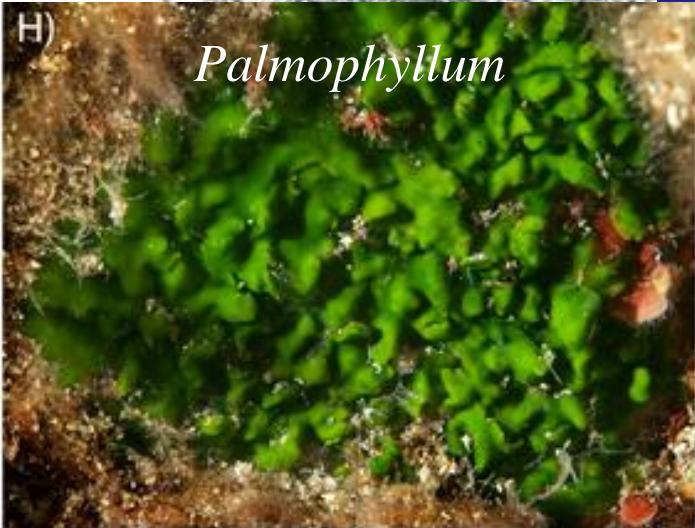
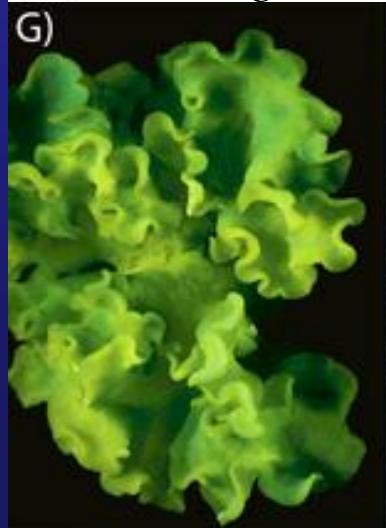
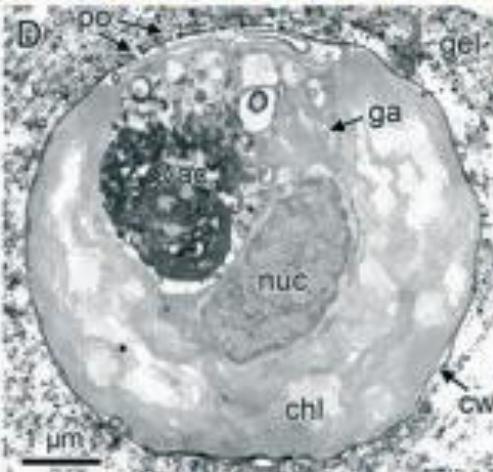
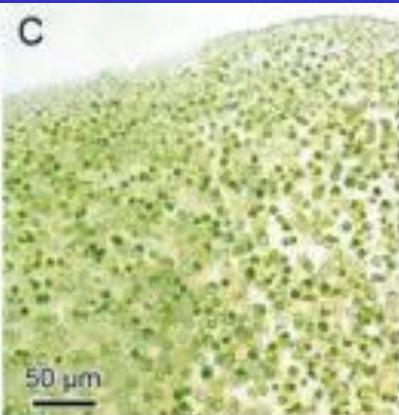
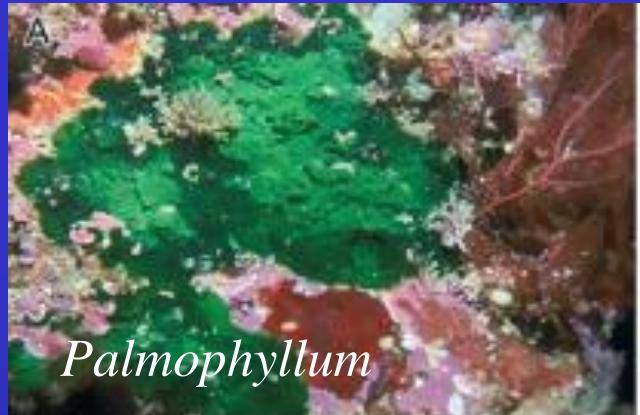
Li et al. 2020
<https://doi.org/10.1038/s41559-020-1221-7>





Yang et al. 2023

<https://doi.org/10.1038/s41467-023-41137-5>





Palmophyllum crassum –
Mediterranean Sea

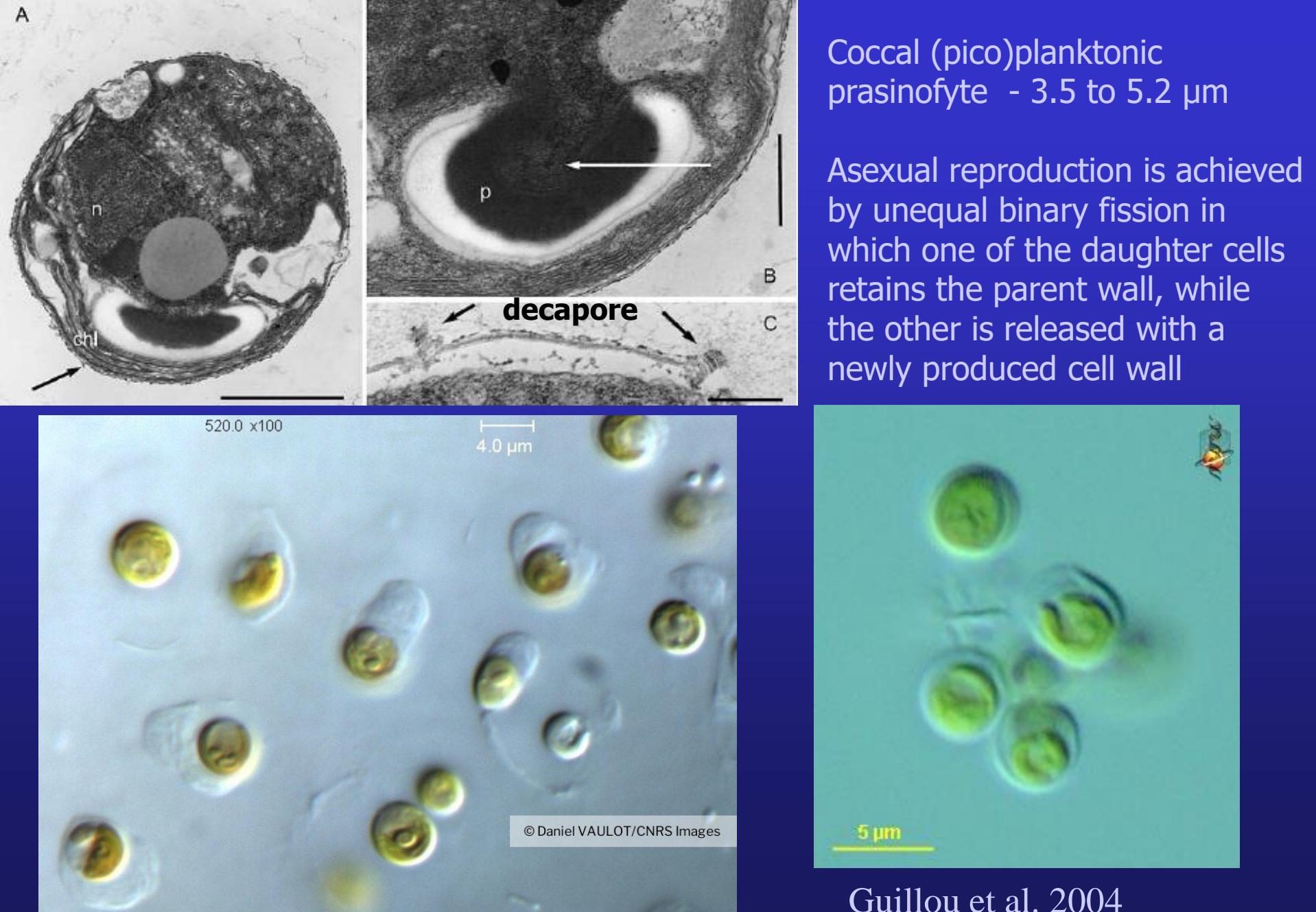
higher ch b/ch a ratio
no siphonaxanthin and siphonein



Palmocladus stipitatus.
This rare and remarkable alga has a stem that
shows annular rings, out of which grows a
delicate, cupshaped, perforated membranous
blade. Individual plants up to 8 years of age
have been recorded
South Australia

Palmophylophyceae

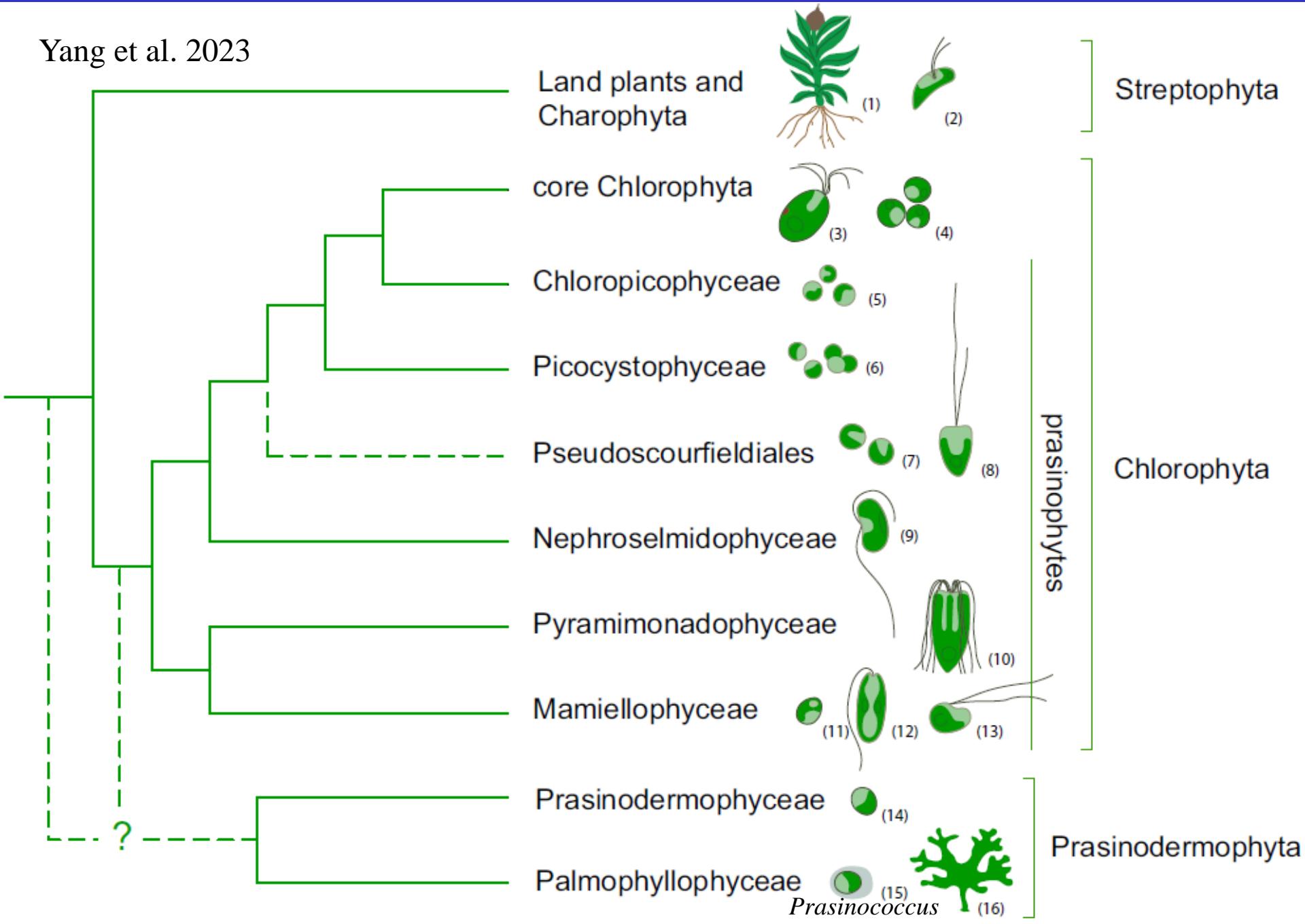
Prasinococcus cf. capsulatus

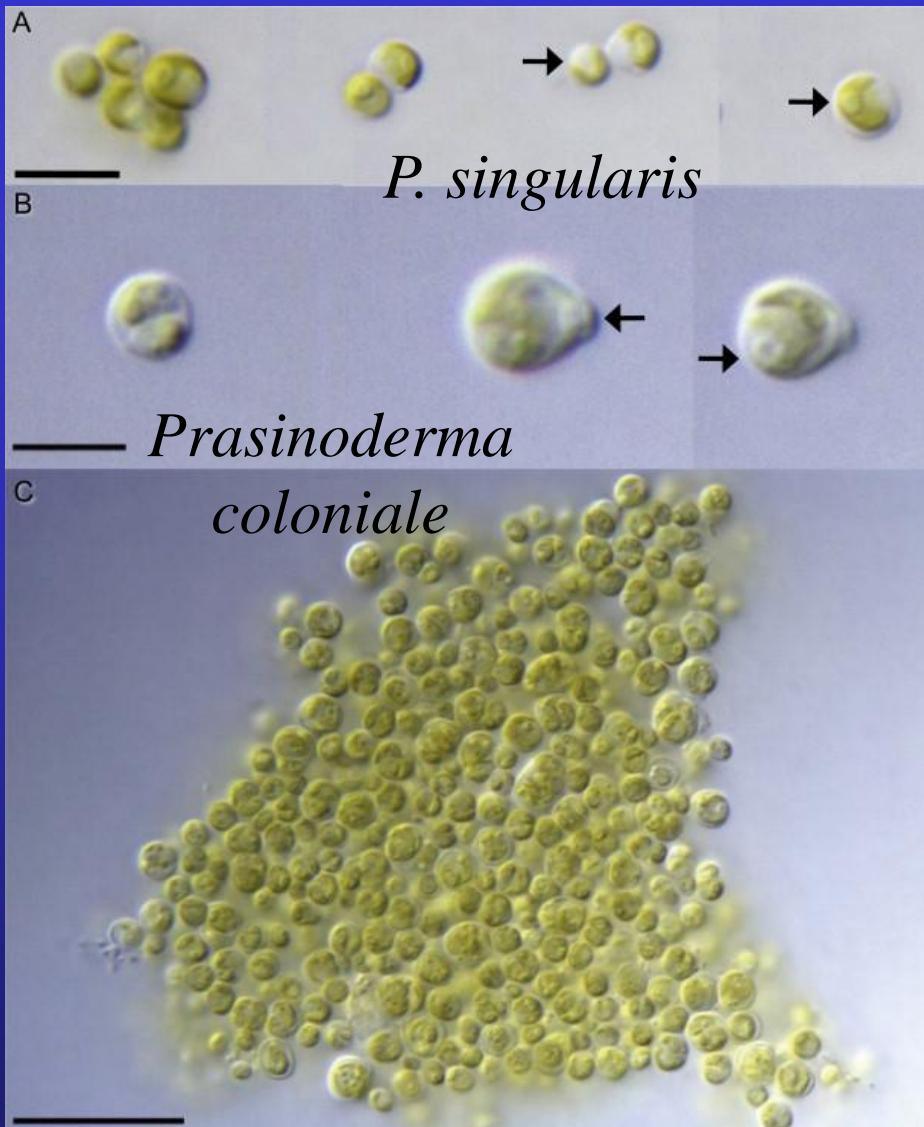


Coccal (pico)planktonic prasino phyte - 3.5 to 5.2 μ m

Asexual reproduction is achieved by unequal binary fission in which one of the daughter cells retains the parent wall, while the other is released with a newly produced cell wall

Guillou et al. 2004





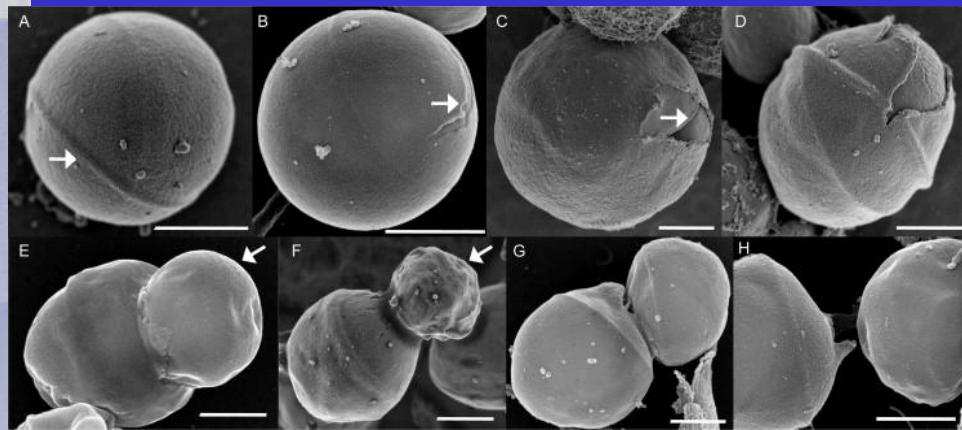
Surface water of Pacific Ocean.

Bioscope expedition 2004

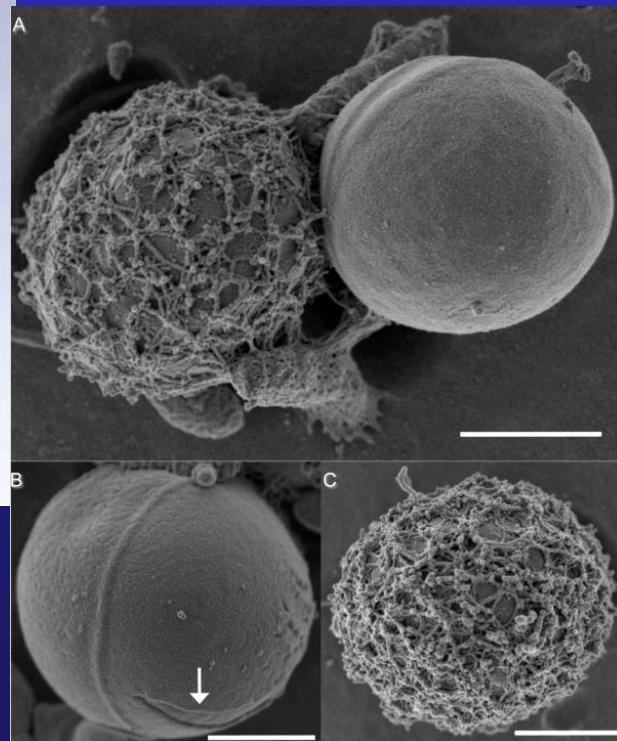
(Jouenne et al. 2011)

Prasinodermophyceae

Prasinoderma



Ch a,b
prasinoxan-
thin,
micromonol
uriolide

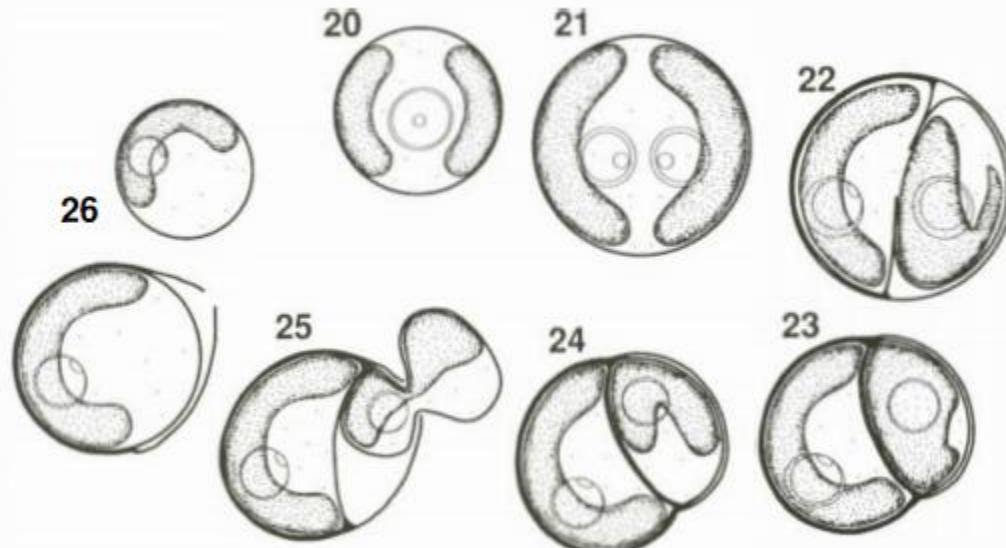


Asexual
reproduction
by unequal
binary
fission and
“budding-
like”
process

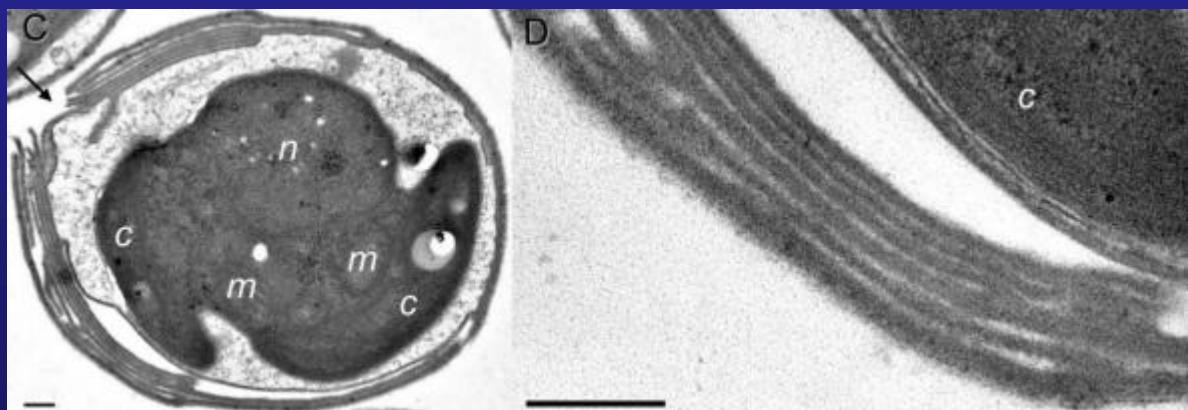
Prasinoderma coloniale

(HASEGAWA et al. 1996)

Hasegawa et al.: *Prasinoderma coloniale* gen. et sp. nov.

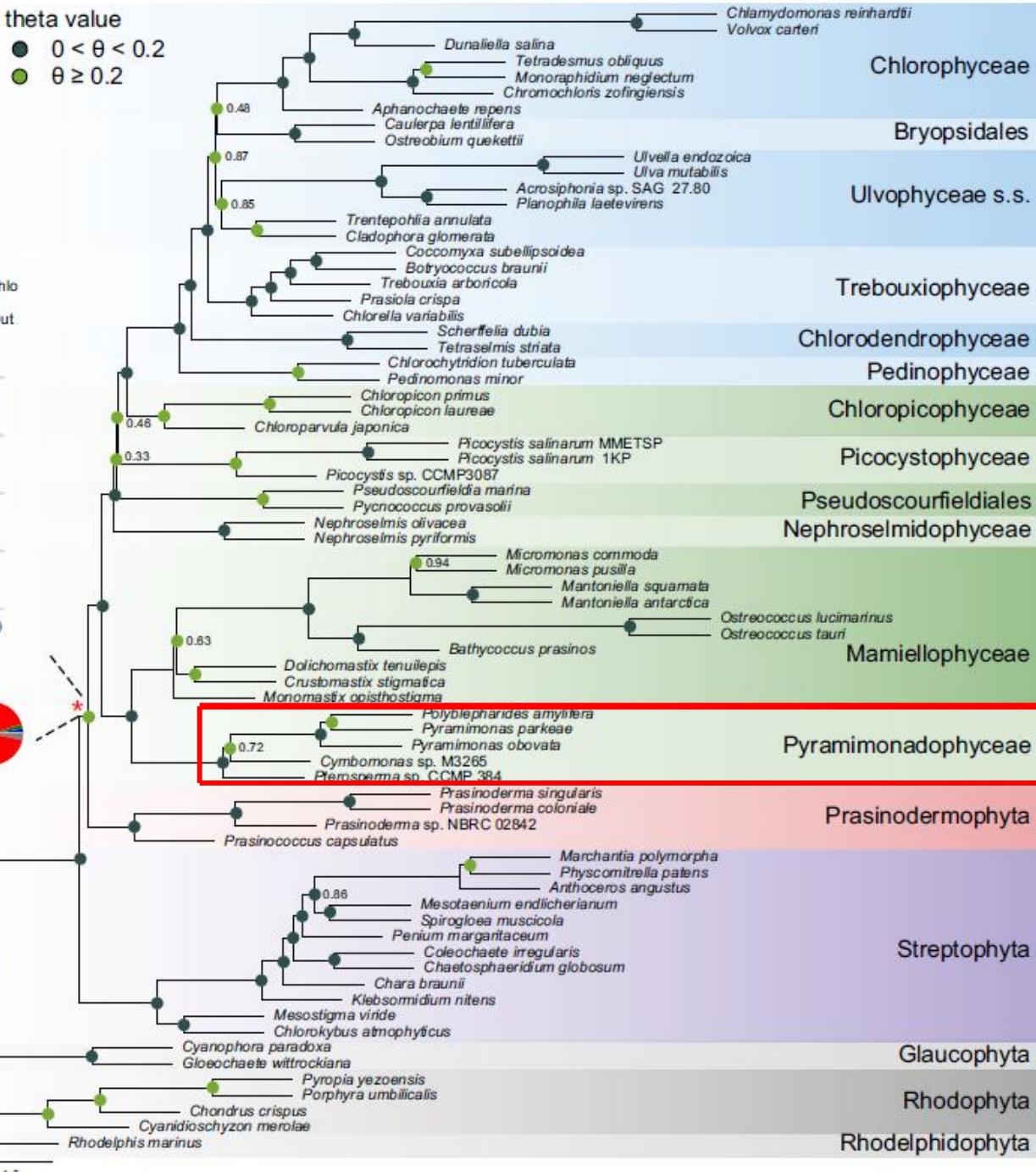


Figs 20–26. *Prasinoderma coloniale*. Schematic illustrations showing the process of asexual reproduction.

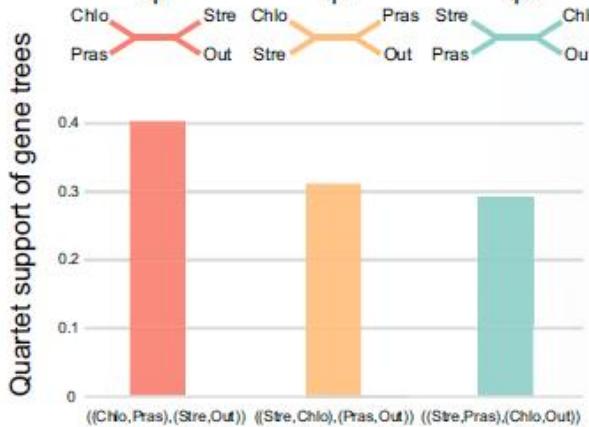


(Jouenne et al. 2011)

b



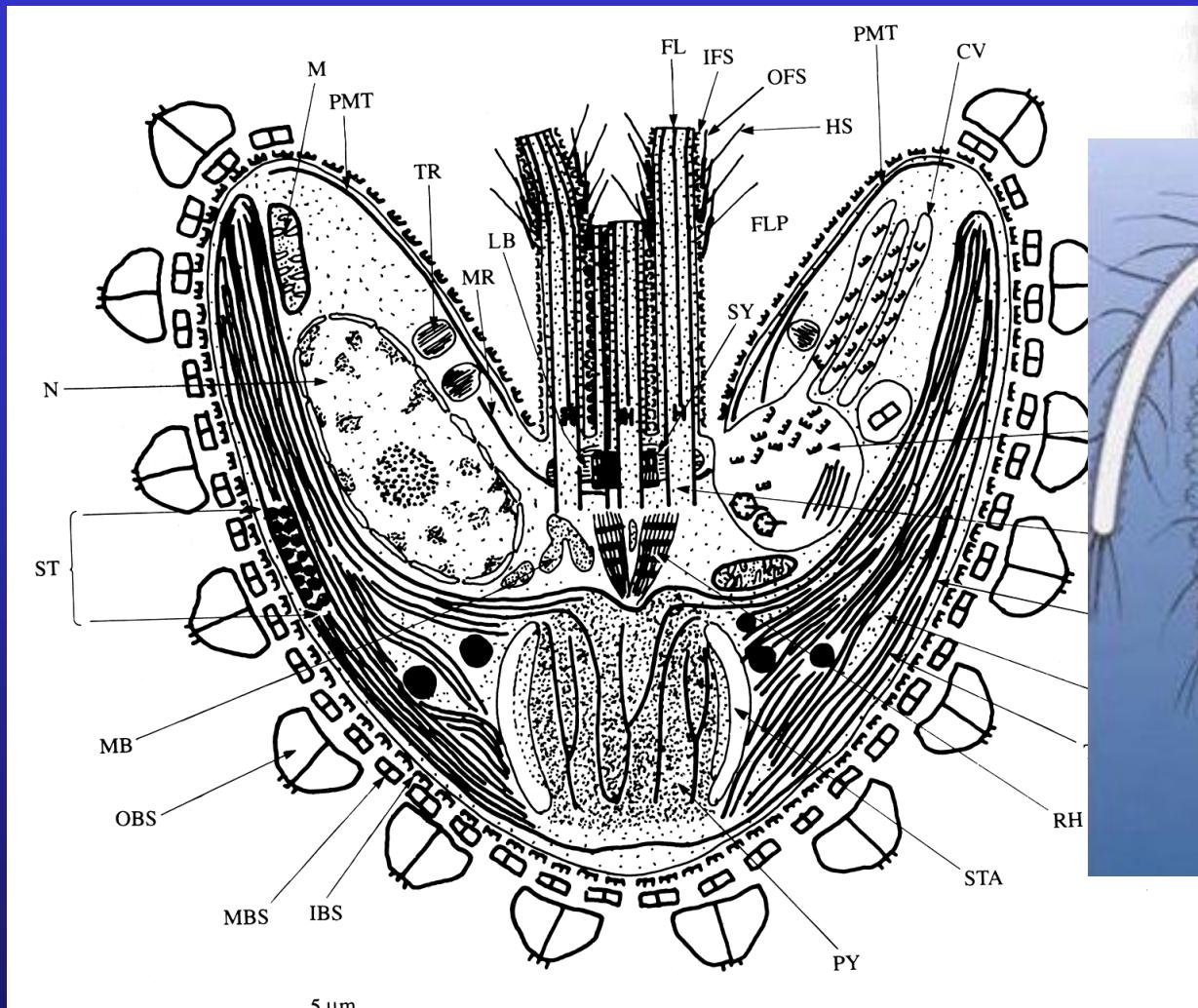
C



Yang et al. 2023

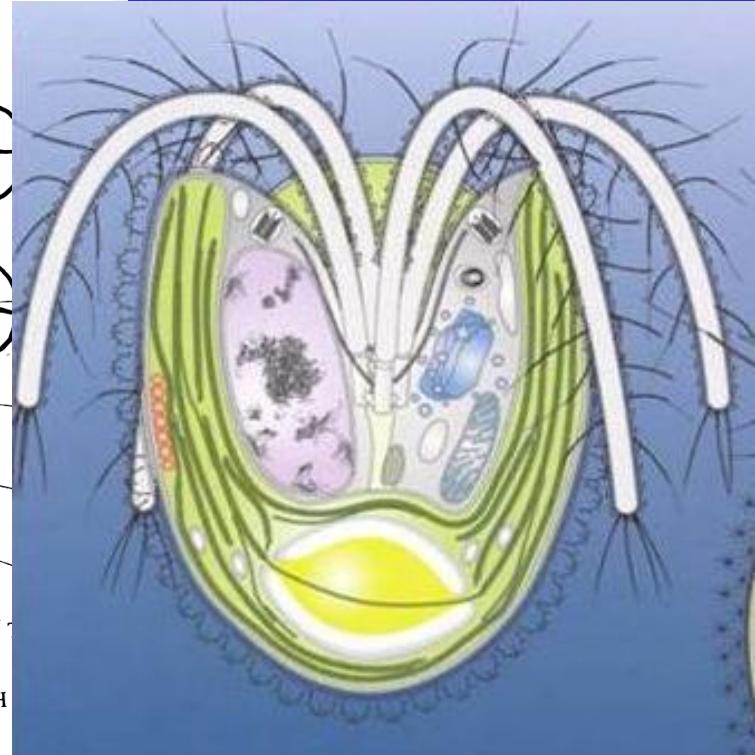
<https://doi.org/10.1038/s41467-023-41137-5>

Pyramimonadales



trichocysts- extracellular digestion

no prasinoxanthin
70 species



rhizoplast attached to the
chloroplast surface
(contraction ATP, Ca²⁺)

ultrastructure of *Pyramimonas*

Pyramimonadales

Cymbomonas

chloroplast

mitochondria

nucleus

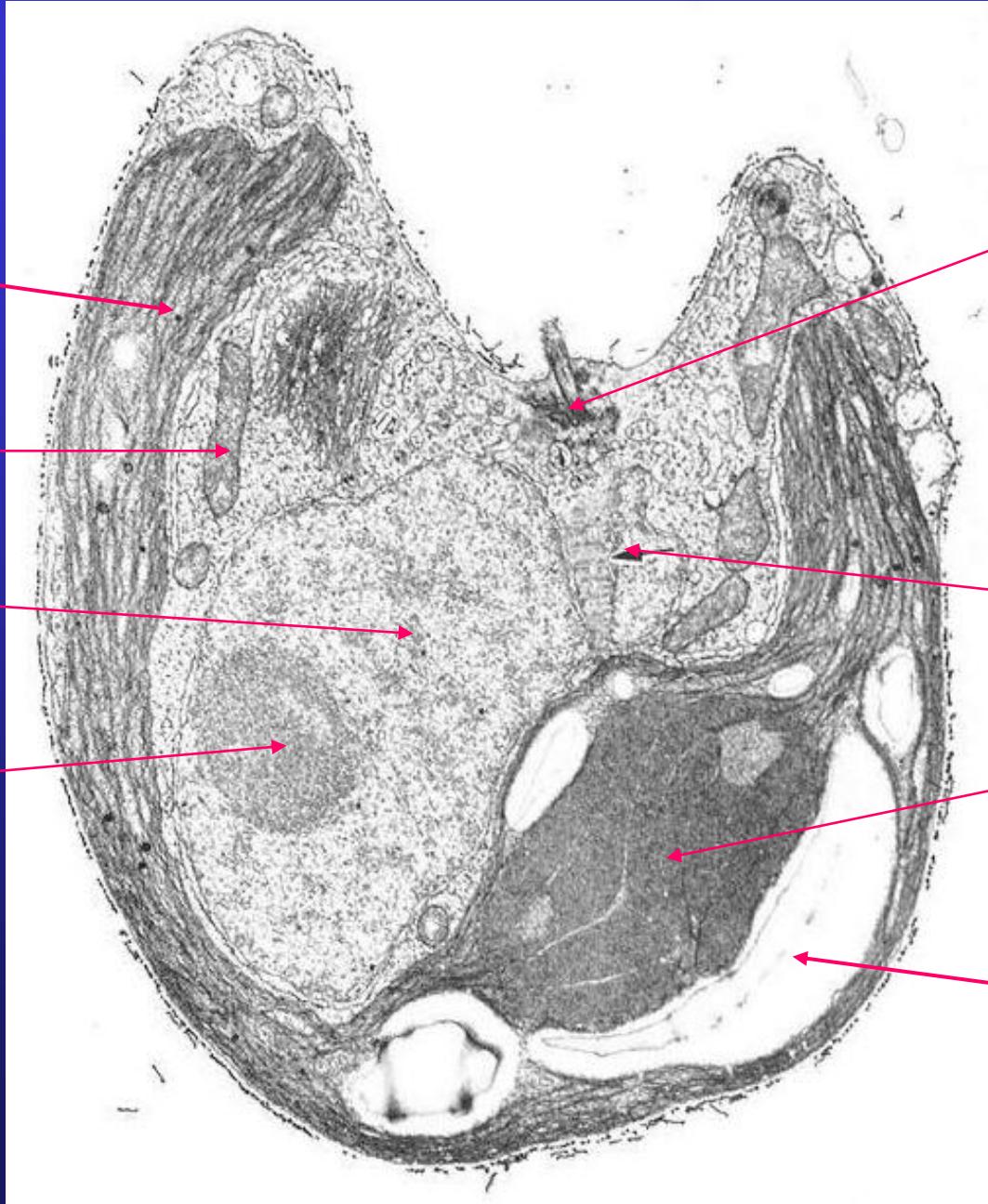
nucleolus

basal bodies

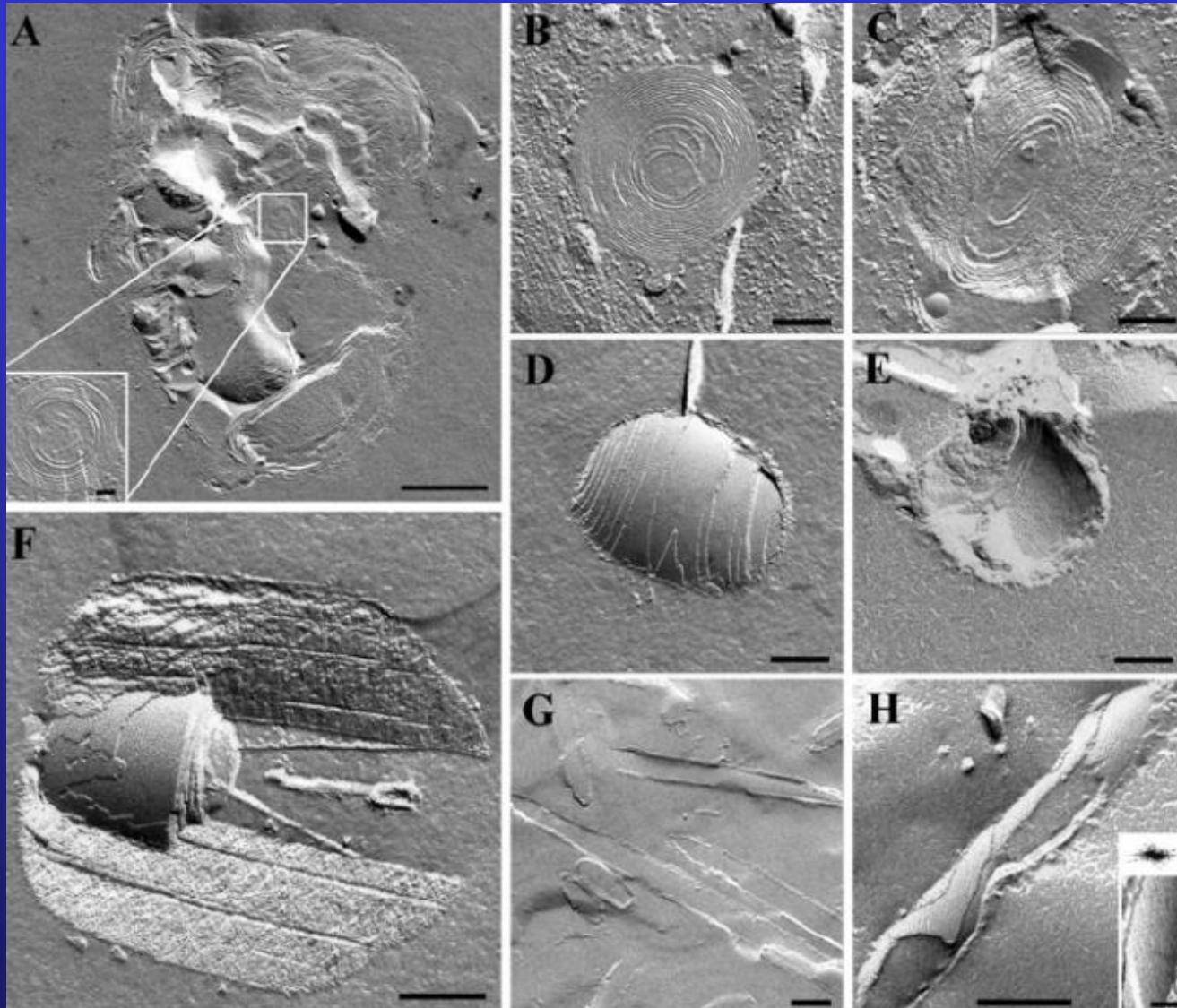
rhizoplast

pyrenoid

starch grain



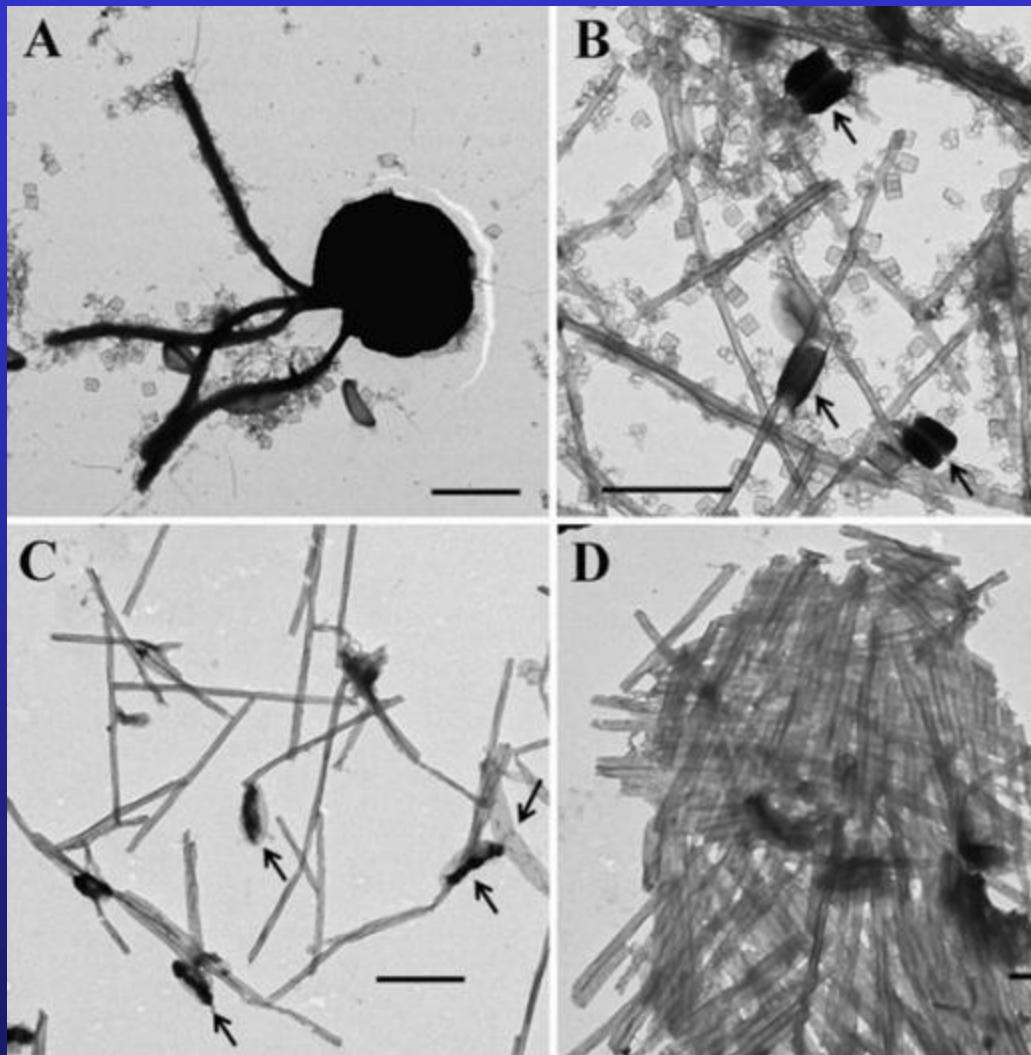
Ejectisomes



Discharged
ejectisomes
measured up to 26
 μm
still furled
measured up to
900 nm in width
and 1 μm in length

Rhiel et al.
Protoplasma 2013

Micrographs of freeze-fractured ejectisomes of *P. grossii*. In freeze-fractured *Pyramimonas* cells

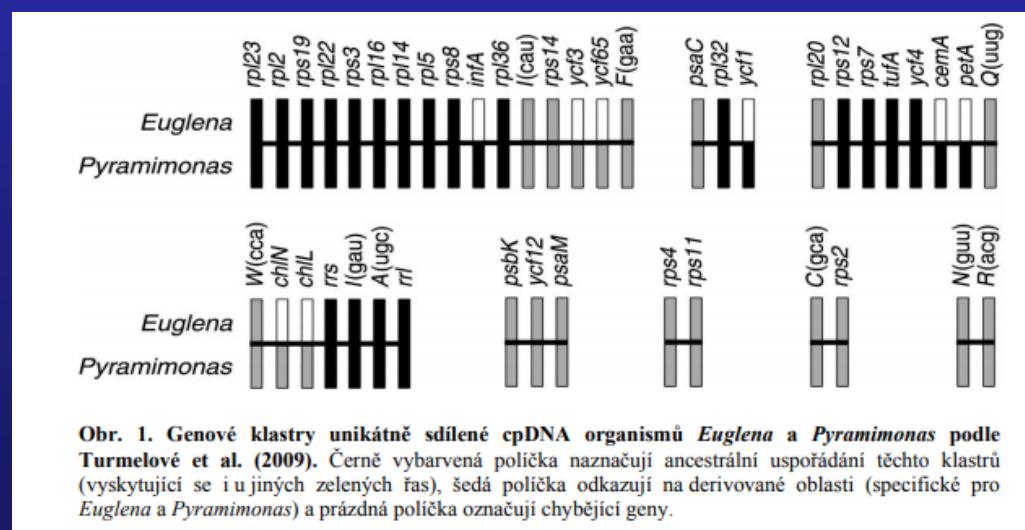


Micrographs of a cell of *P. grossii* (a) and of crude (b) and enriched (c, d) ejectisome fractions after negative staining.



Pyramimonas parkeae - the closest relative to the euglena chloroplast

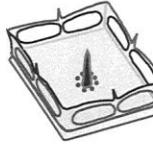
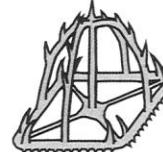
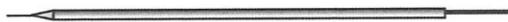
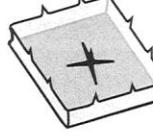
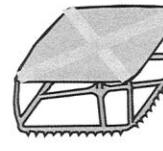
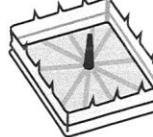
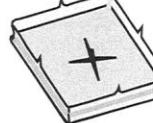
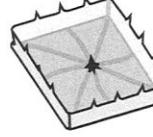
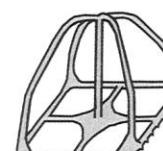
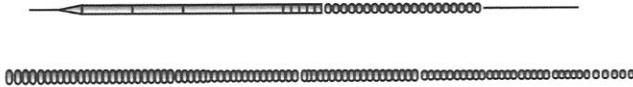
Plastid genomes share unique
gene clusters



Obr. 1. Genové klastry unikátně sdílené cpDNA organismů *Euglena* a *Pyramimonas* podle Turmelové et al. (2009). Černé vybarvená polička naznačují ancestrální uspořádání těchto klastrů (vyskytující se i u jiných zelených řas), šedá polička odkazují na derivované oblasti (specifické pro *Euglena* a *Pyramimonas*) a prázdná polička označují chybějící geny.

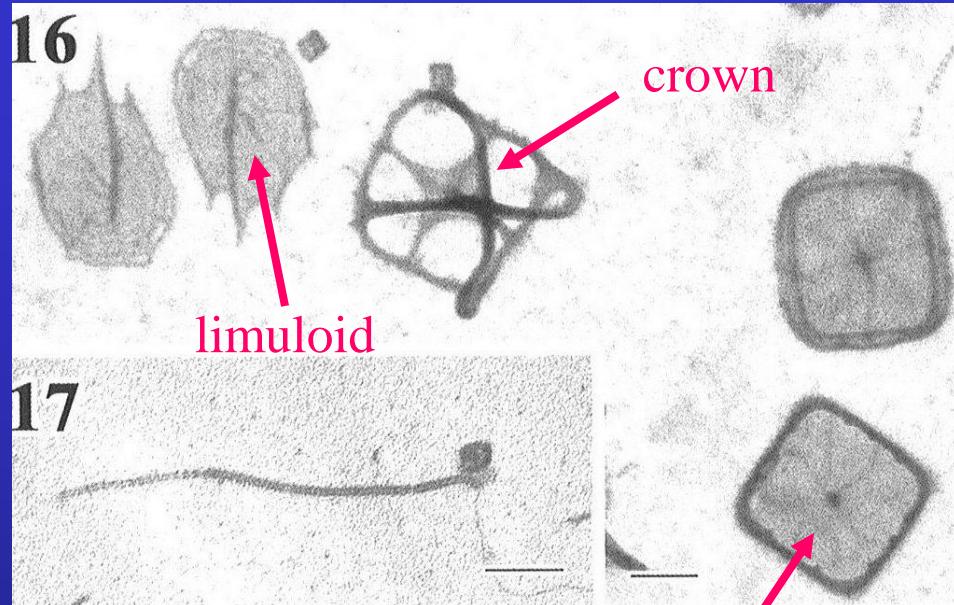
Organic scales

Table 3. Comparison of the body (box and crown) scales and the flagellar (limuloid and hair) scales among species of *Pyramimonas* subgenus *Punctatae*.

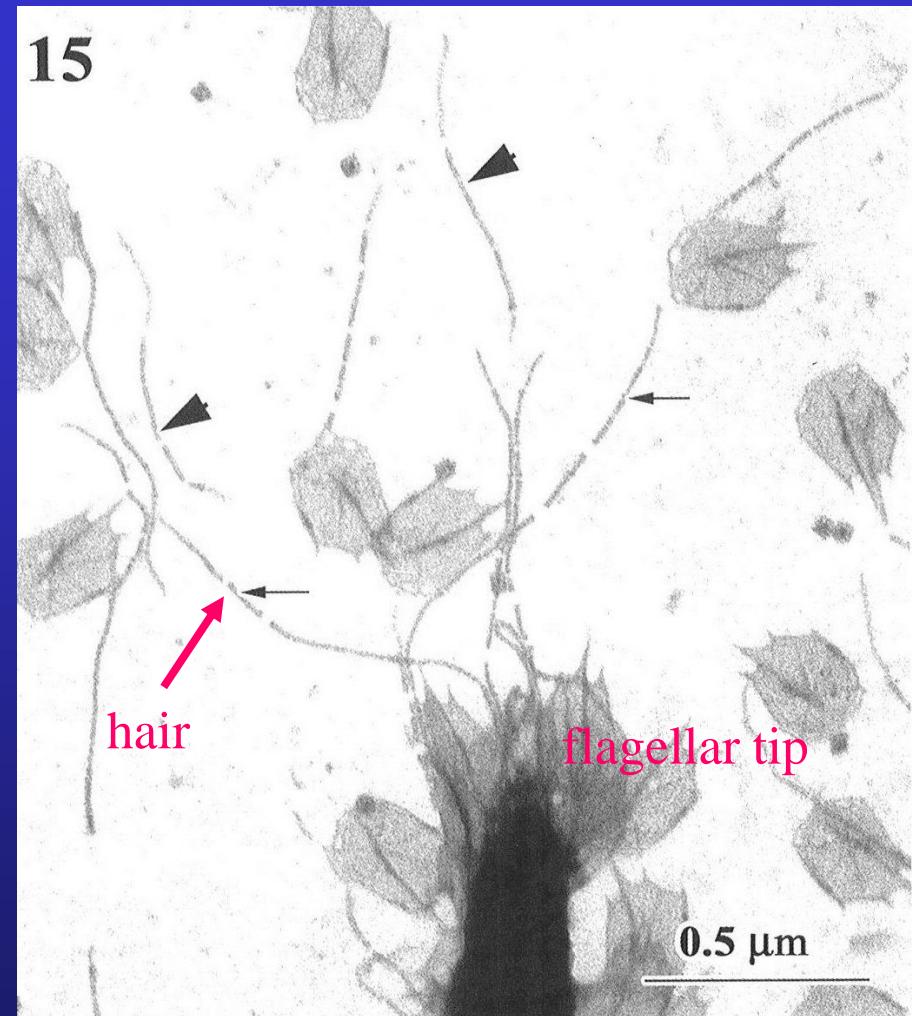
species	body scales		flagellar scales	
	box scale	crown scale	limuloid scale	hair scale
<i>P. formosa</i> ¹				
<i>P. mucifera</i> ²				
<i>P. olivacea</i> ³				
<i>P. robusta</i> ⁴				
<i>P. aurea</i> ⁵				

¹Sym & Pienaar 1999; ²Sym & Pienaar 1991, Pienaar & Sym 1997; ³McFadden *et al.* 1987, Pienaar & Sym 1997; ⁴Pienaar & Sym 1997; ⁵this study

Organic scales



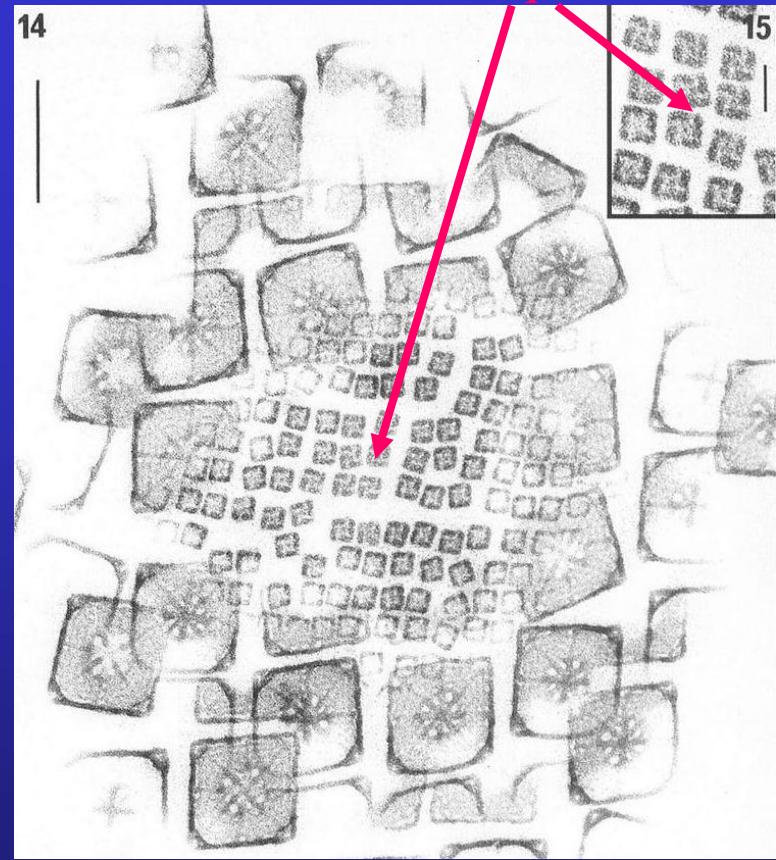
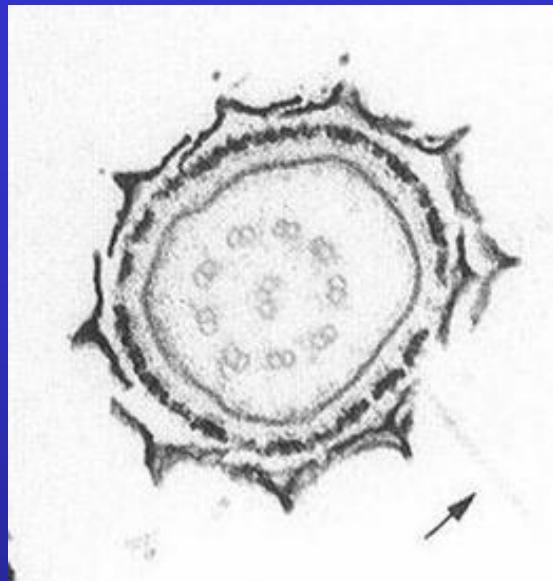
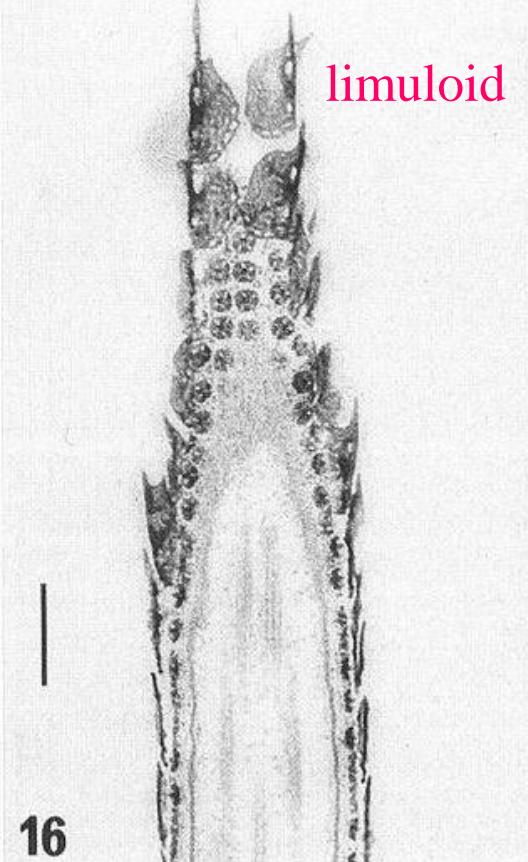
Pyramimonas aurea



carbohydrate similar to pectin

Organic scales

limuloid



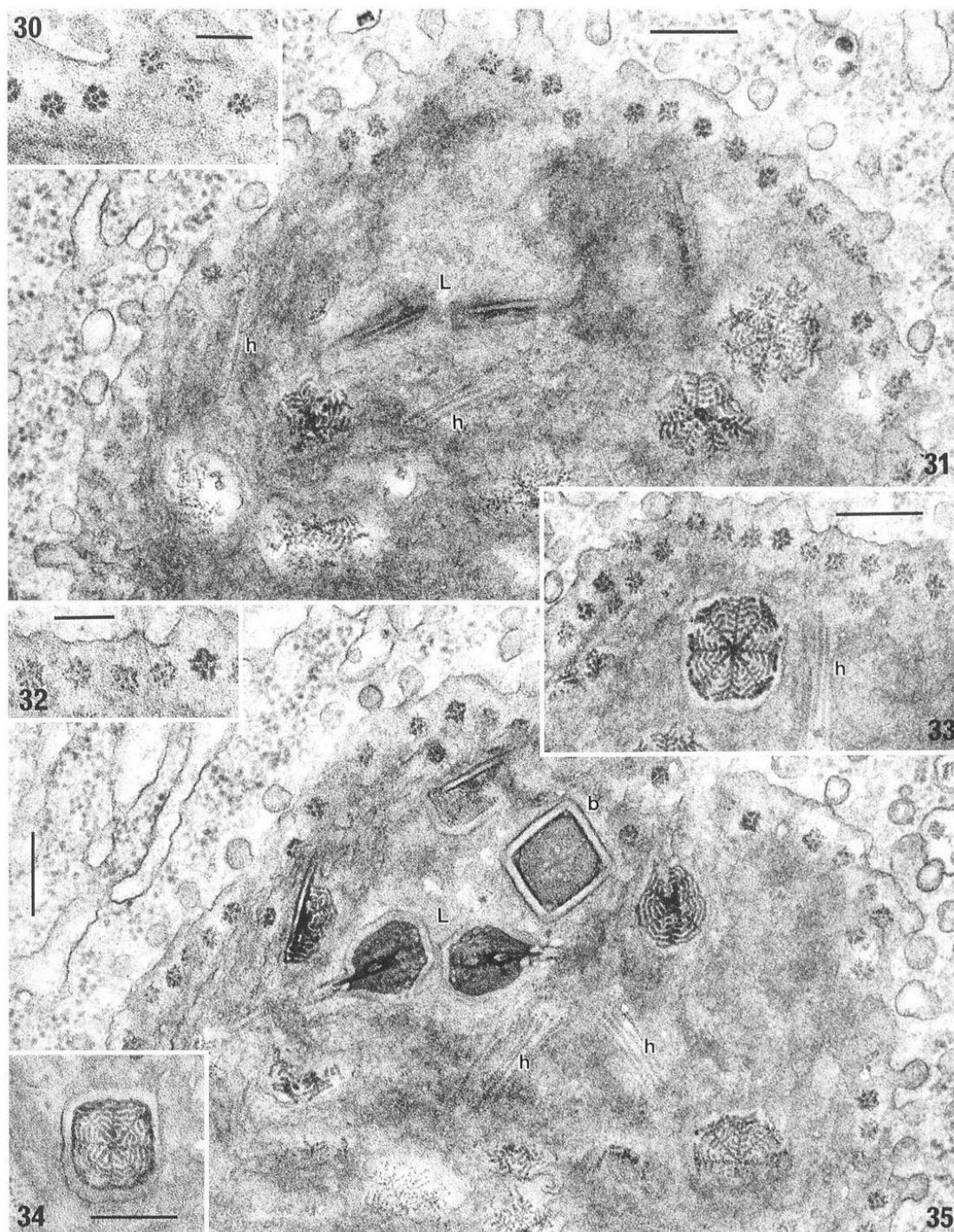
*Cymbomonas
tetramitiformis*

box

Scales produced by GA
– all types of body and flagellar scales can be produced in a single GA cisterna, secreted to the surface in the area of flagellar insertion (flagellar pit).

x chrysophyte silica scales

Cymbomonas

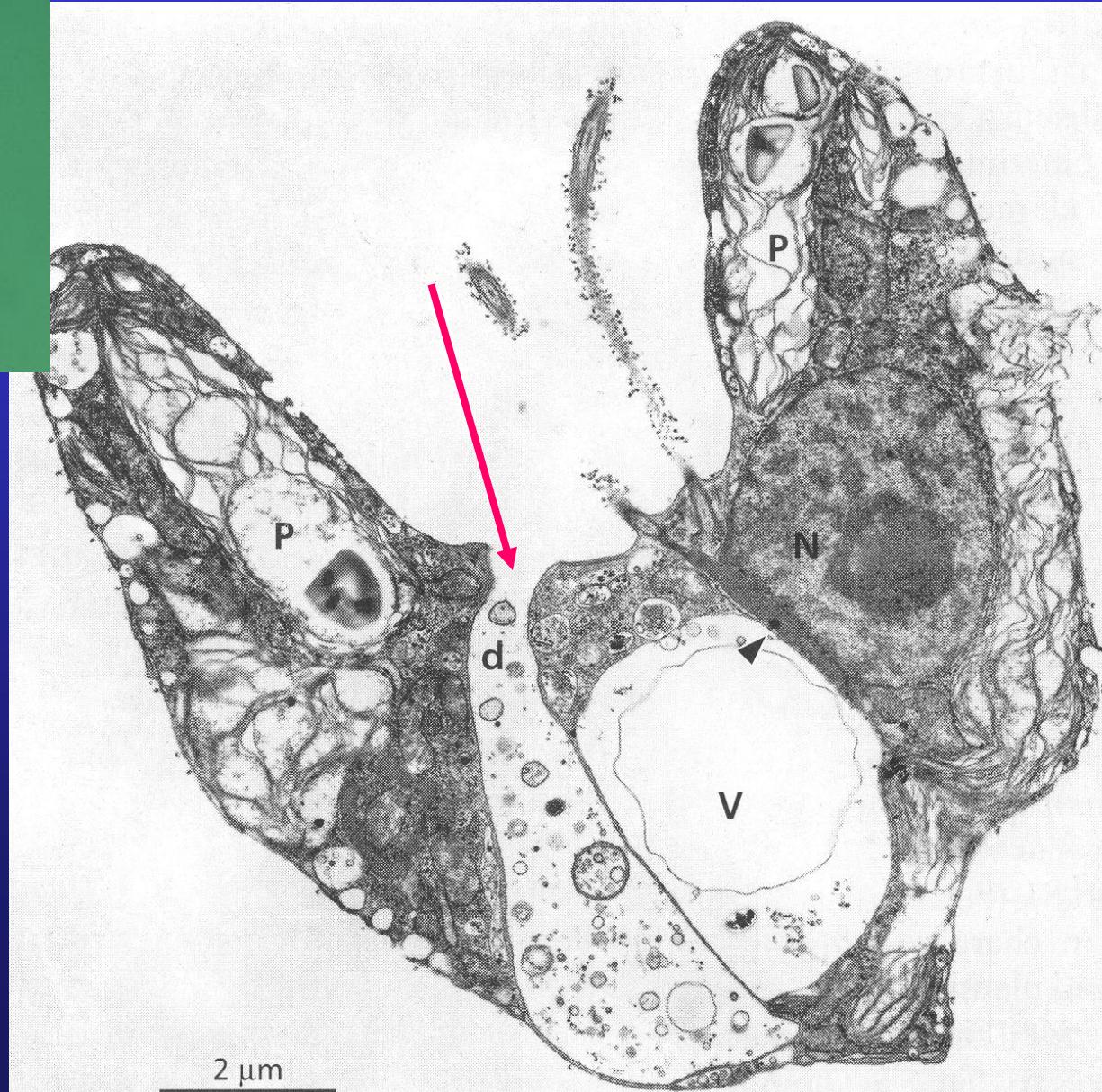


Organic scales

Halosphaera minor
Ostenfeld

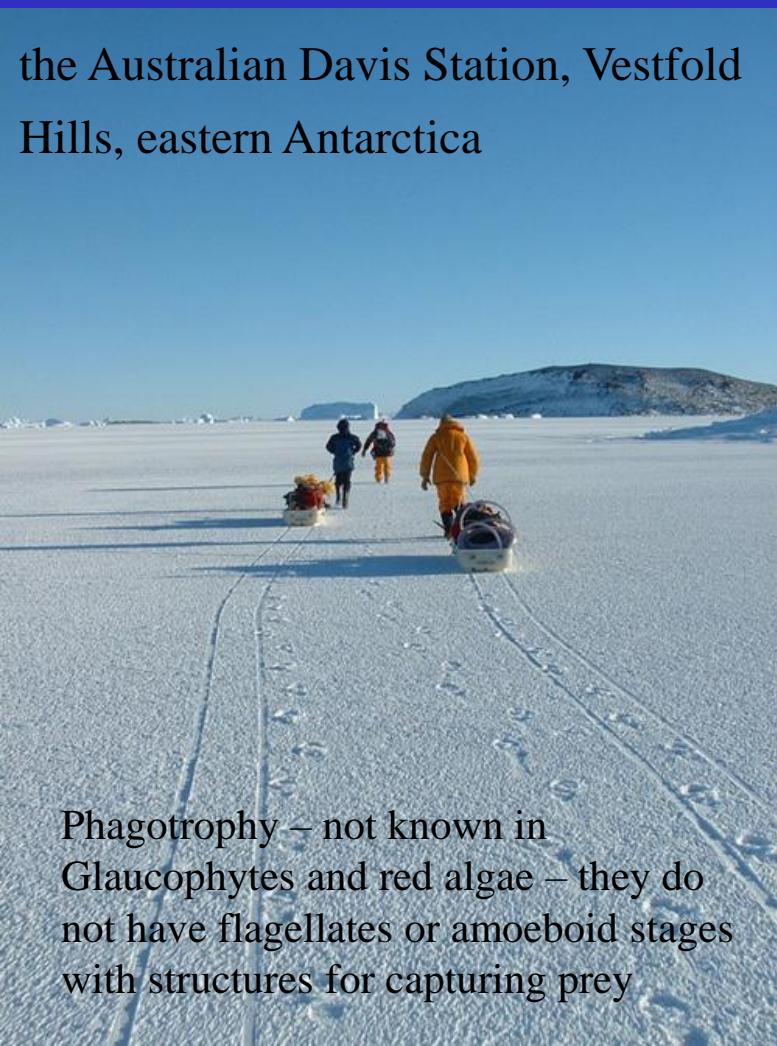


Halosphaera –
specialized channel
through which scales
are released, vestigial
cytopharynx of
ancestral phagotrophic
cells



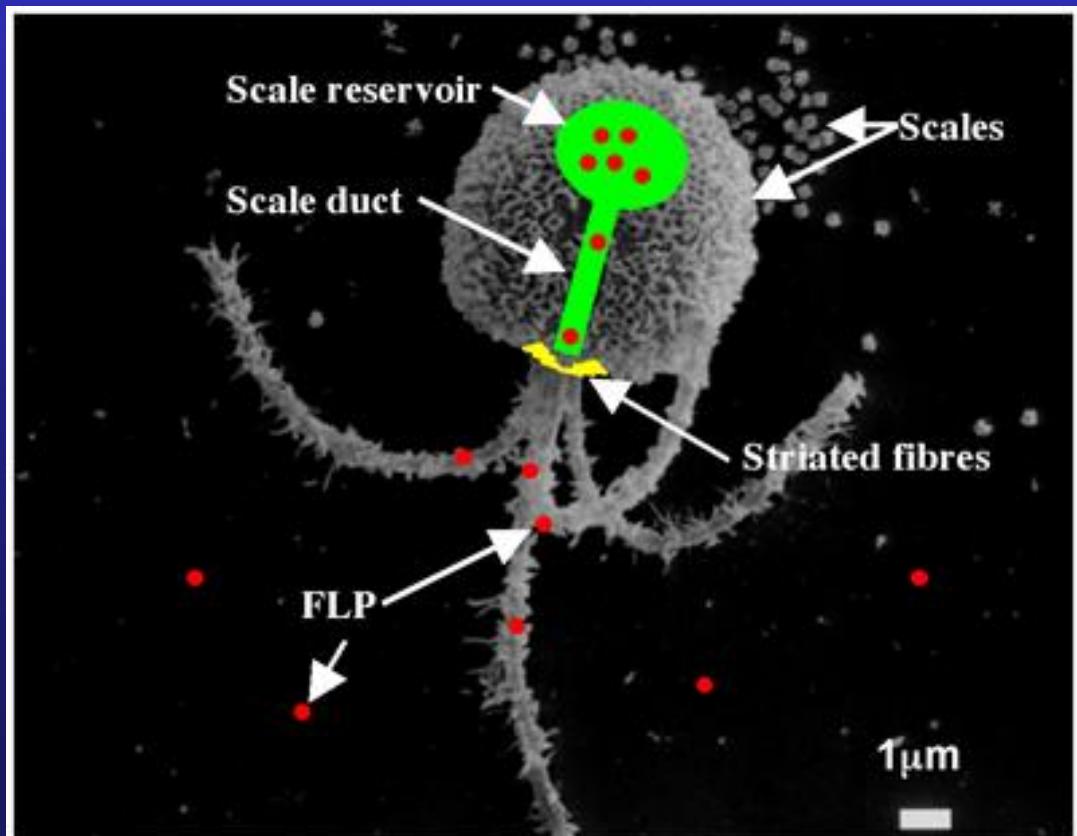
Mixotrophy 'activated' only under extreme conditions (total winter darkness and again when ice thickness was maximal and PAR levels in the water column low) – trophic plasticity

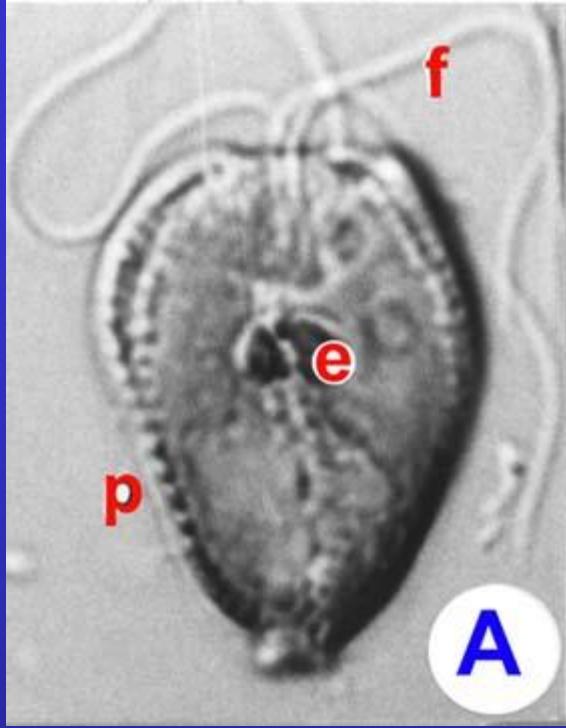
the Australian Davis Station, Vestfold Hills, eastern Antarctica



Phagotrophy – not known in Glaucophytes and red algae – they do not have flagellates or amoeboid stages with structures for capturing prey

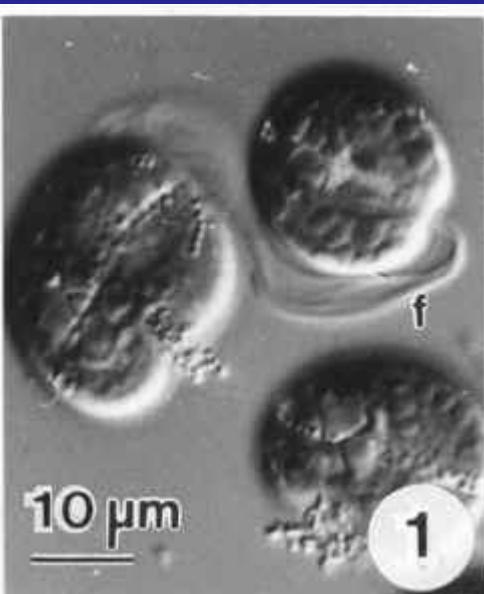
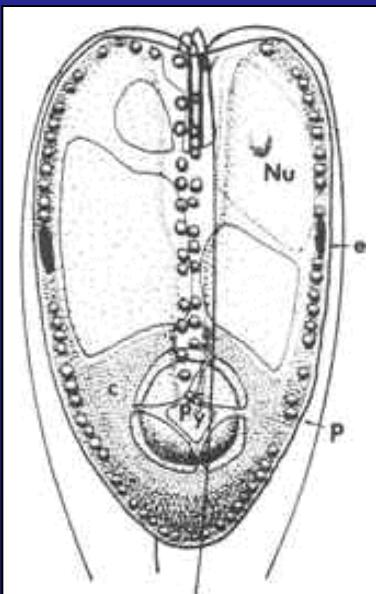
Diagrammatic representation of *Pyramimonas gelidicola* ultrastructure illustrating scale duct and reservoir thought to be involved in prey uptake. FLP, fluorescently labelled prey.



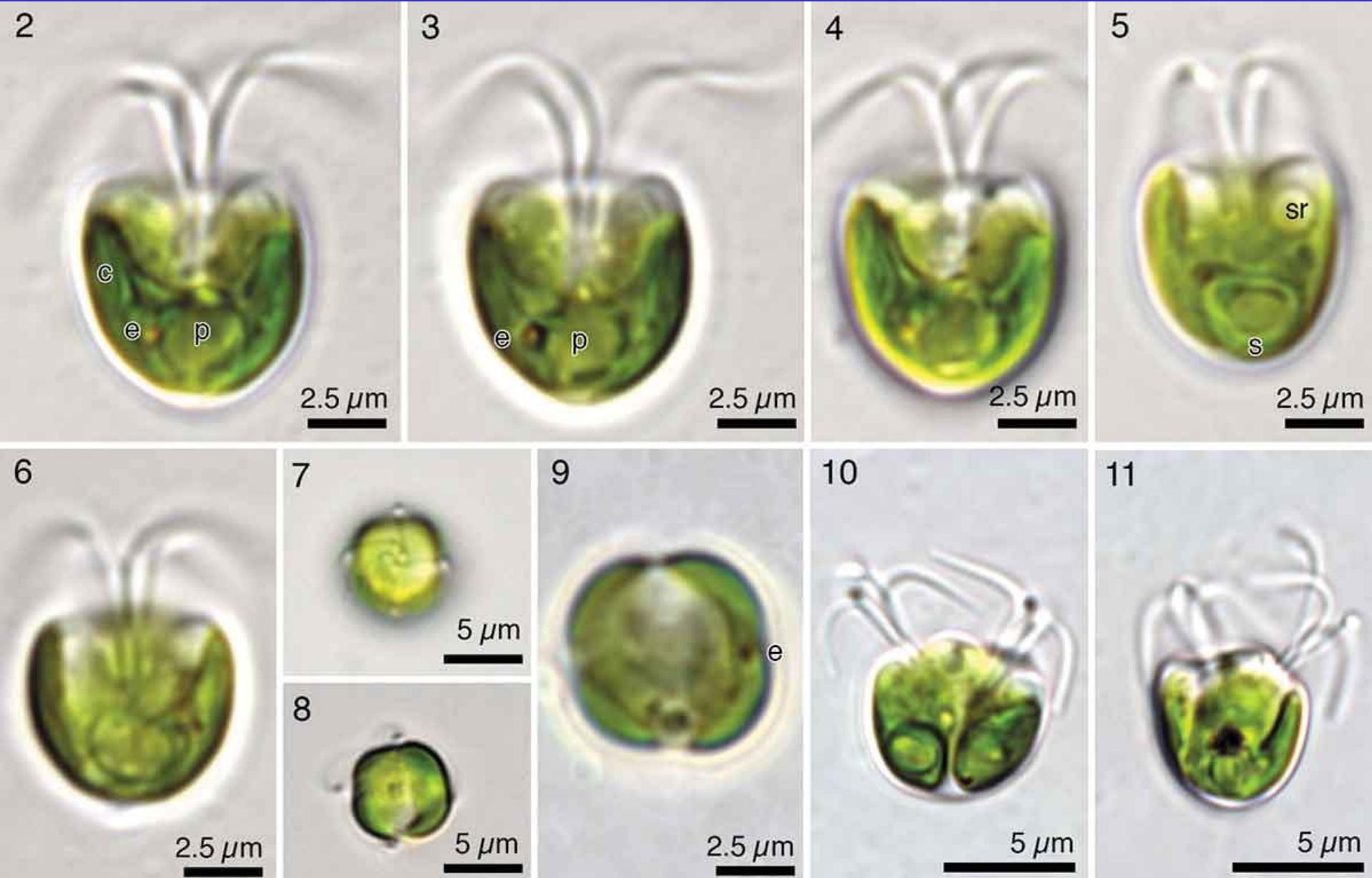


Pyramimonas mucifera

benthic species,
produces slime –
adaptation to benthic
environment



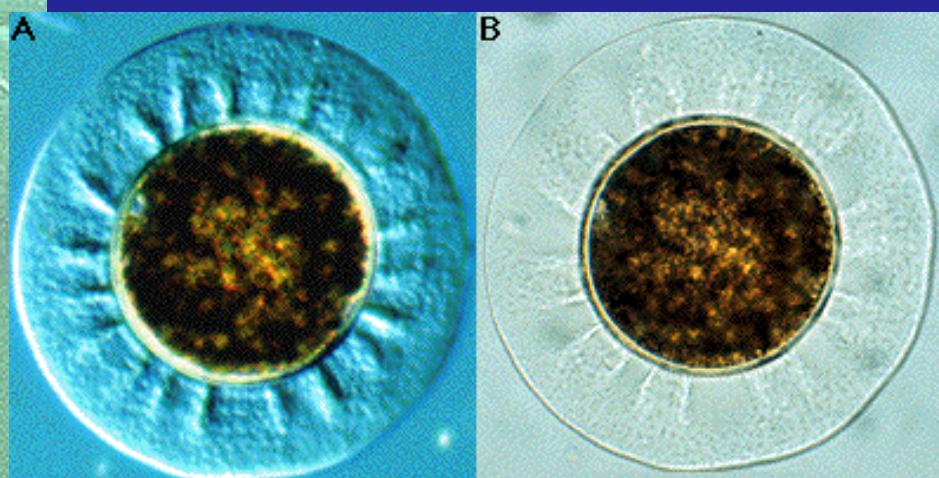
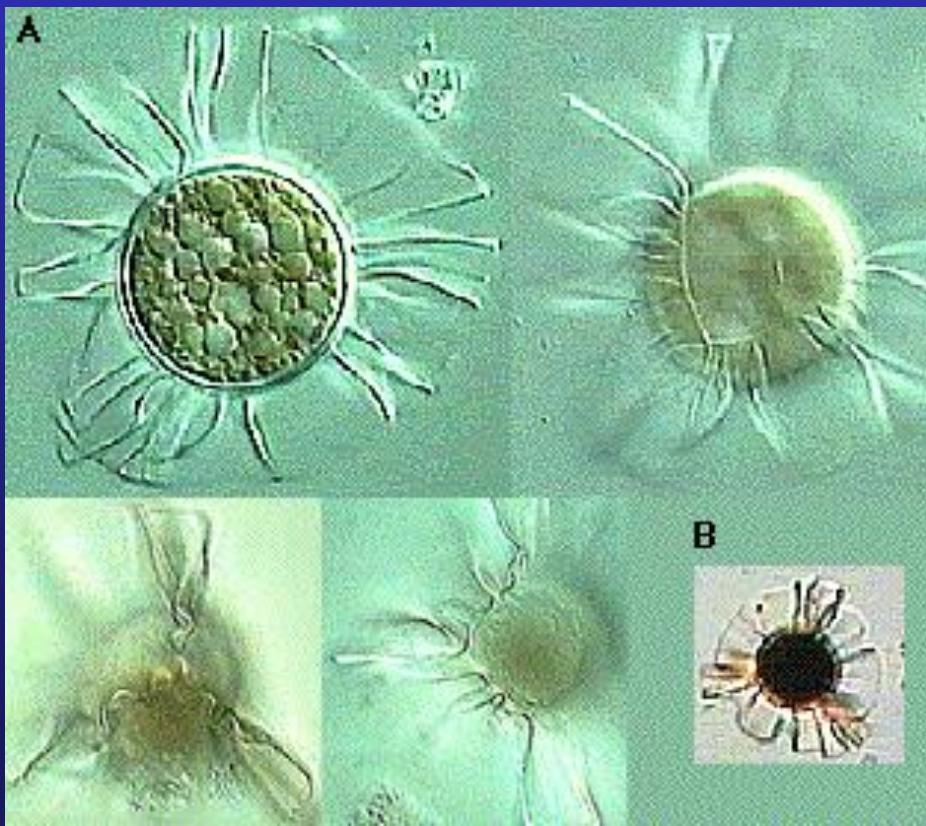
Pyramimonas tatianae longitudinal sections through division stage



Phycoma

- asexual cysts, resistant outer cell wall
- fossil records from Precambrium (250-540 mya)

Halosphaera



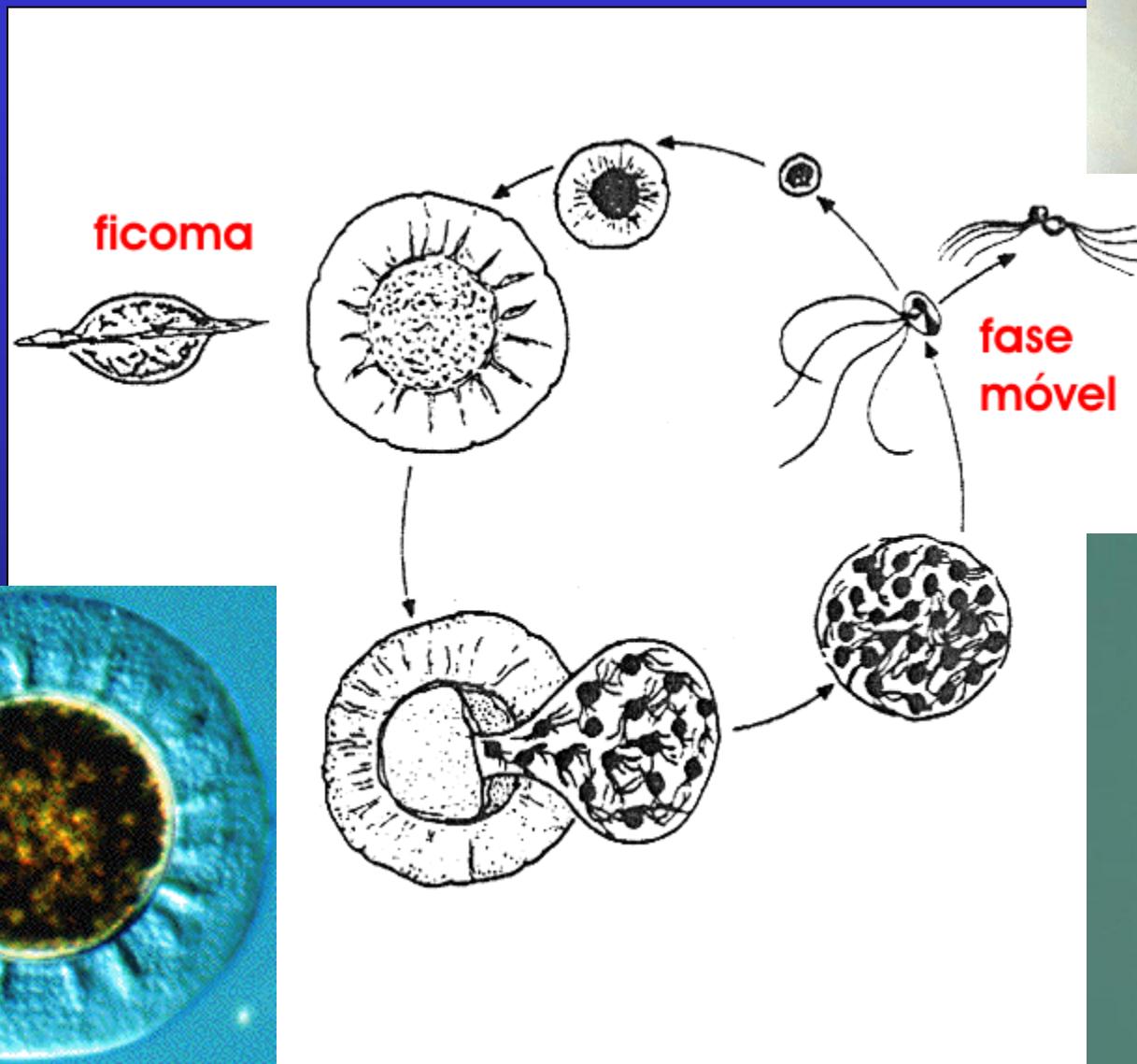
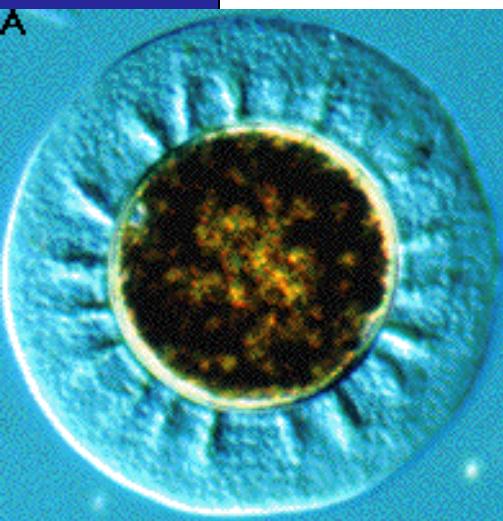
Pterosperma

Pyramimonadales

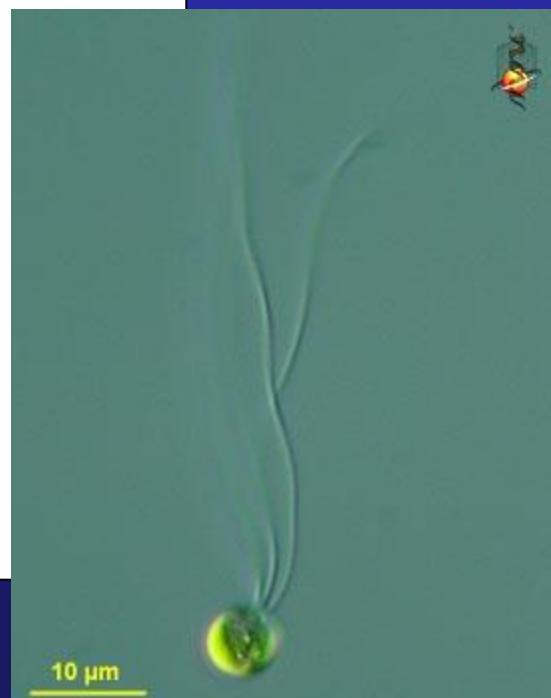


Halosphaera – during spring and early summer, phycomata can accumulate in surface layers (carried by currents and wind) and cause a water blooms – green oily water.

Pyramimonadales



lipids
sporopolenin

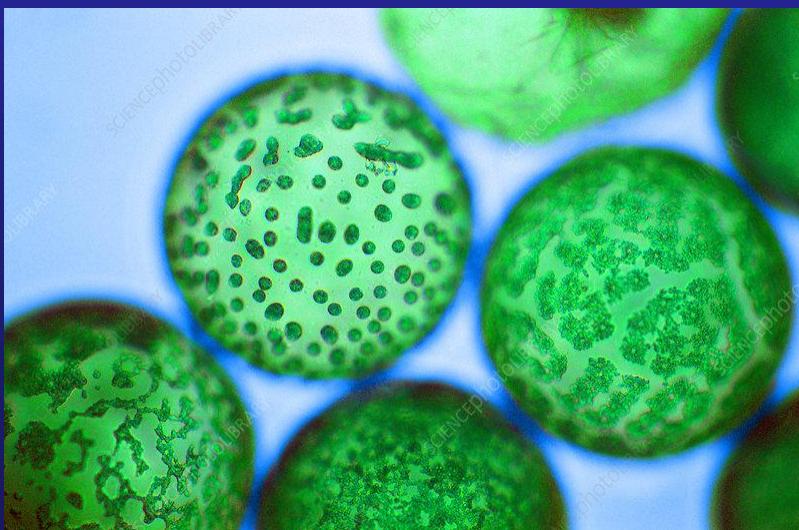


Pterosperma – live cycle

Pyramimonadales

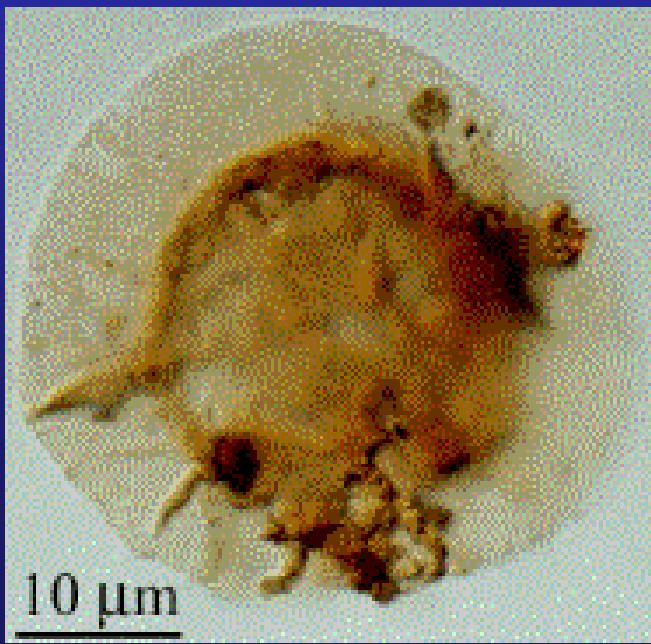


Halosphaera - fagotrophy,
mixotrophic nutrition



calm winter seas
Phycoma (=asexual cyst)

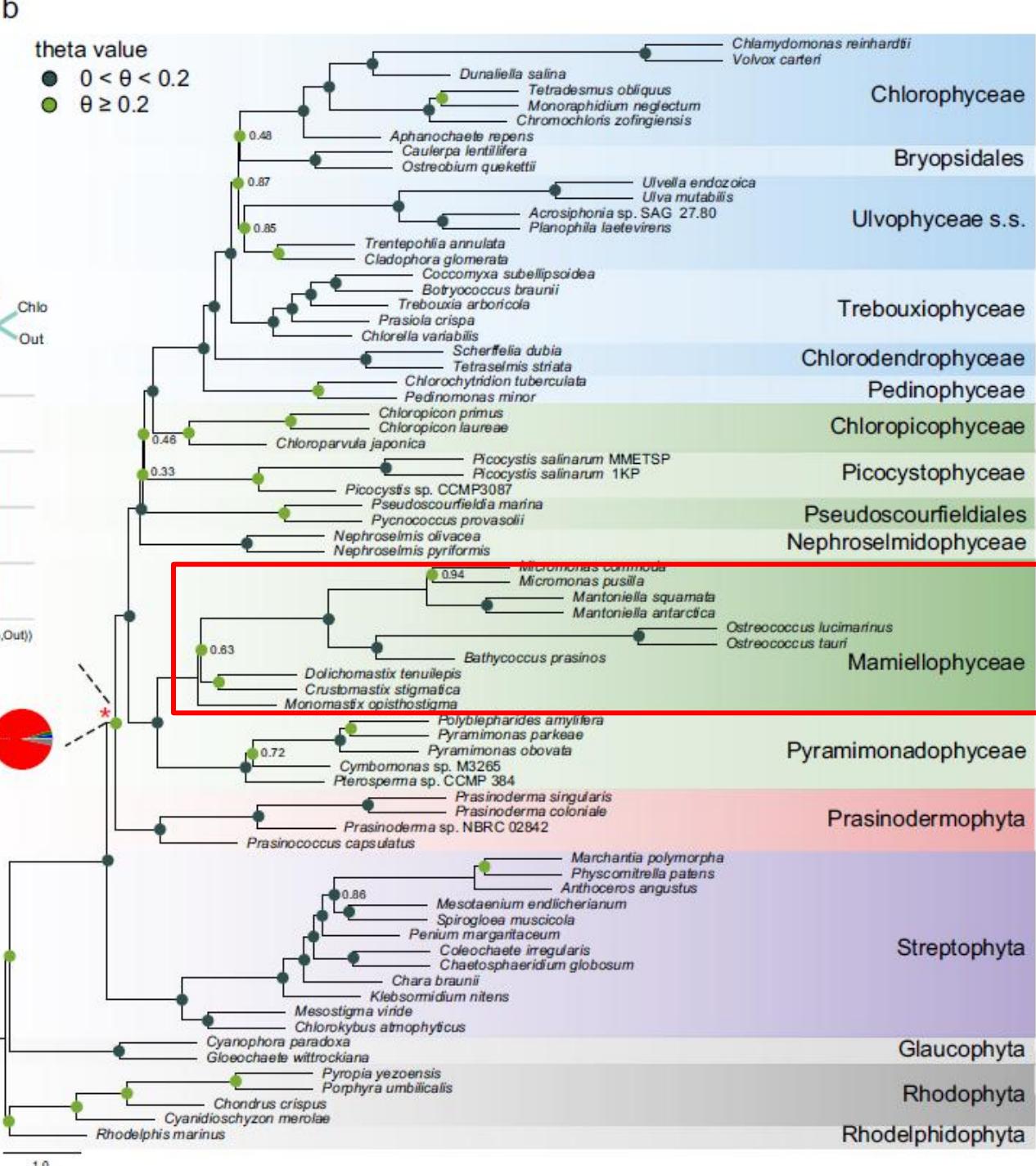
Fossil species



Tasmanites

600 mya - Precambrium

Pterospermella



Yang et al. 2023

<https://doi.org/10.1038/s41467-023-41137-5>

Monomastigales



trychocysts=
ejectosomes

Light micrographs of *Monomastix opisthostigma*

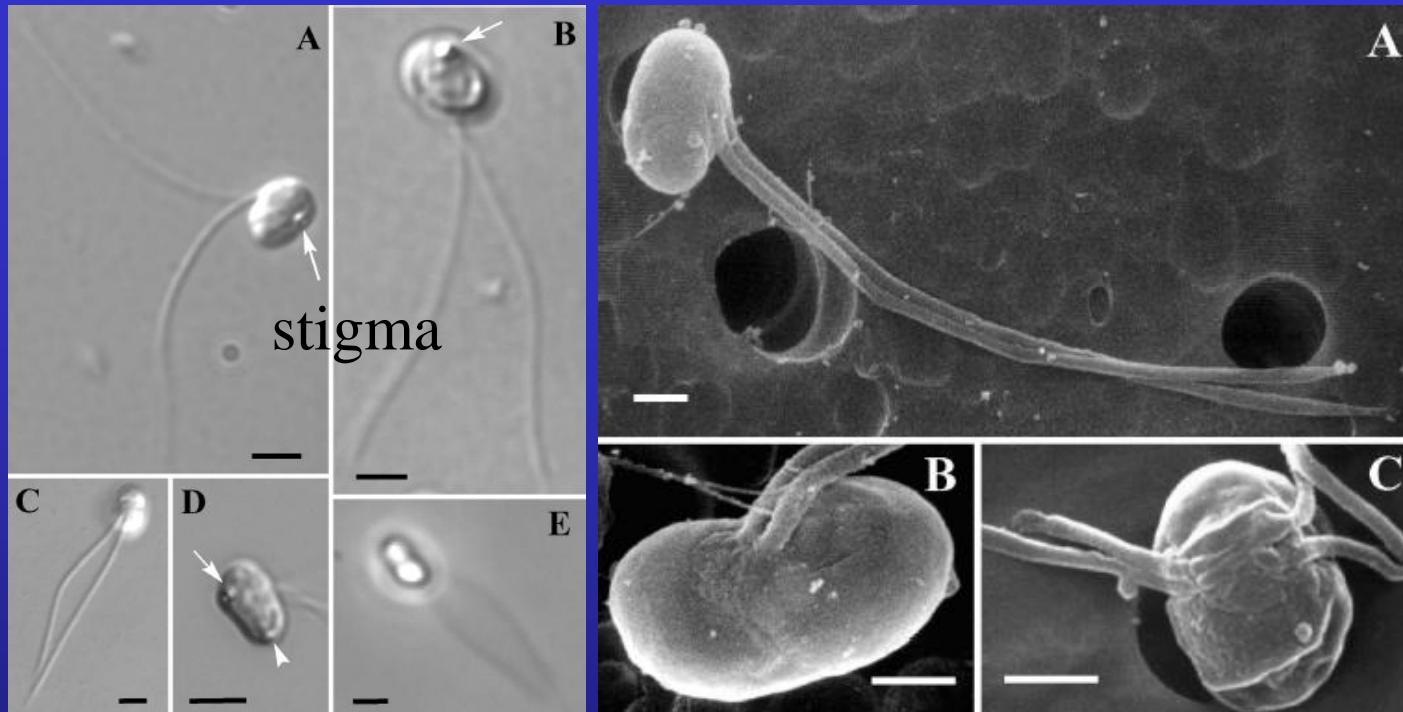
basal lineage – large cells up to 20 μm

Dolichomastigales and the Mamiellales cells are usually no longer than 5 μm and in two lineages (Bathycoccaceae and *Micromonas*) are reduced to the size of the smallest eukaryotes (1 μm ; Courties et al. 1994). Cell size reduction is accompanied by loss of the flagellar apparatus and eyespot (Bathycoccaceae) and/or the scaly covering (*Ostreococcus* and *Micromonas*).

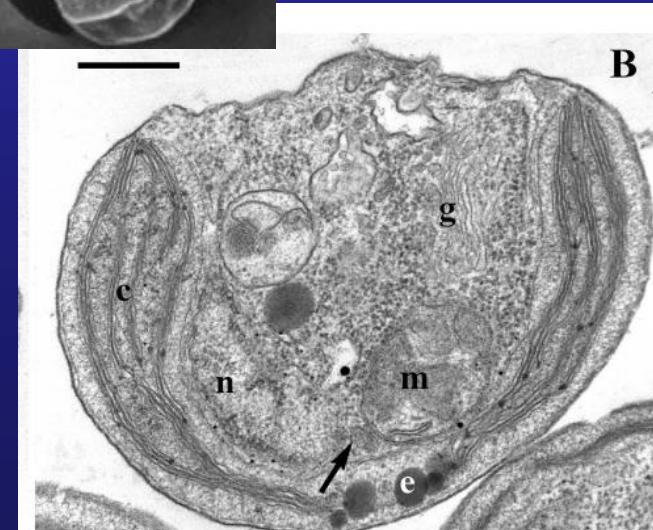
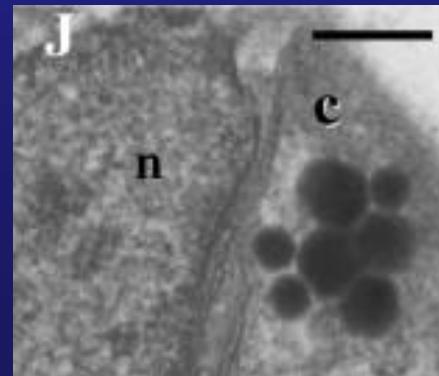
Dolichomastigales

Crustomastix stigmatica

Zingone et al. 2002

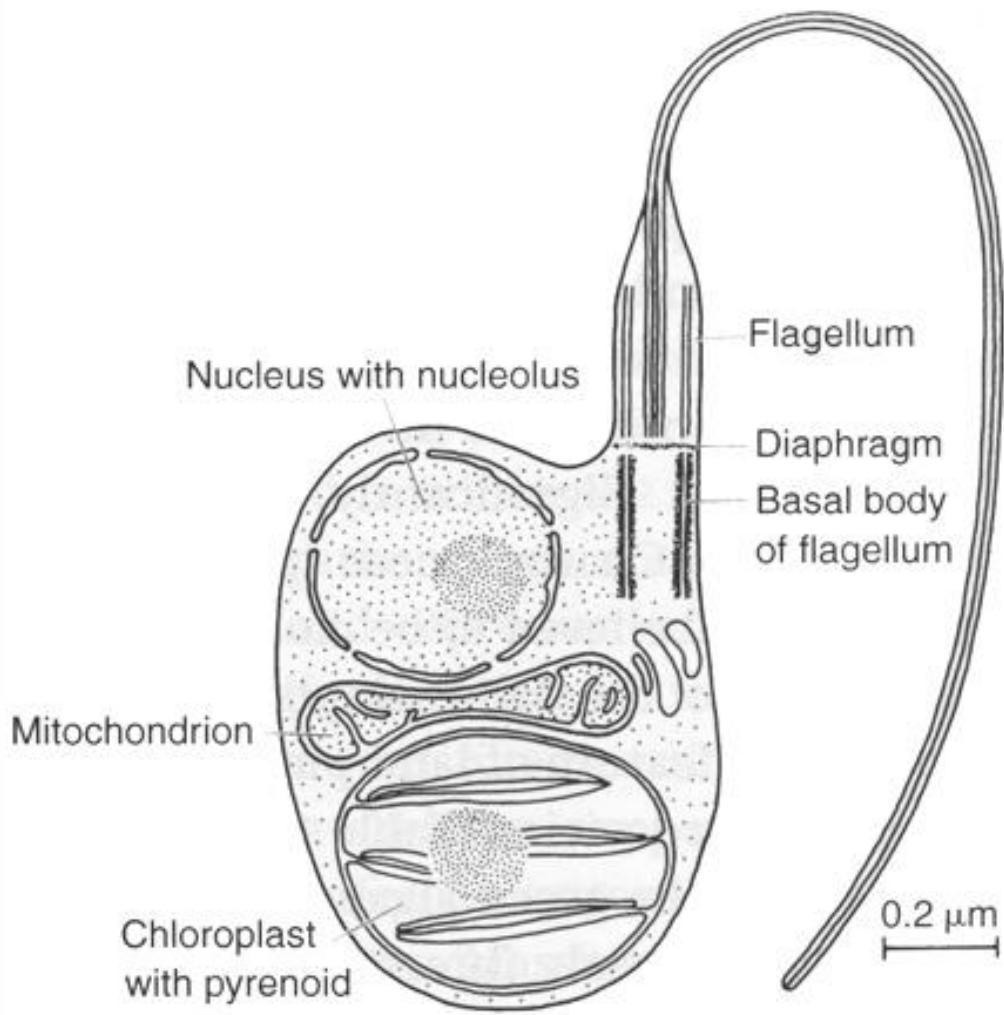


3 different types of hair-like scales on the flagella.



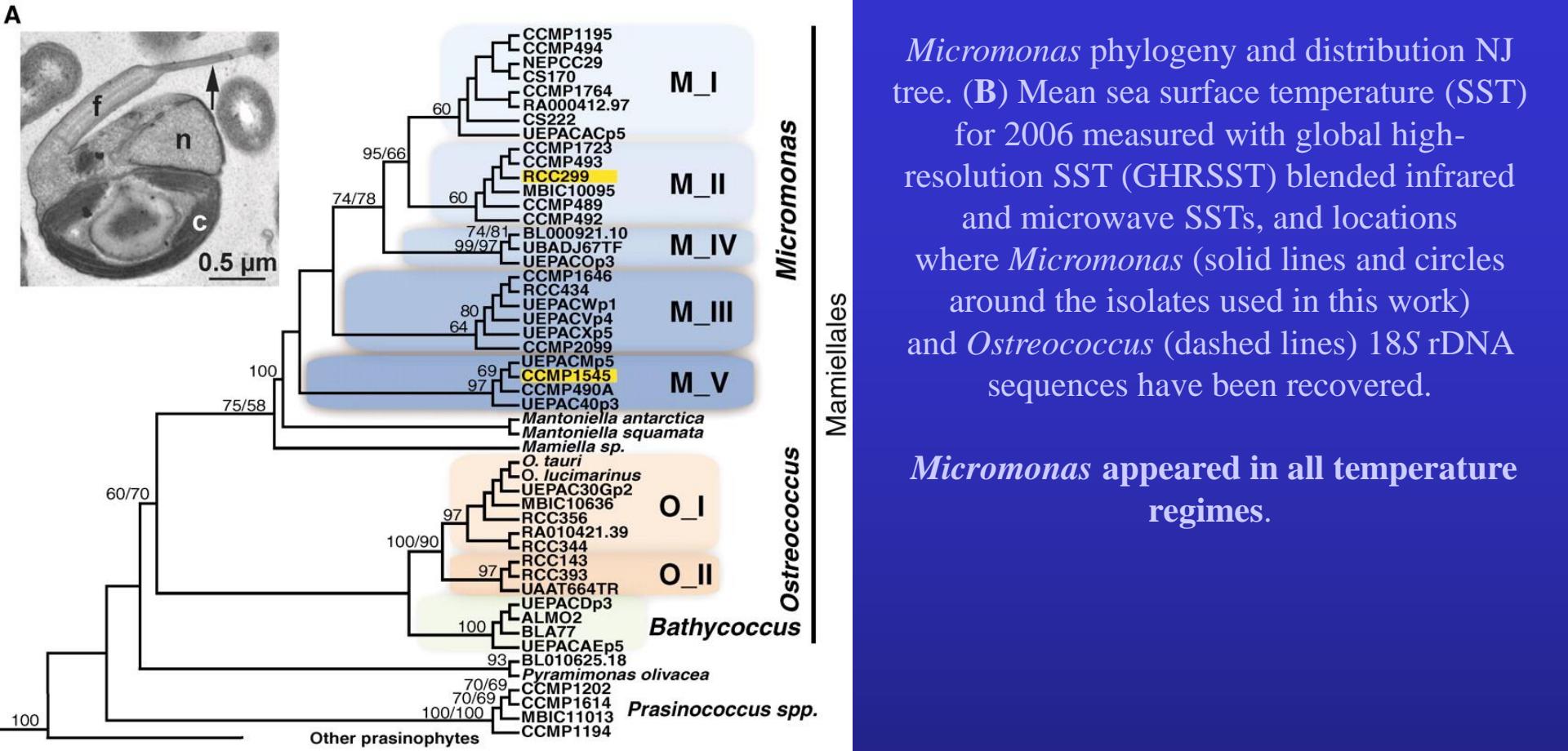
Mamiellales

Micromonas pusilla –
without scales



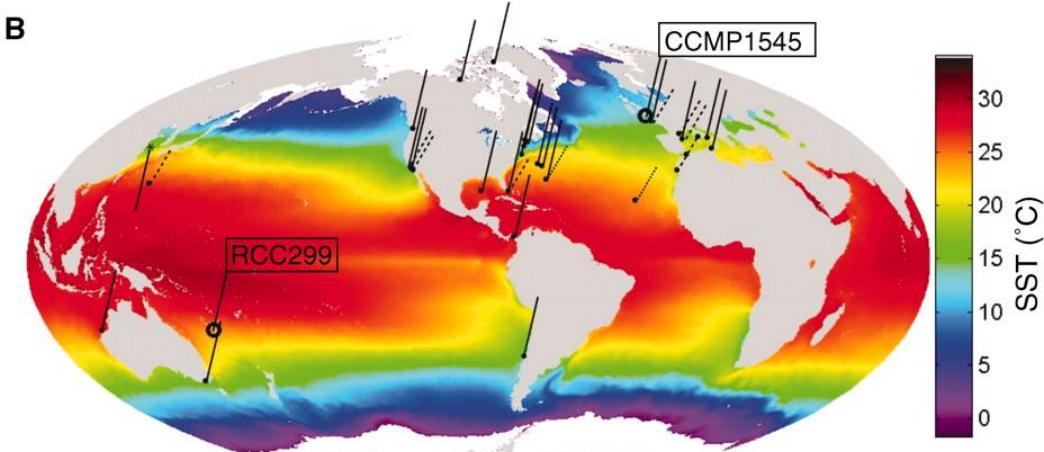
oceanic picoplankton





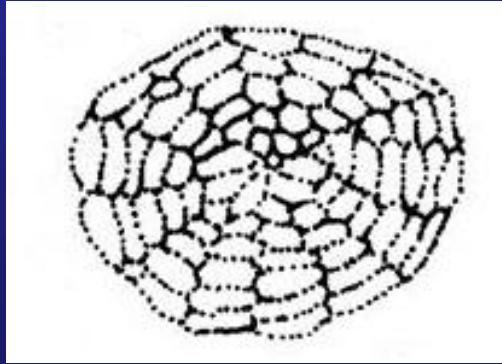
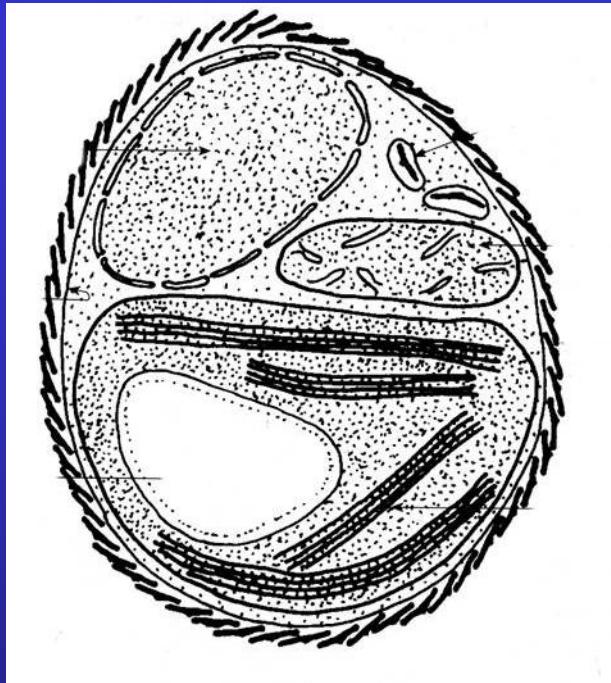
Micromonas phylogeny and distribution NJ tree. **(B)** Mean sea surface temperature (SST) for 2006 measured with global high-resolution SST (GHRSSST) blended infrared and microwave SSTs, and locations where *Micromonas* (solid lines and circles) and *Ostreococcus* (dashed lines) 18S rDNA sequences have been recovered.

Micromonas appeared in all temperature regimes.

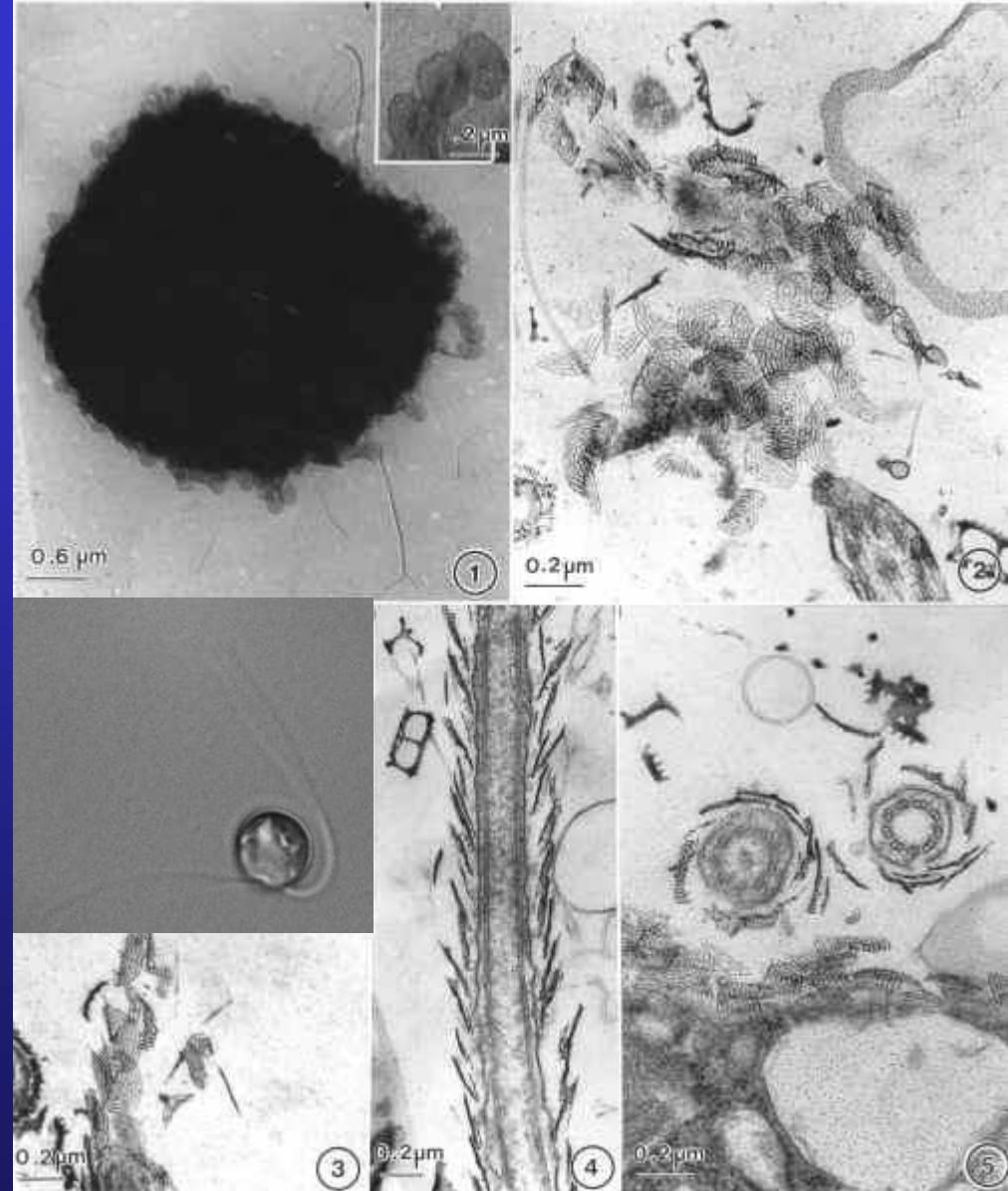


Worden *et al.* 2009

Mamiellophyceae

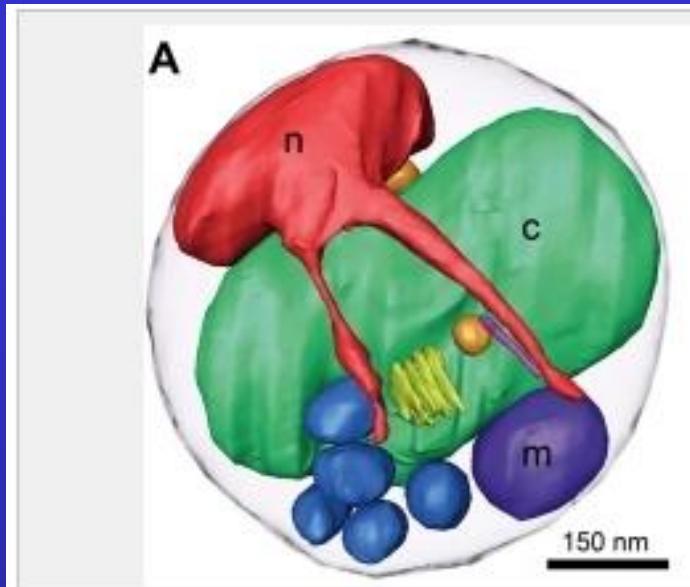


Bathycoccus – lost of flagellum
North Atlantic



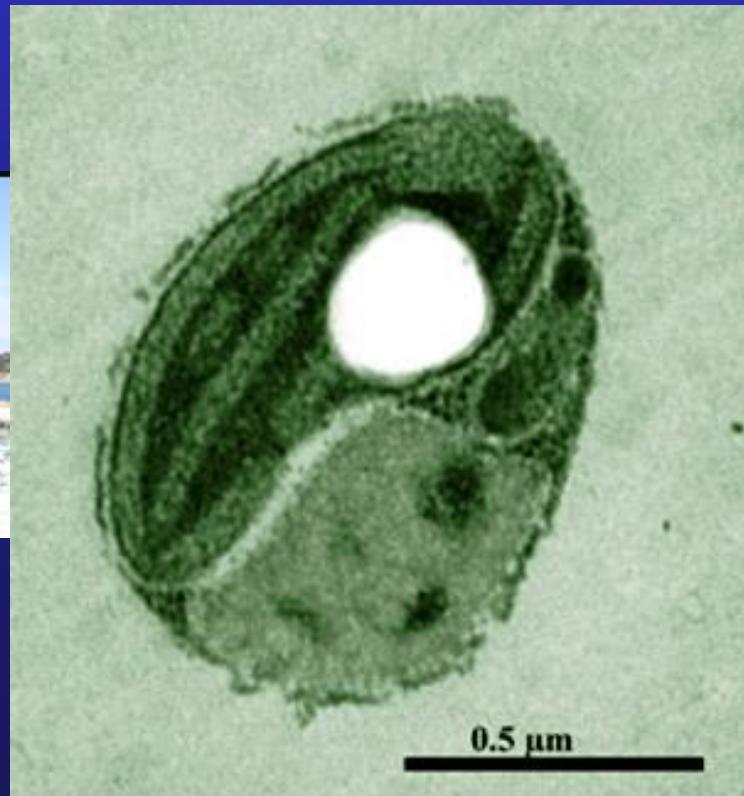
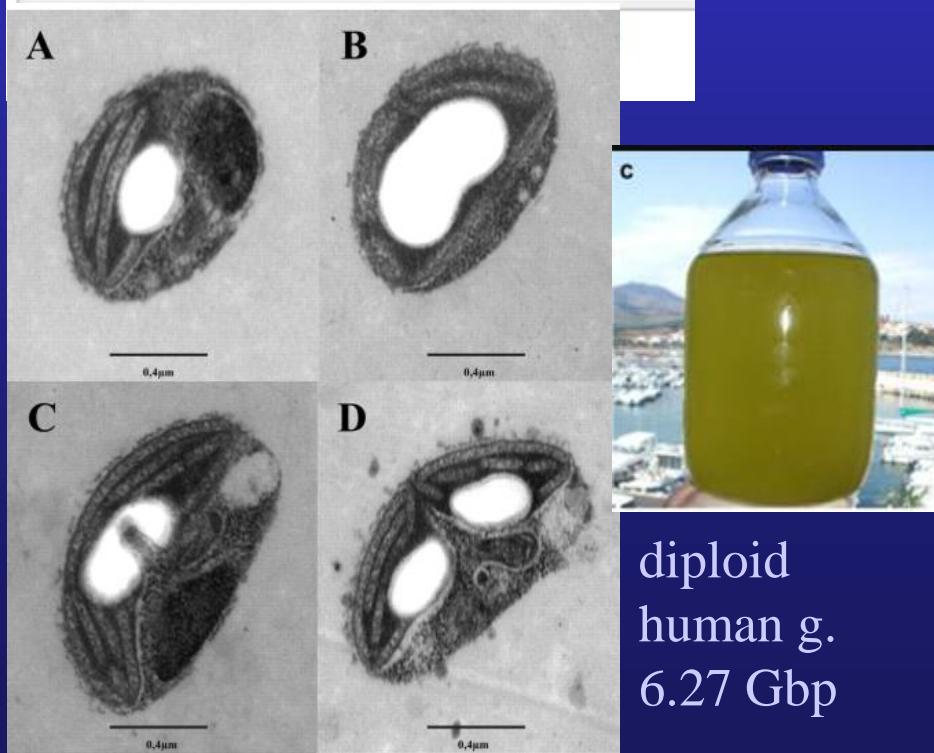
Mamiella gilva

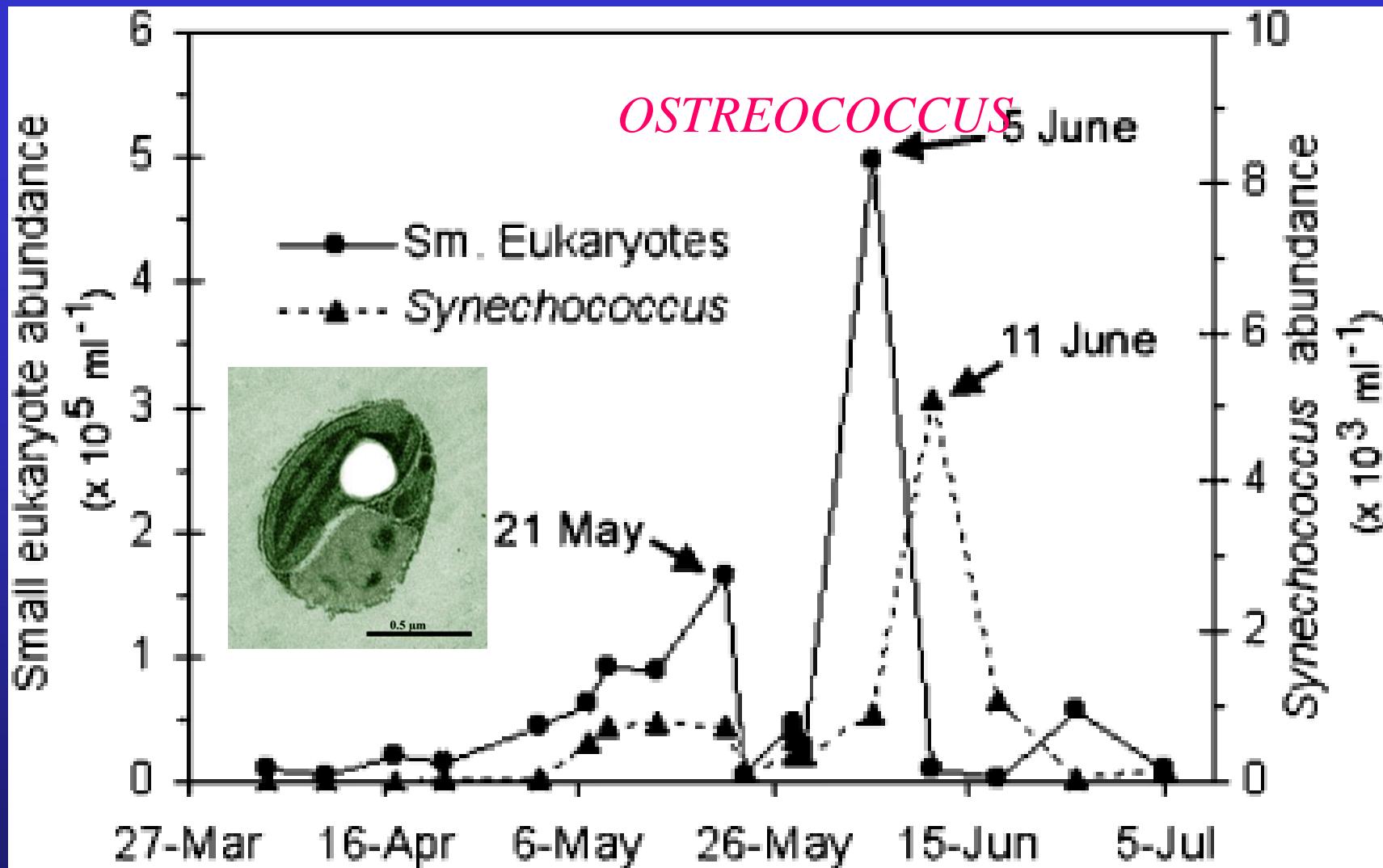
Mamiellales



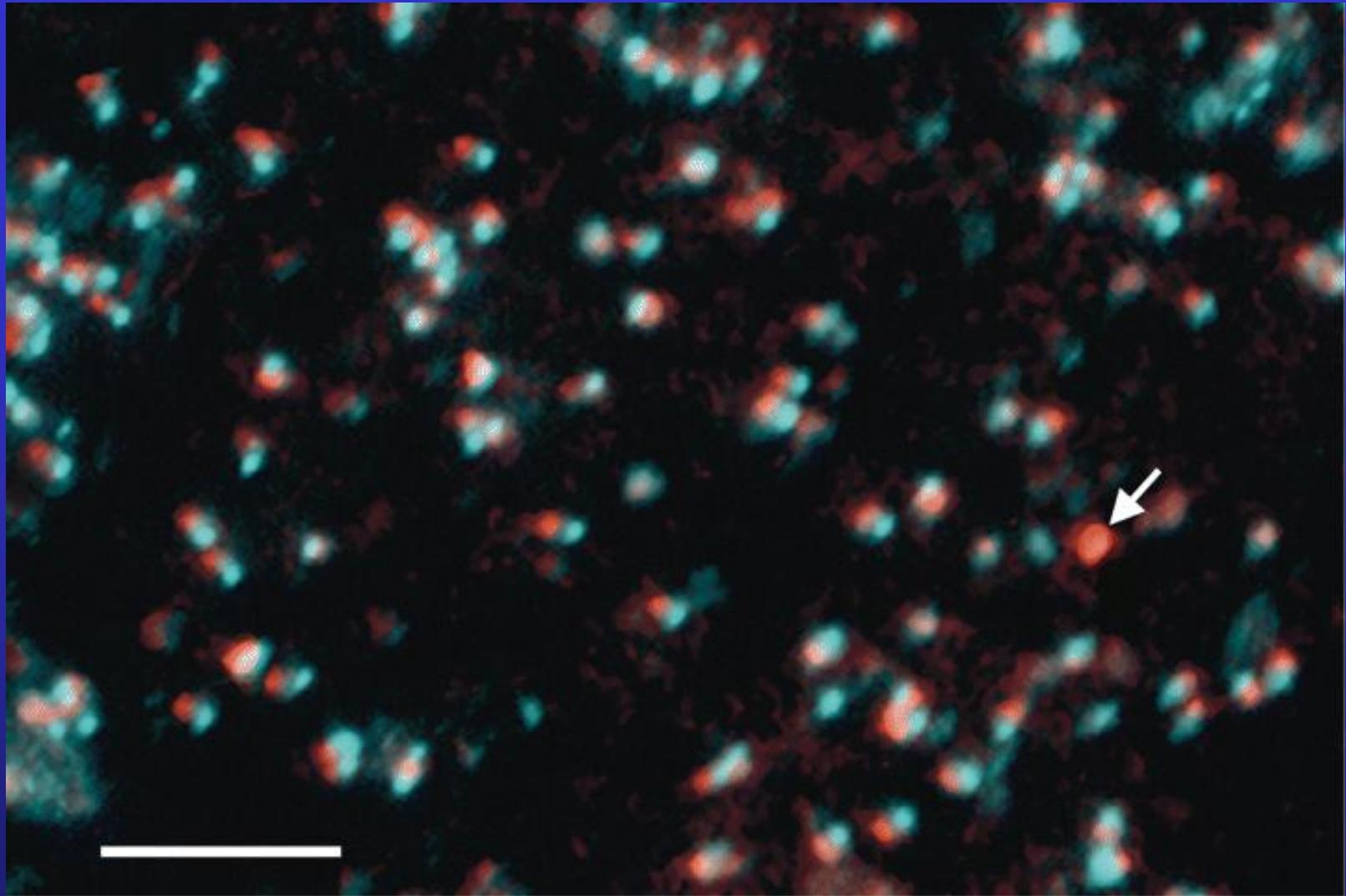
Ostreococcus belongs to the **Mamiellales** and is reported as a globally abundant, single-celled alga thriving in the upper (illuminated) water column of the oceans.

Blooms in the sea, the smallest eukaryotic genome (12 Mbp) – densely packed genome





1. Abundances of small eukaryotes and *Synechococcus* cells over the 2001 study period in West Neck Bay, New York as counted by flow cytometry. Major bloom events are indicated. Note difference in scales.



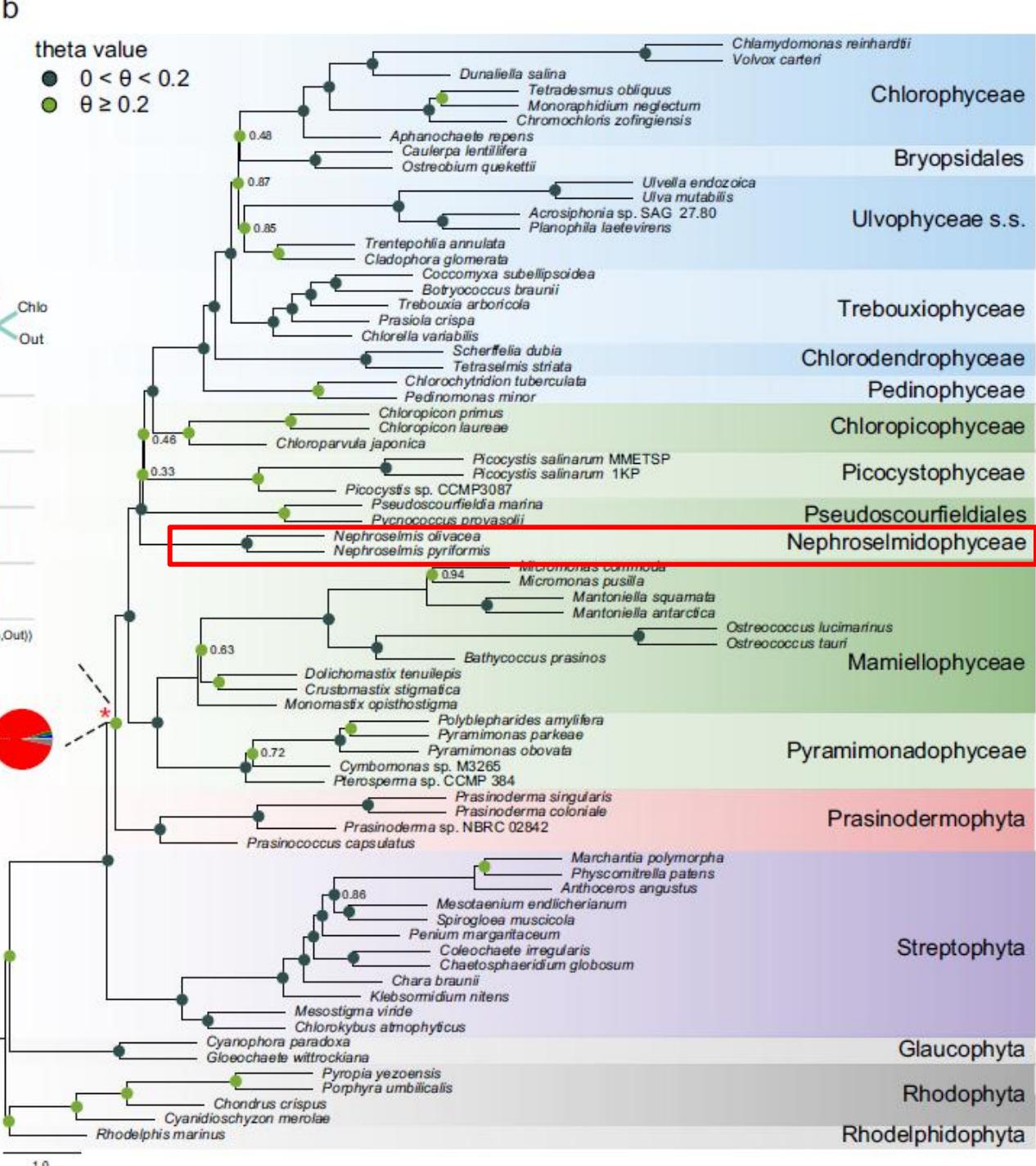
Color composite epifluorescence micrograph showing DAPI-stained *Ostreococcus*-like cells with nuclei (cyan) and chloroplasts (red autofluorescence). A single *Synechococcus* cell (arrow) is also present. Scale bar, 10 μm .

O'Kelly et al. 2003

Mamiellophyceae

Do they reproduced sexually???

Ostreococcus as well as *Micromonas* may contain meiotic genes and thus reproduce sexually (Derelle et al. 2006; Worden et al. 2009), but experimental proof for sexual reproduction in the Mamiellophyceae is still lacking.



Yang et al. 2023

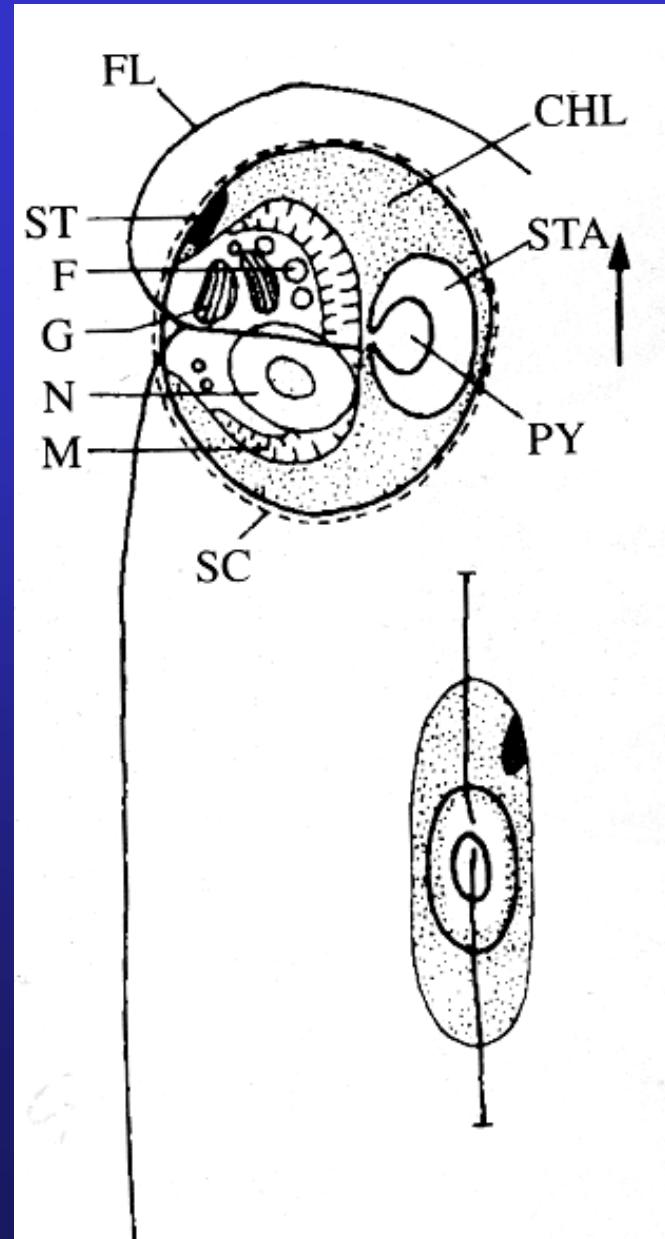
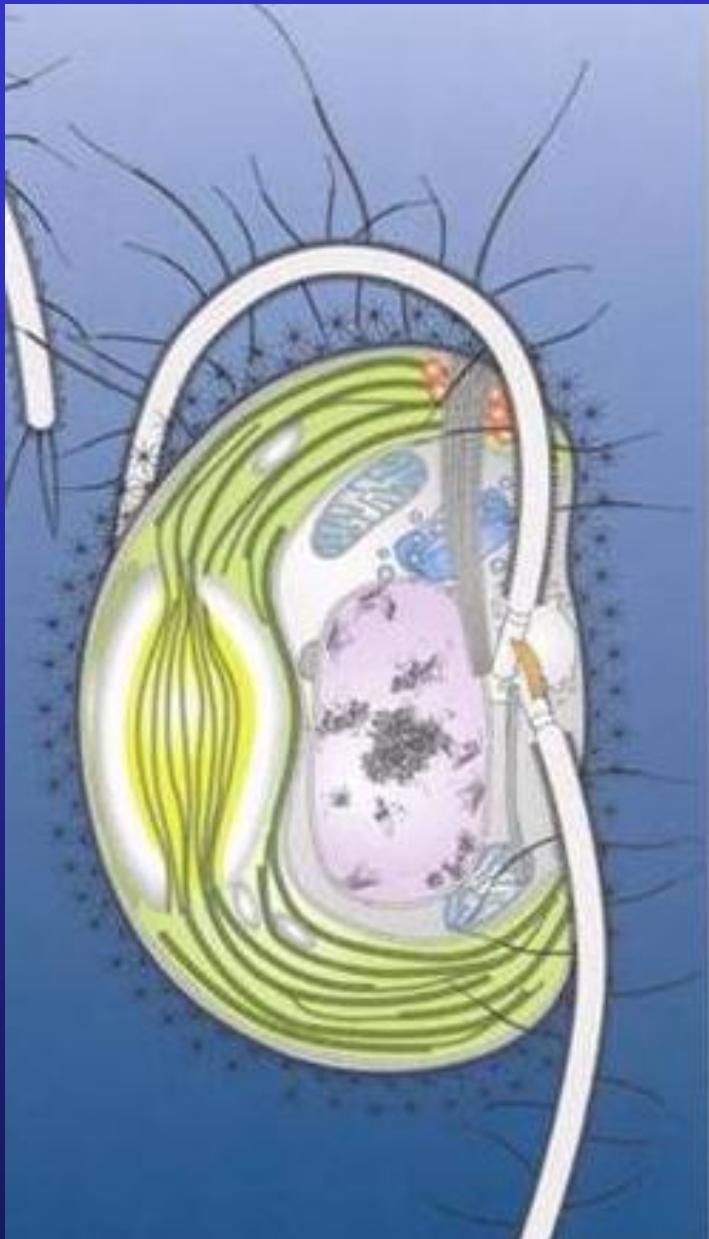
<https://doi.org/10.1038/s41467-023-41137-5>

Nephroselmidophyceae

Nephroselmis



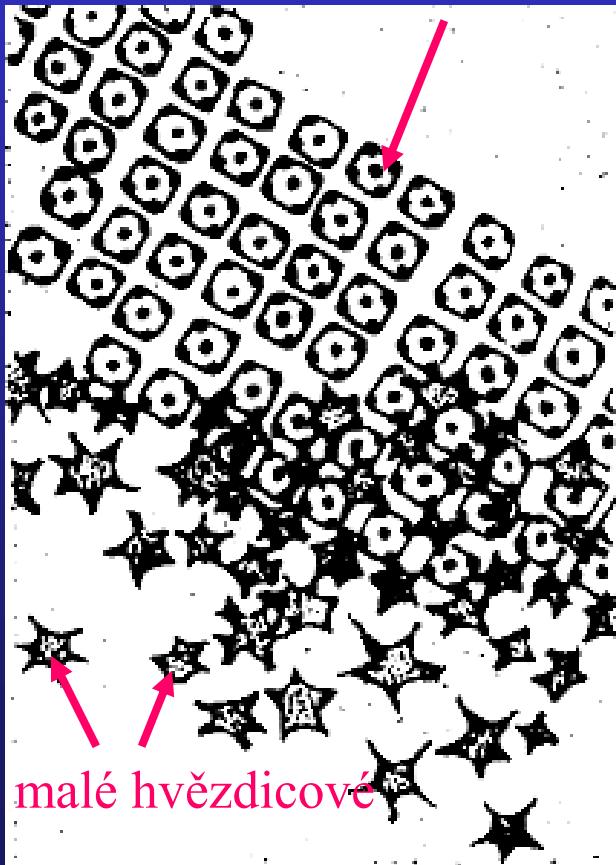
swiming



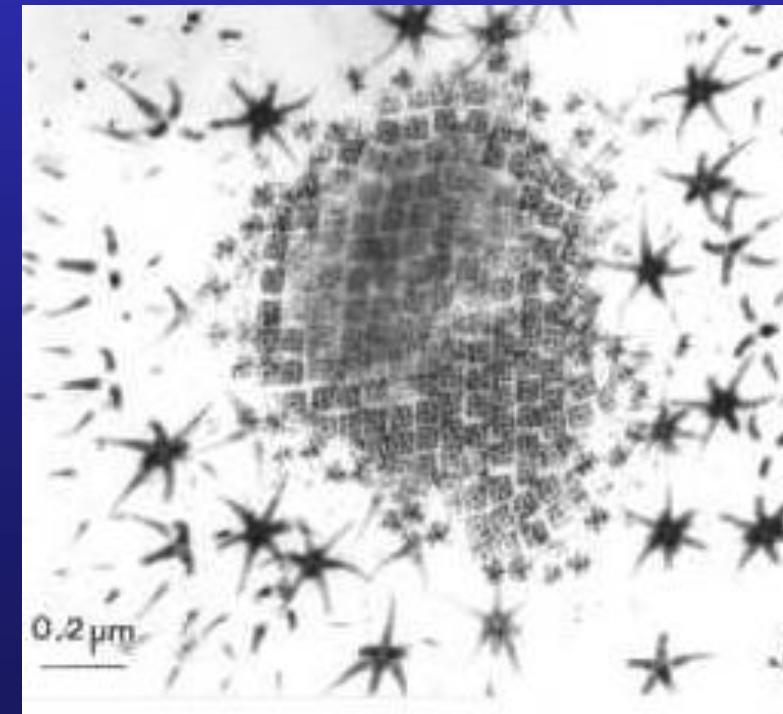
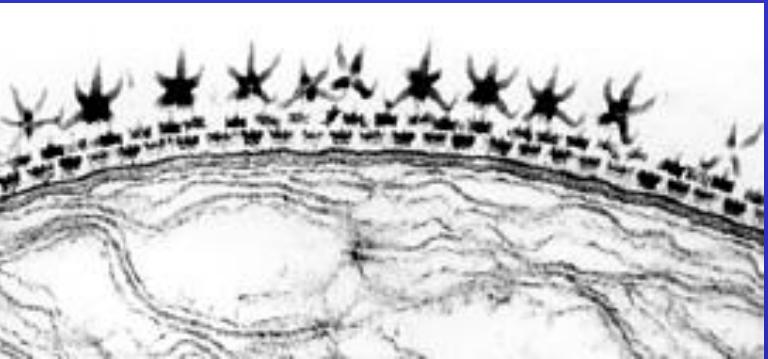
Organic scales

Nephroselmis

malé čtvercové

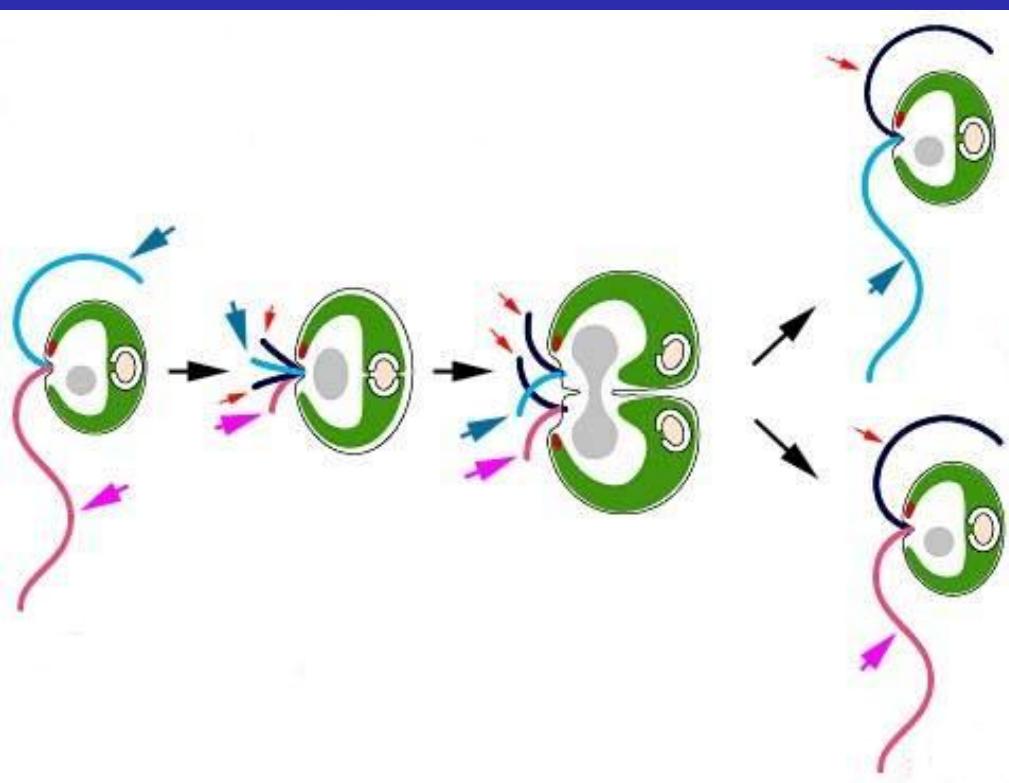
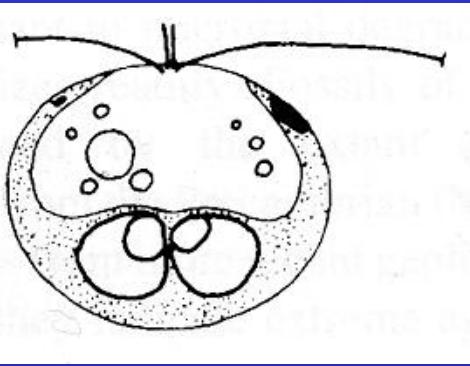


velké hvězdicové



Mitosis and cytokinesis

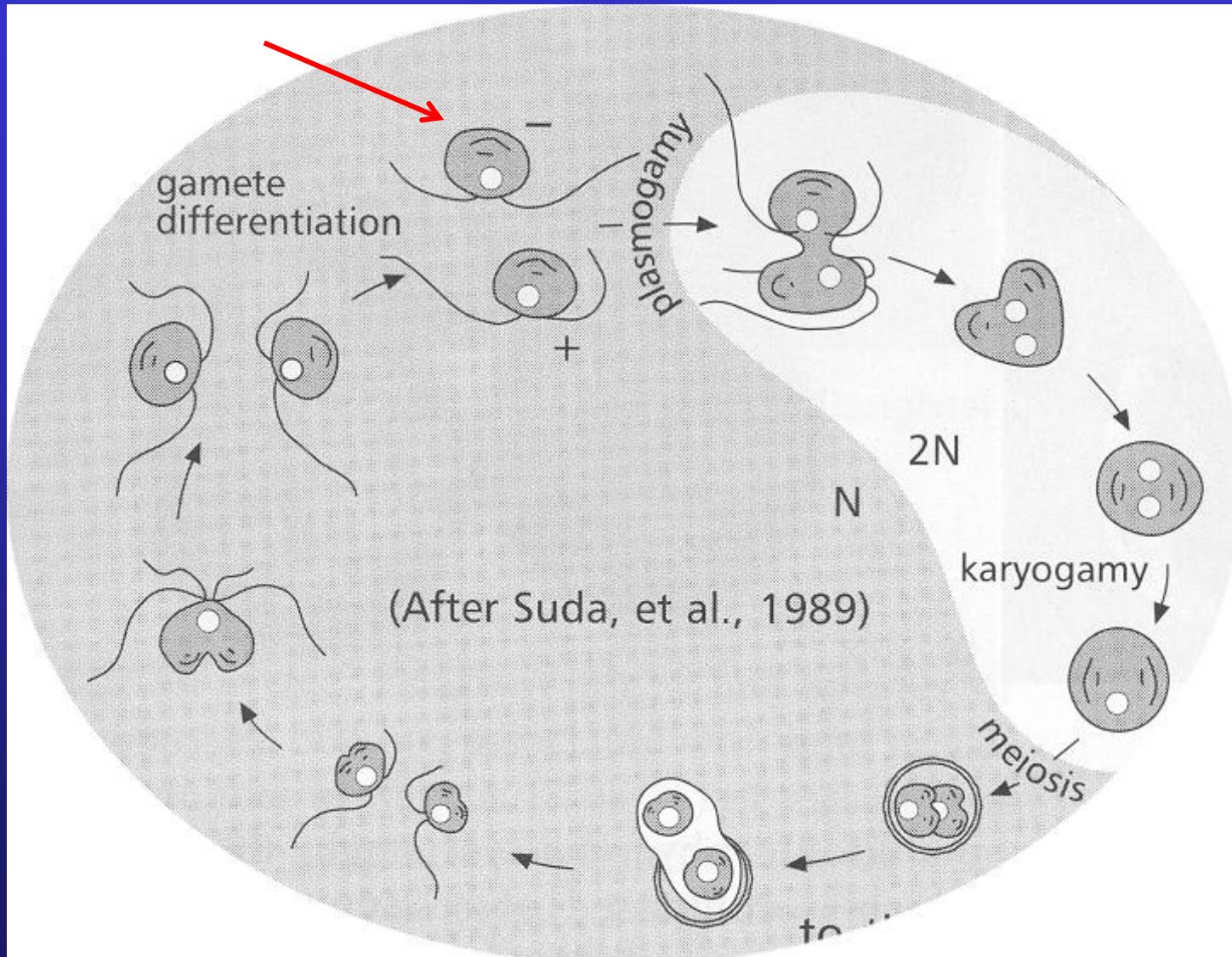
- schizotomy



Nephroselmis rotunda

Sexual reproduction

Haplontic life cycle, izogamy, heterothalism



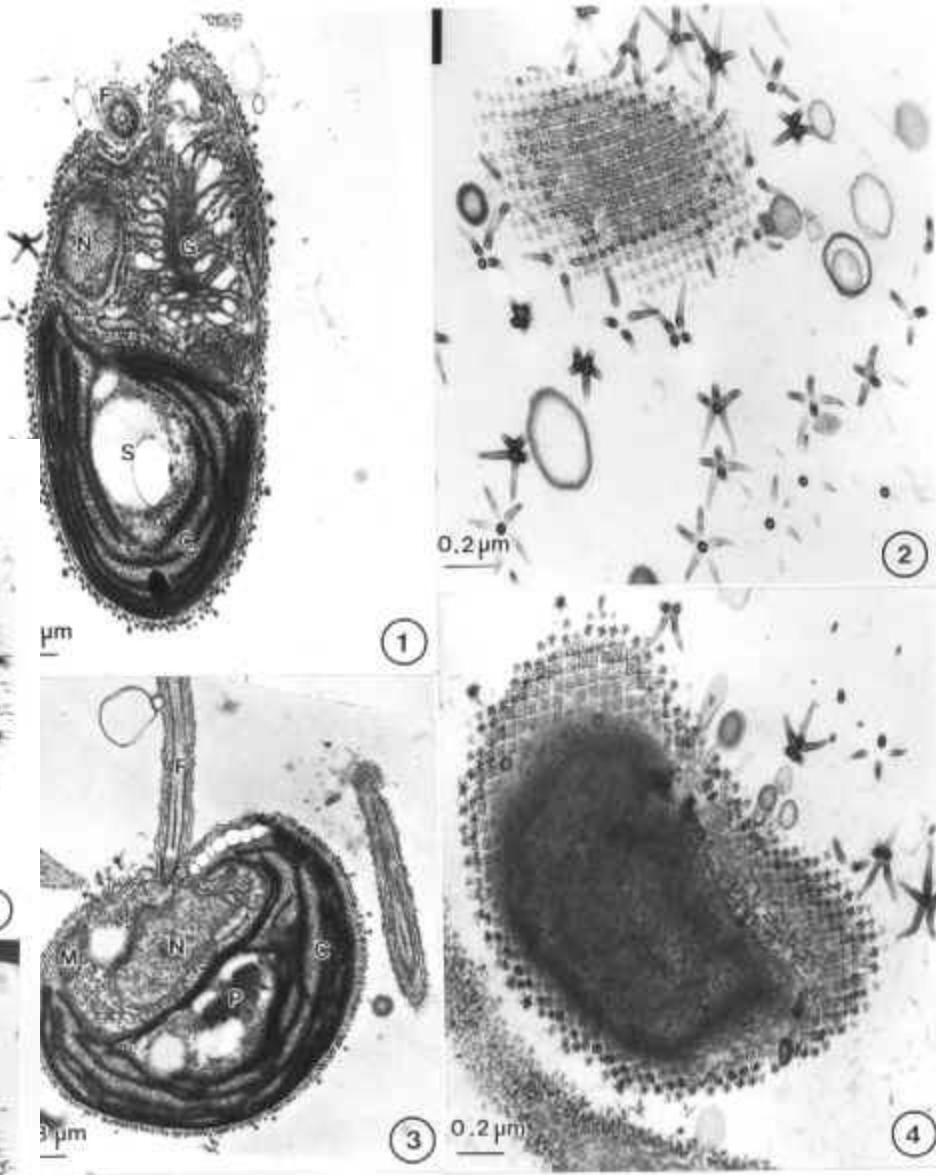
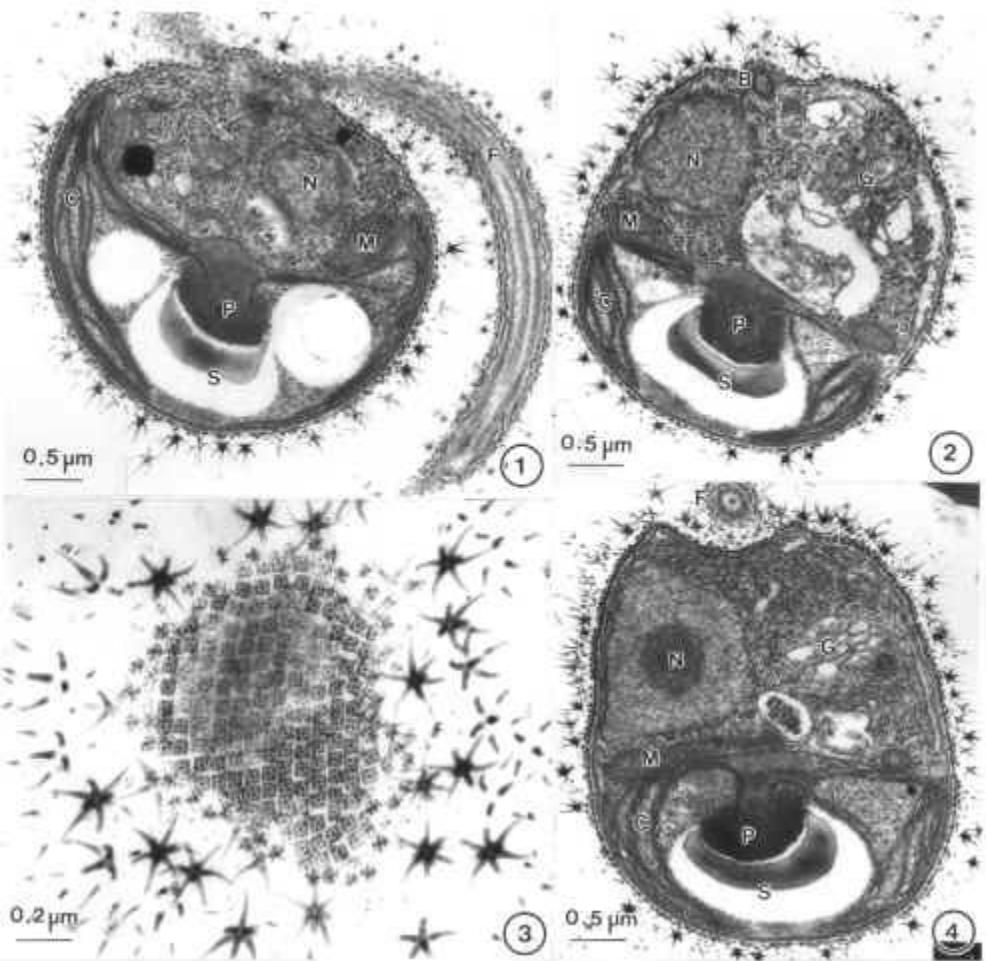
Nephroselmis olivacea

Nephroselmis - ultrastructure

malé čtvercové

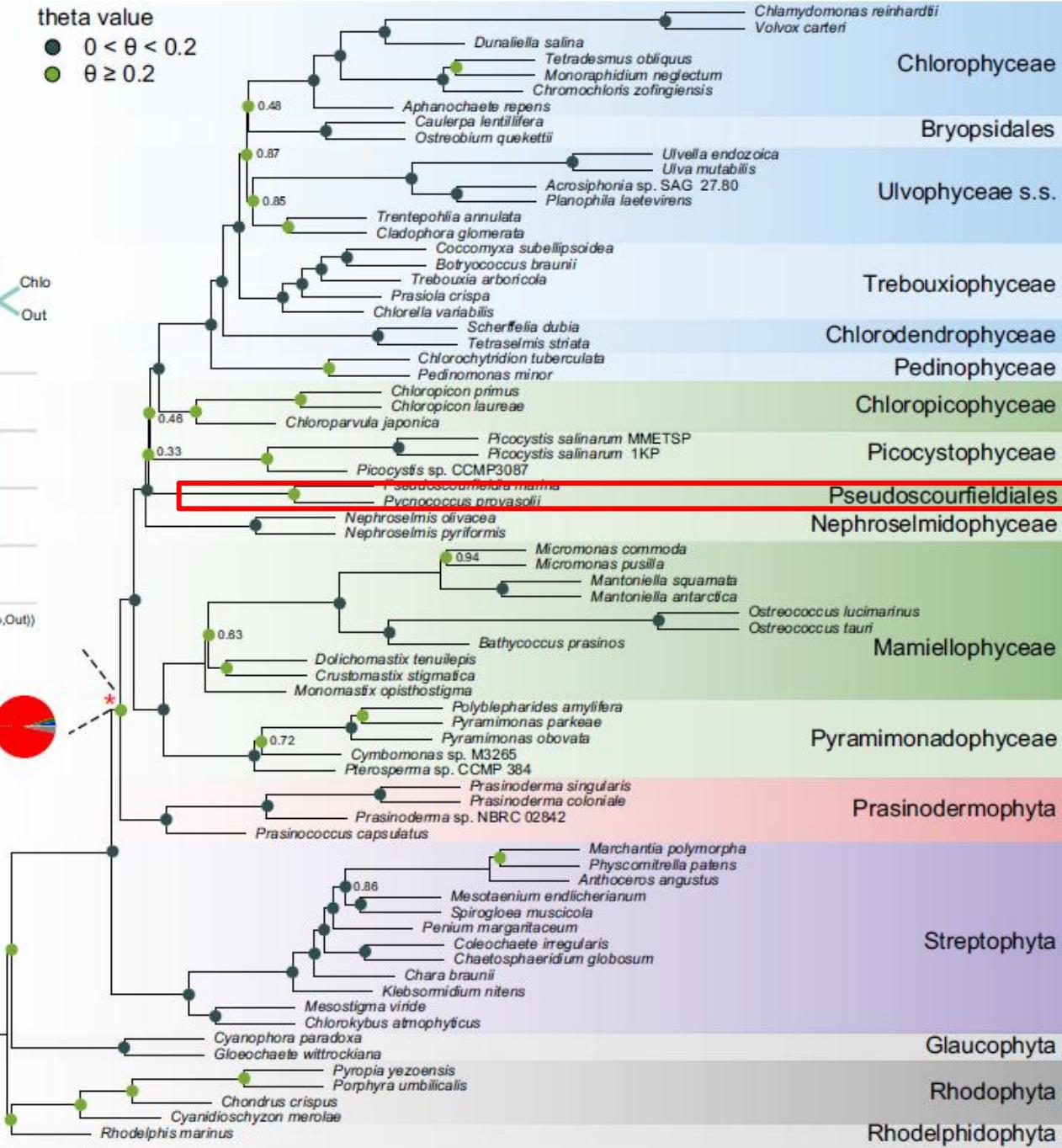
malé hvězdicové

velké hvězdicové

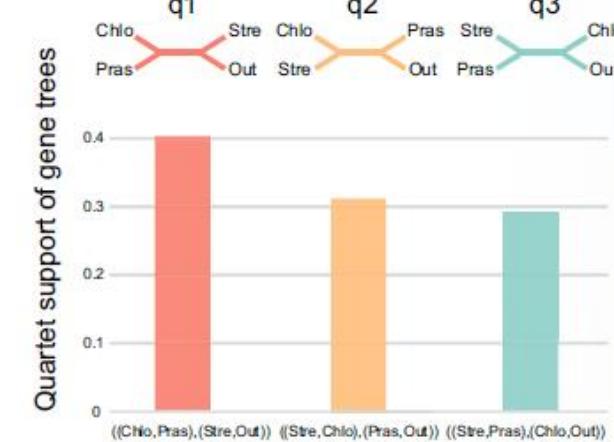


b

theta value
 ● $0 < \theta < 0.2$
 ● $\theta \geq 0.2$



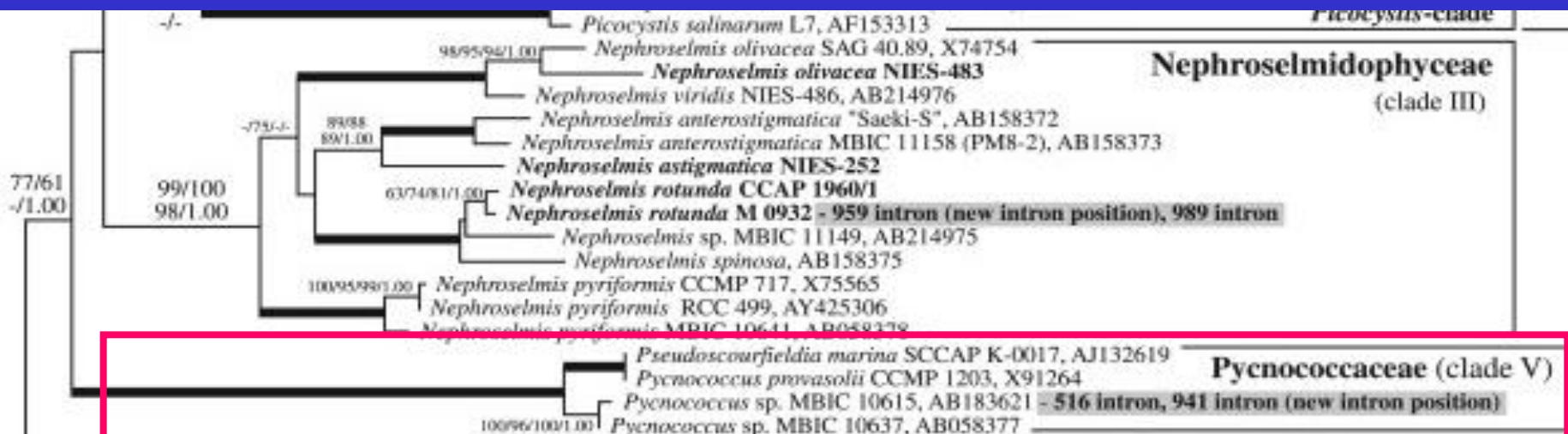
c



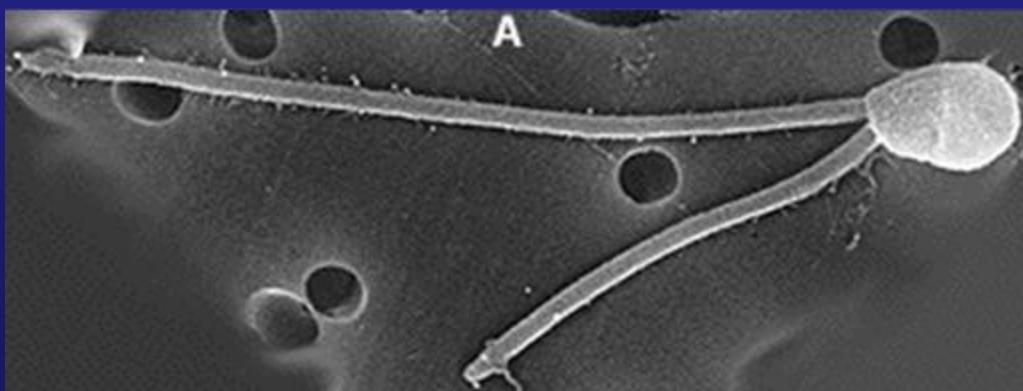
Yang et al. 2023

<https://doi.org/10.1038/s41467-023-41137-5>

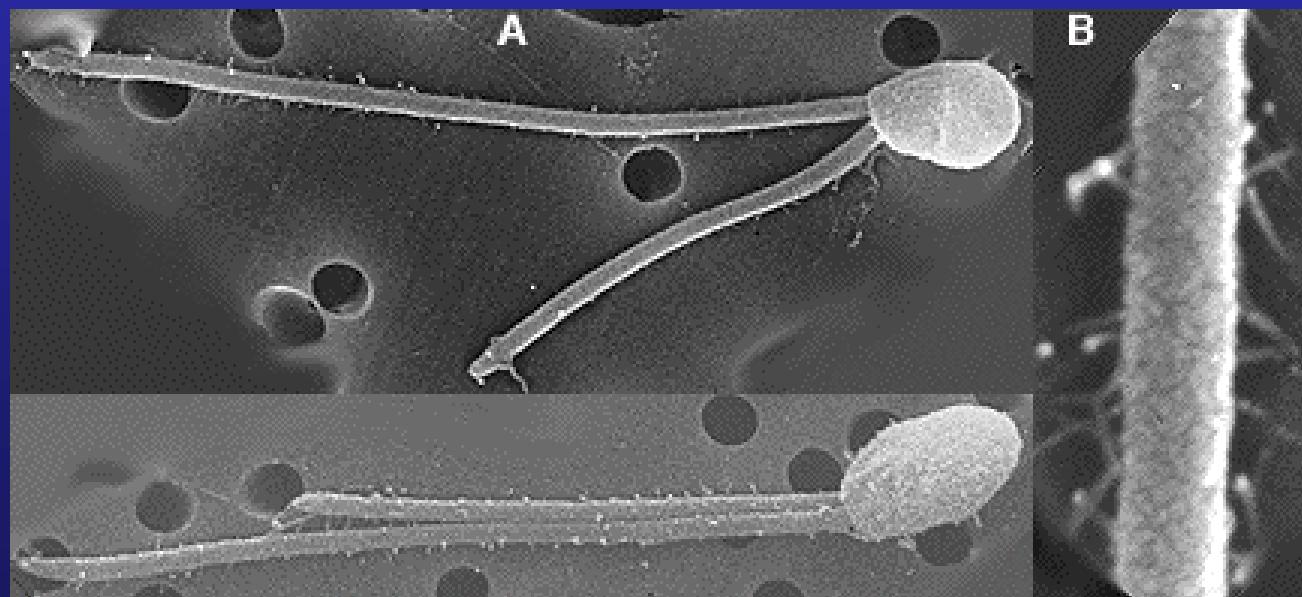
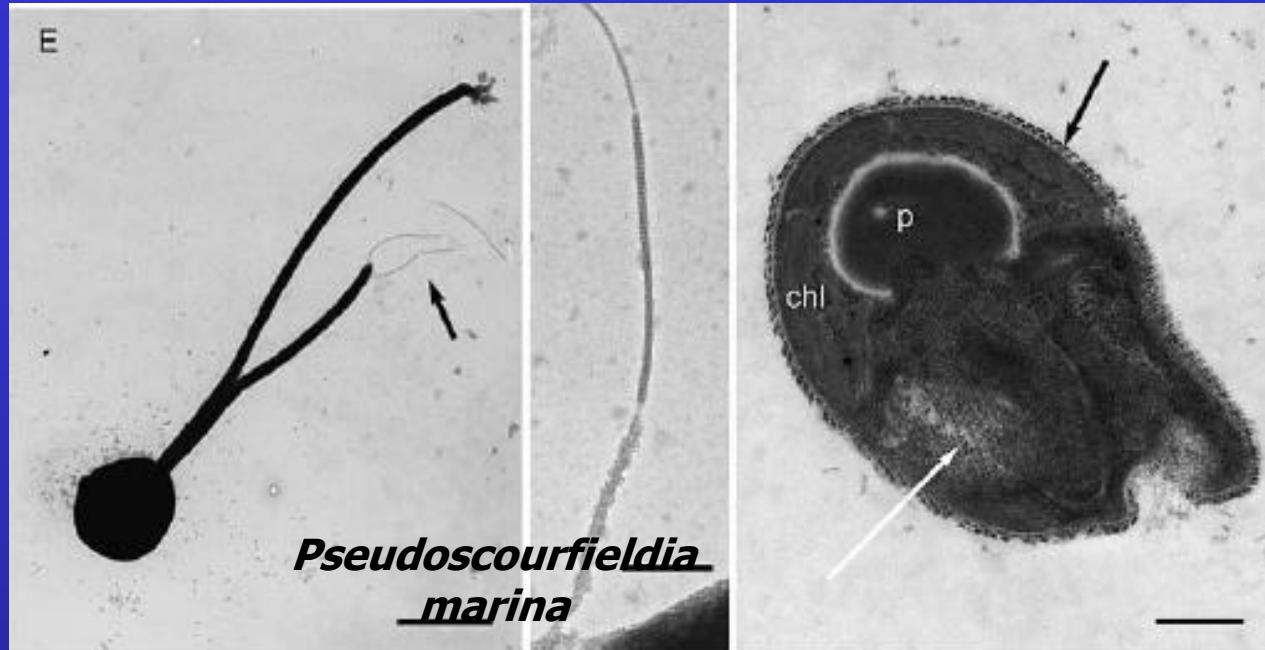
Pseudoscourfieldiales

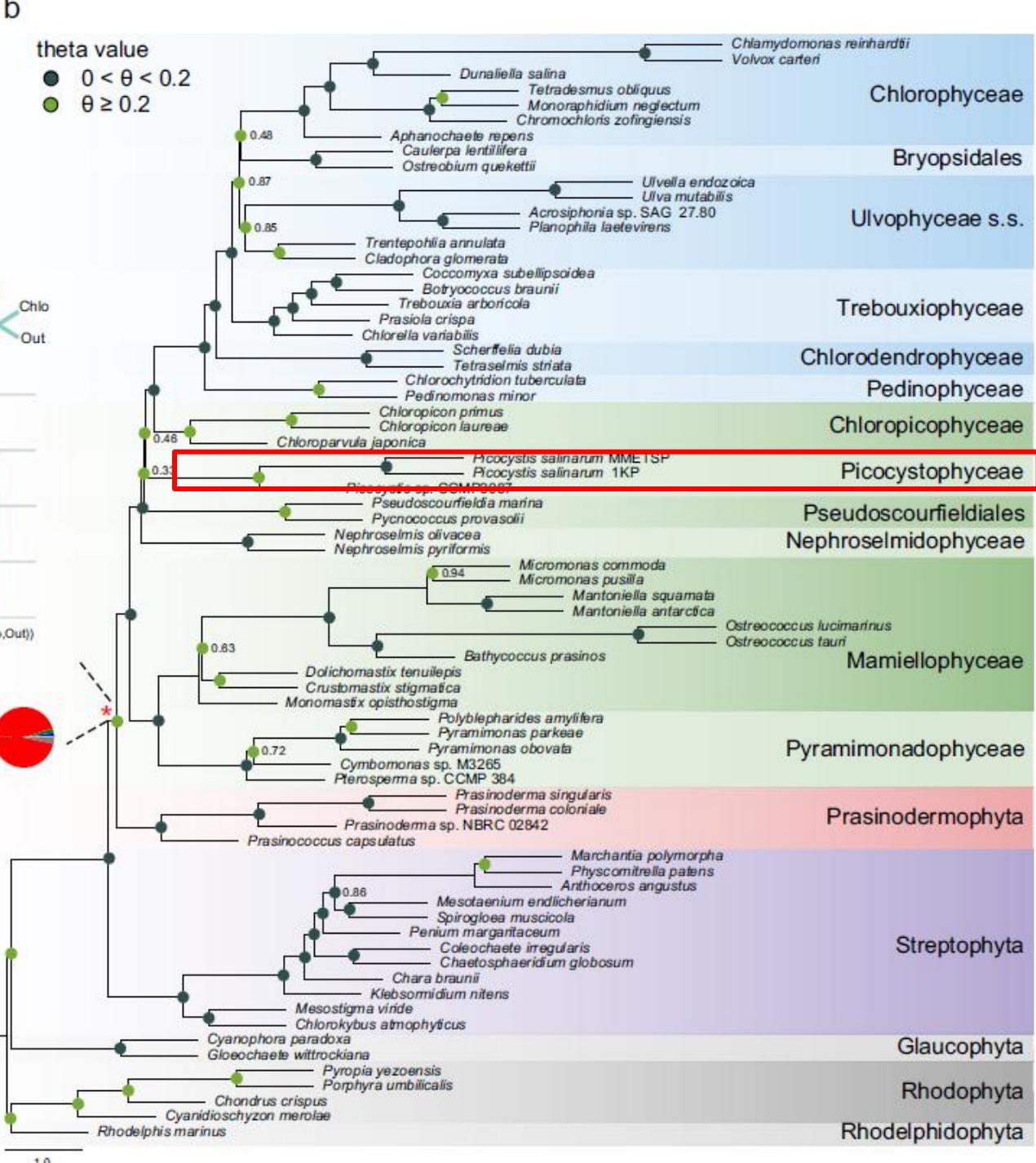


Pseudoscourfieldia marina and *Pycnococcus provasolii* are genetically identical and presumably represent different life history stages of a single taxon; unpubl.



Guillou et al.
2004

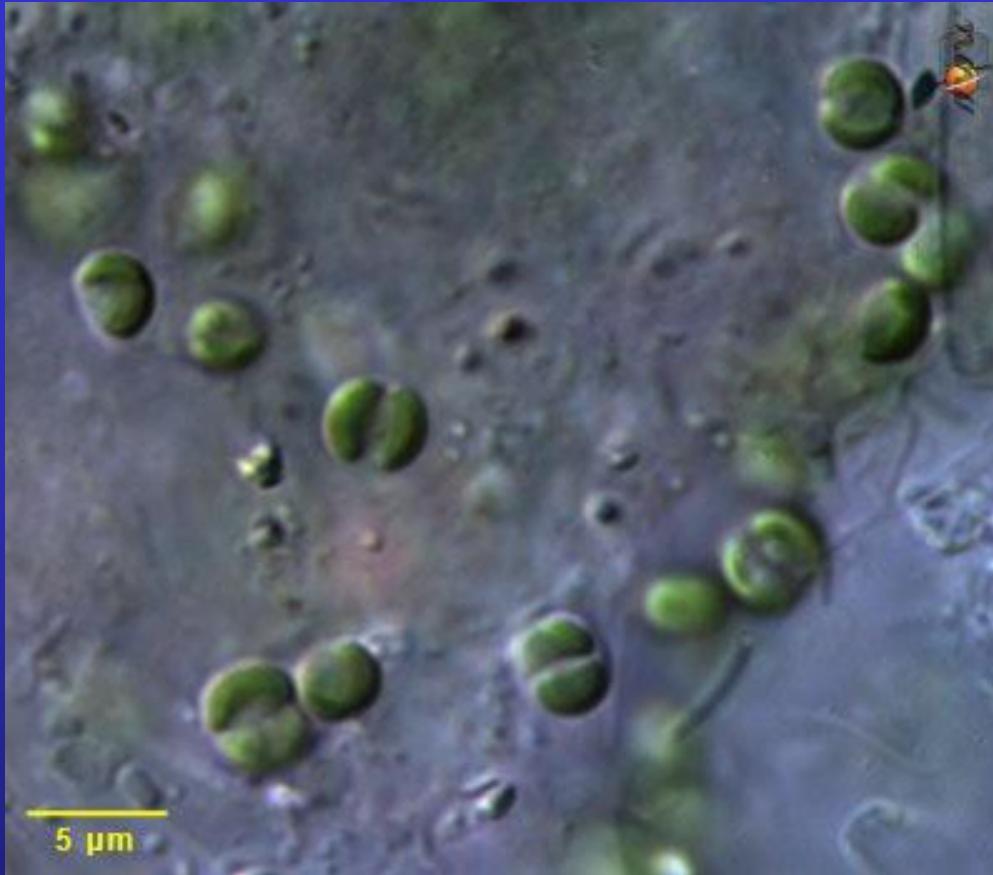




Yang et al. 2023

<https://doi.org/10.1038/s41467-023-41137-5>

Picocystophyceae - *Picocystis salinarum* halophytic



Mono Lake California

Layered cell wall
containing polyarabinose,
mannose, galactose and
glucose



Krienitz et al. 2012

Picocystis salinarum
(Chlorophyta) in saline lakes
and hot springs of East Africa



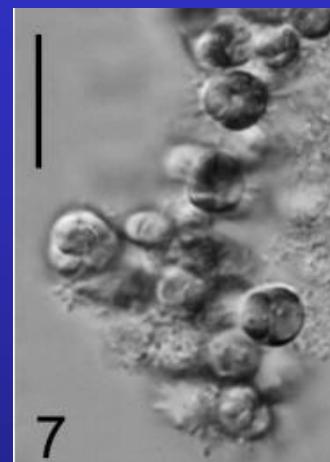
1



2



3



7



Picocystis occasionally replaces the dominant cyanobacterium (*Limnospira fusiformis*), which is the main food resource of Lesser Flamingos, in soda lakes of Bogoria and Nakuru

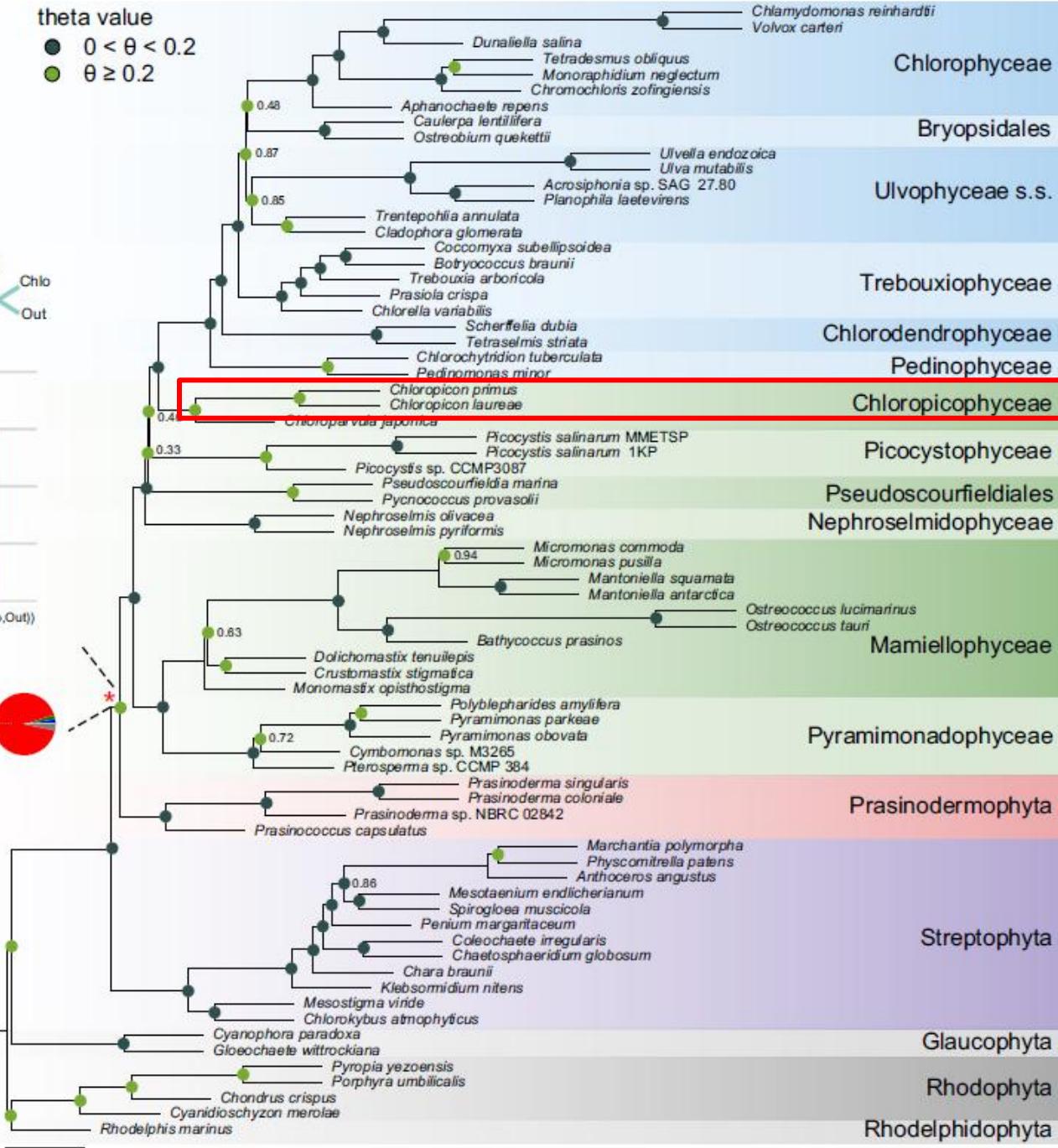
Východní Afrika – alkalická „soda lakes“



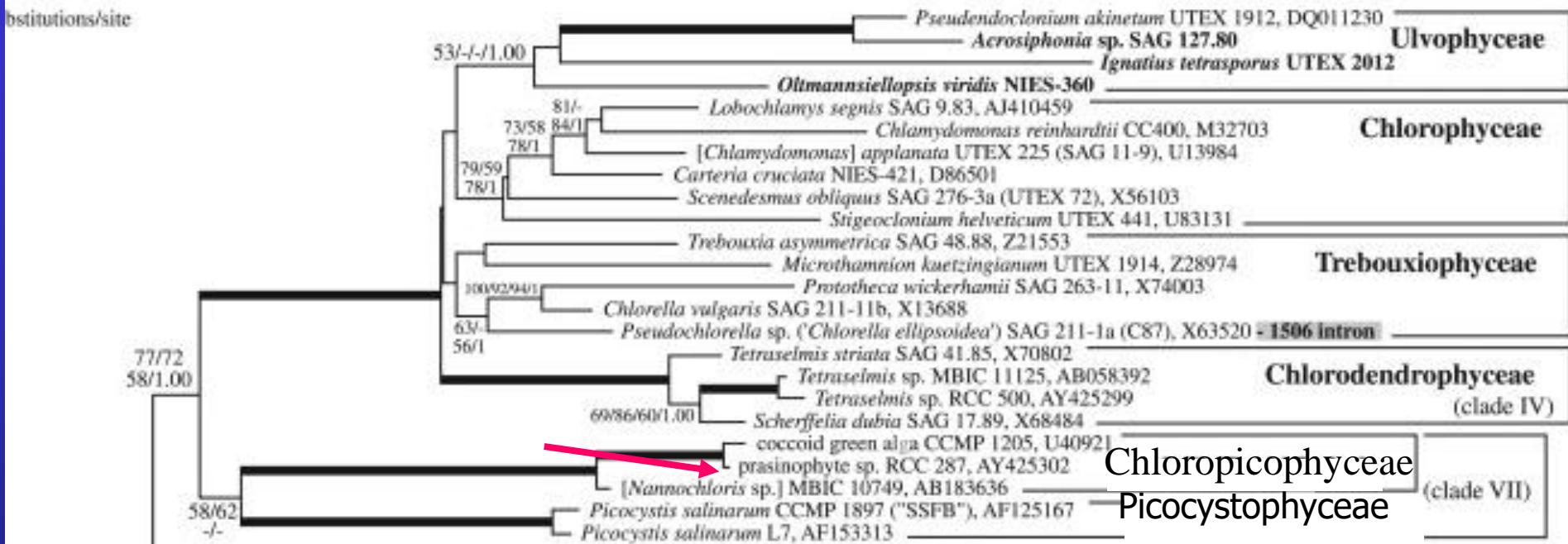
např. jezero Bogoria

b

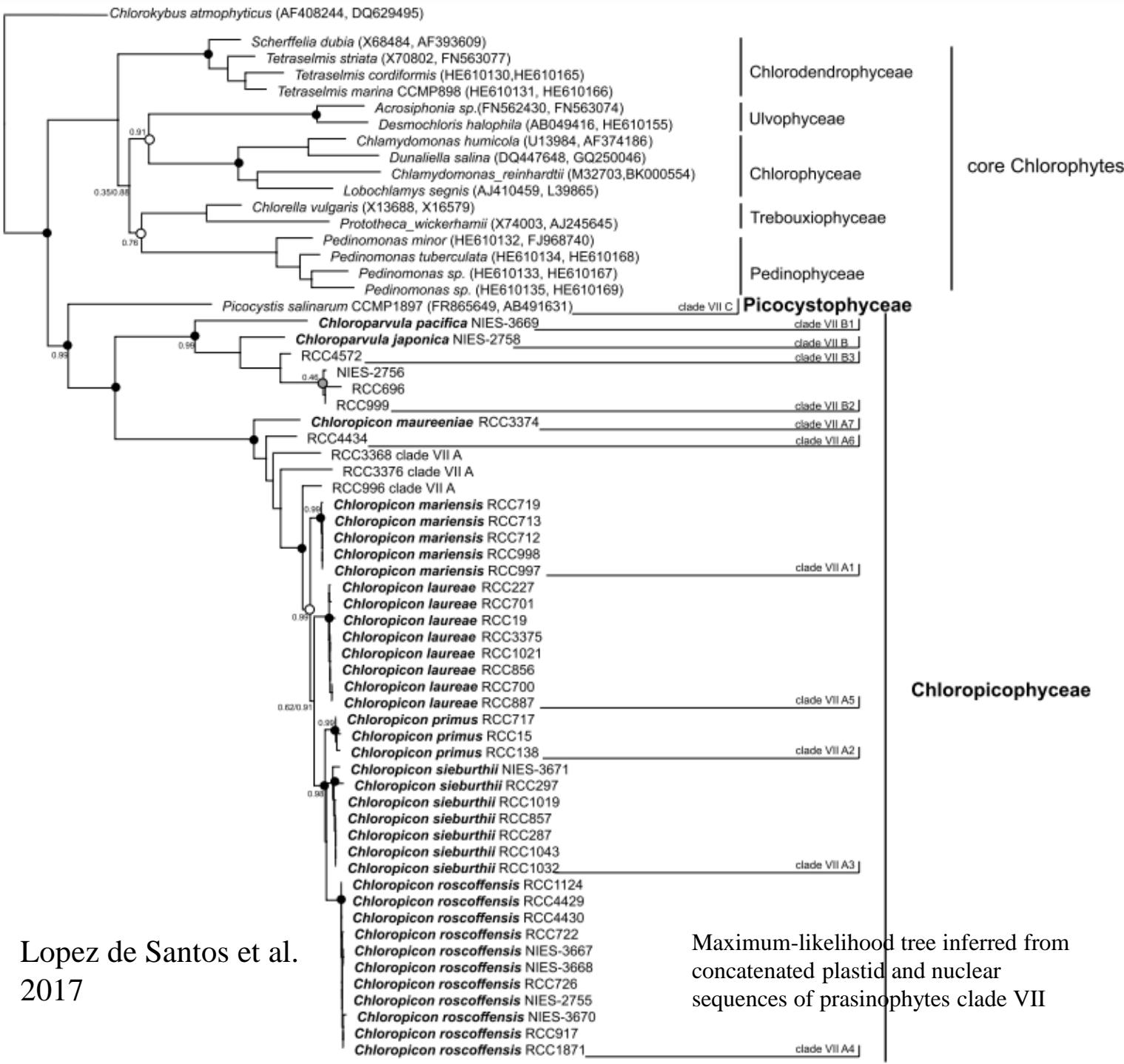
theta value
 ● $0 < \theta < 0.2$
 ● $\theta \geq 0.2$



Chloropicophyceae



Chloropicon sieburthii
a diameter of 1.5–4 µm,
found in oligotrophic marine
waters



Lopez de Santos et al.
2017

Maximum-likelihood tree inferred from concatenated plastid and nuclear sequences of prasinophytes clade VII

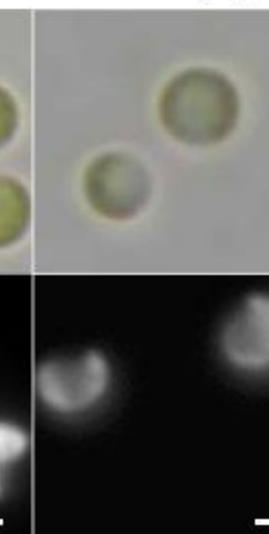
C. sieburthii
RCC287 (A3)



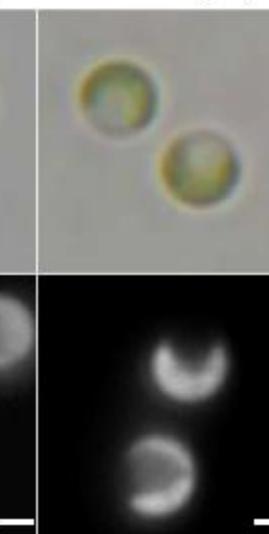
C. primus
RCC15 (A2)



C. laureae
RCC856 (A5)



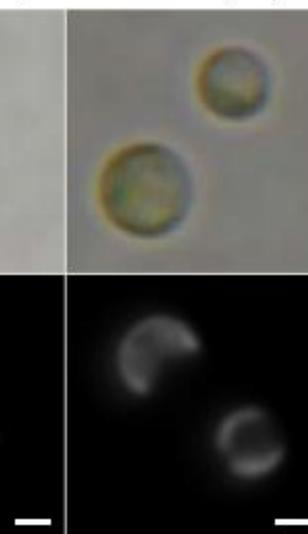
C. mariensis
RCC998 (A1)



C. roscoffensis
RCC1871 (A4)



C. maureeniae
RCC3374 (A7)

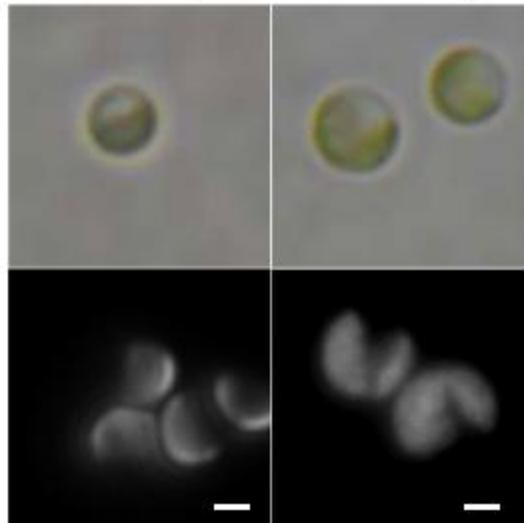


A

B

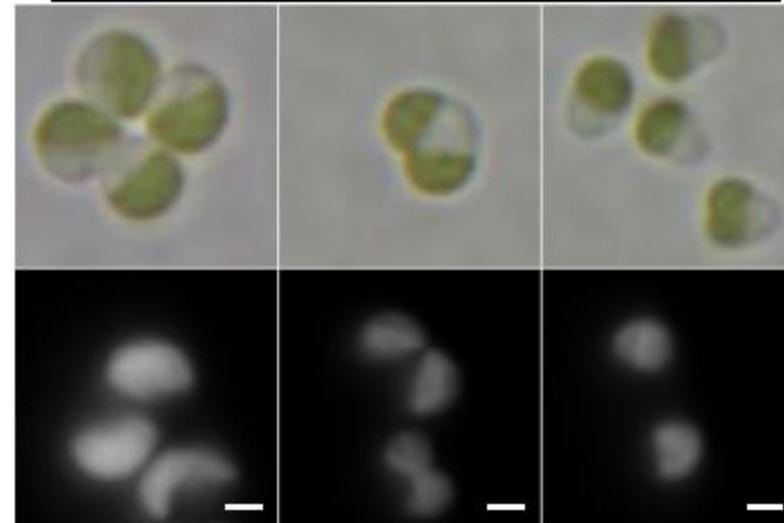
Chloropicon

C. pacifica
NIES-3669 (B1)



C. japonica
NIES-2758 (B)

P. salinarum
RCC3402



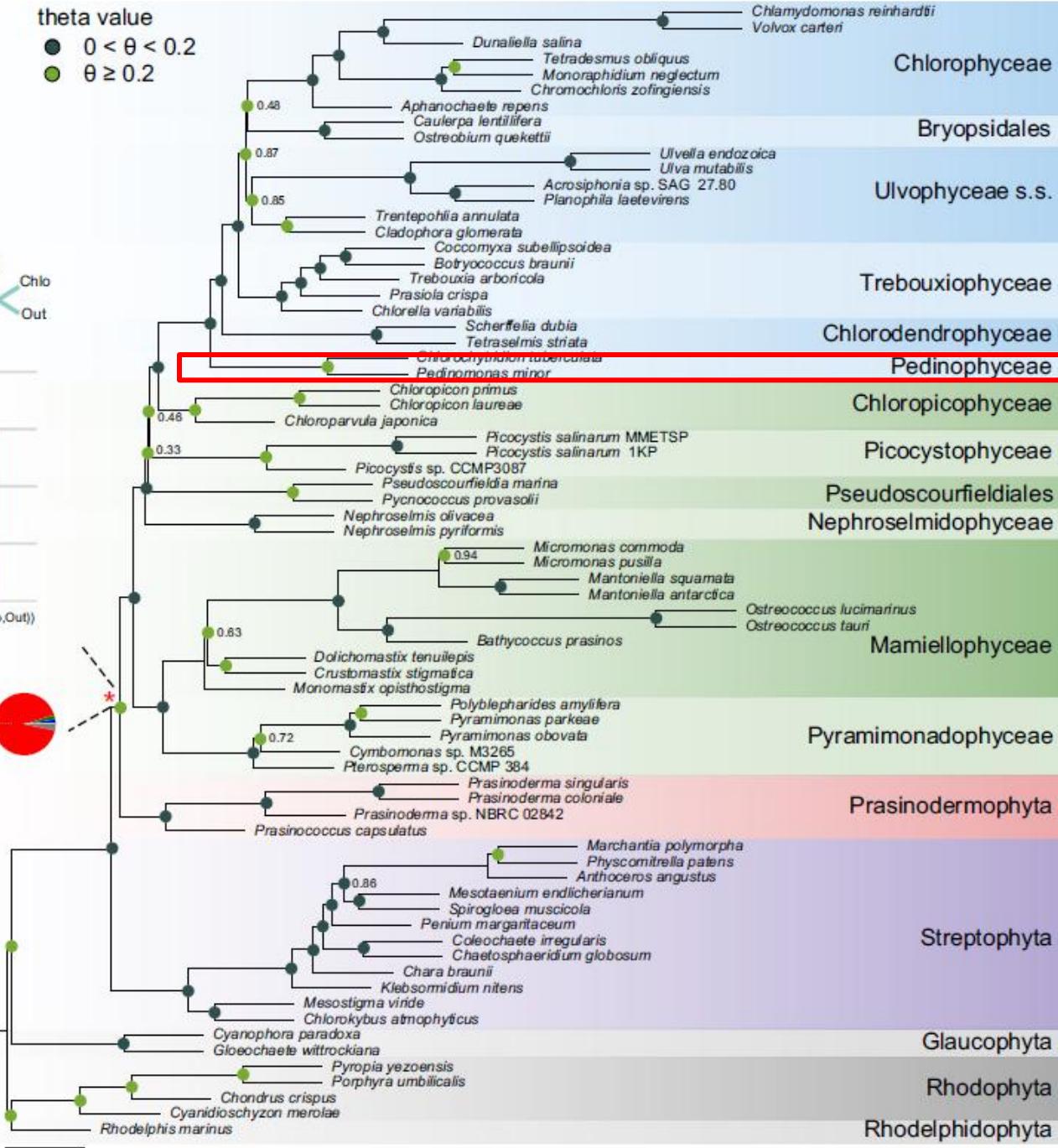
A

B

Core chlorophytes:
Ulvophyceae/Chlorophyceae/Trebouxiophyceae/Chlorodendrophyceae/
Pedinophyceae

b

theta value
 ● $0 < \theta < 0.2$
 ● $\theta \geq 0.2$



Yang et al. 2023

<https://doi.org/10.1038/s41467-023-41137-5>

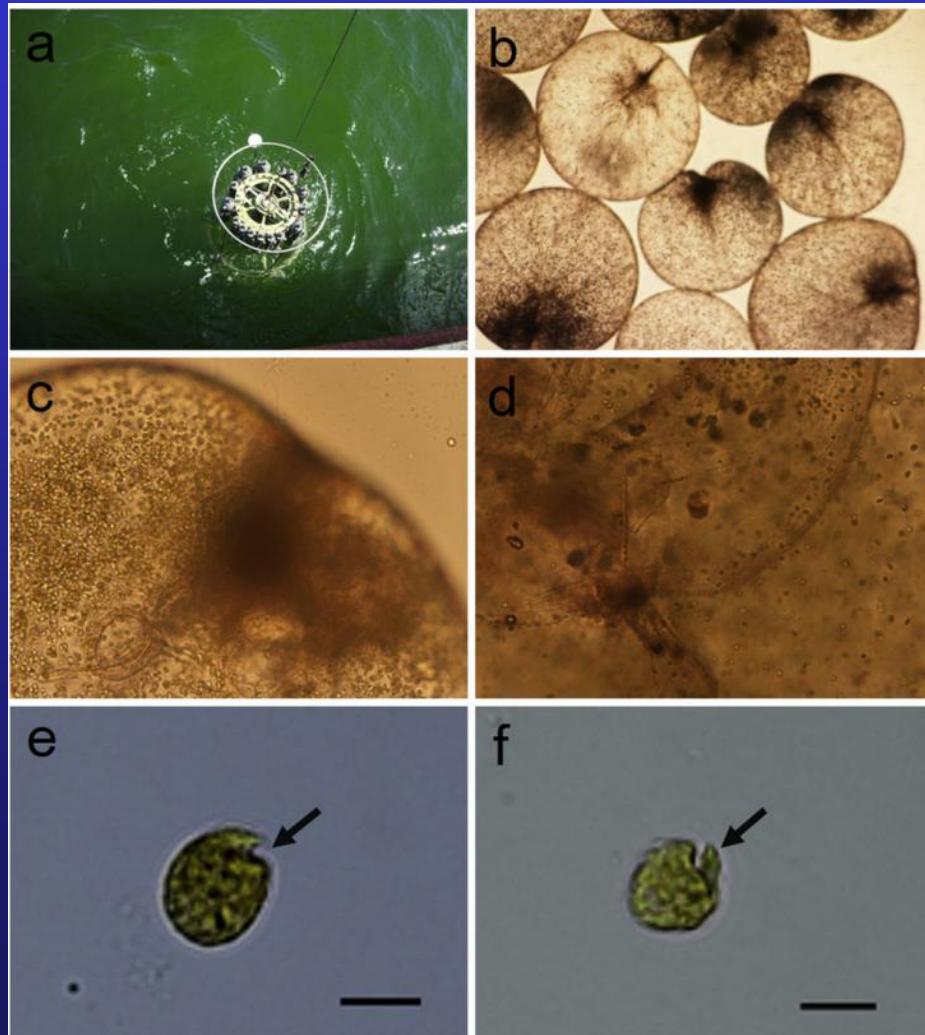
Pedinophyceae

Marsupiomonadales – live in seas, in salt or brackish waters, and are part of plankton. The second lineage – Pedinomonadales – is characterized by freshwater species or species living in soil

2.5 - 10 μm

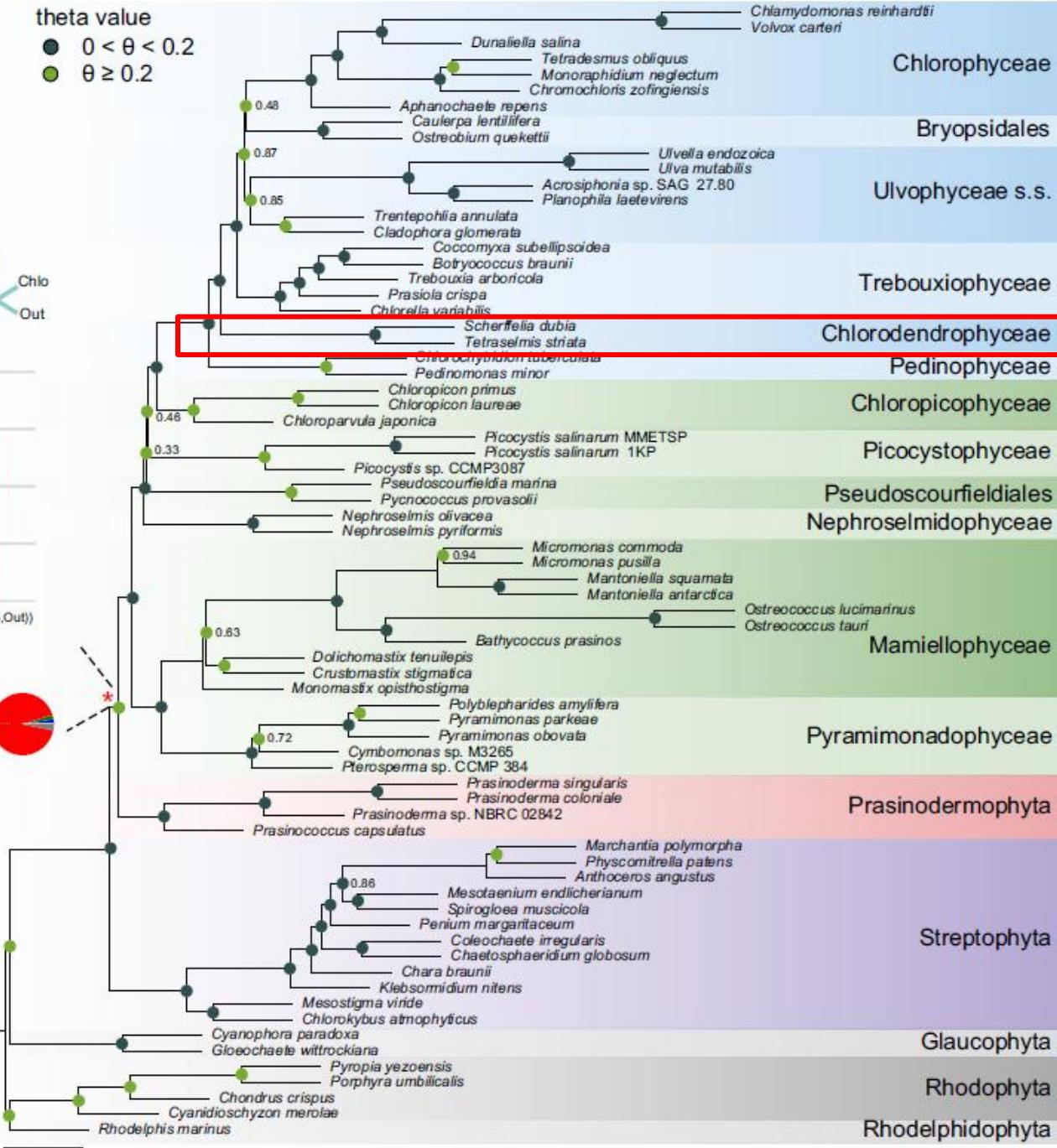
green *Noctiluca* contains a large number of endosymbiotic algal cells – blooms in the northern Arabian Sea

Pedinomonas noctilucae

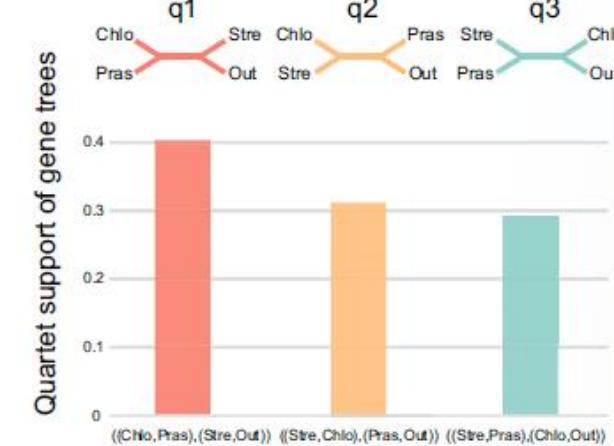


b

theta value
 ● $0 < \theta < 0.2$
 ● $\theta \geq 0.2$



c



Yang et al. 2023

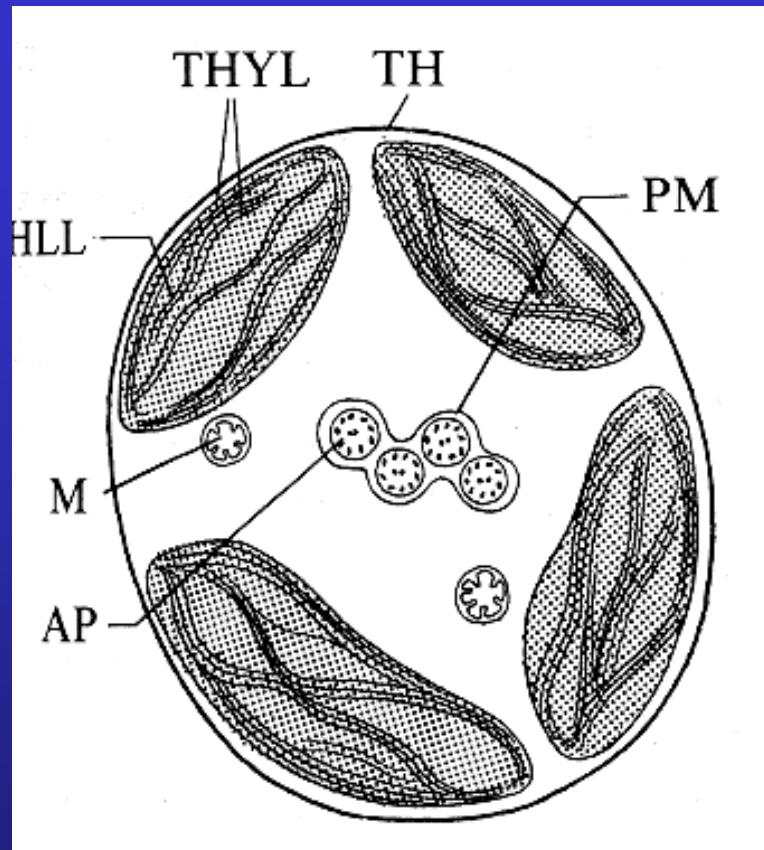
<https://doi.org/10.1038/s41467-023-41137-5>

Chlorodendrophyceae

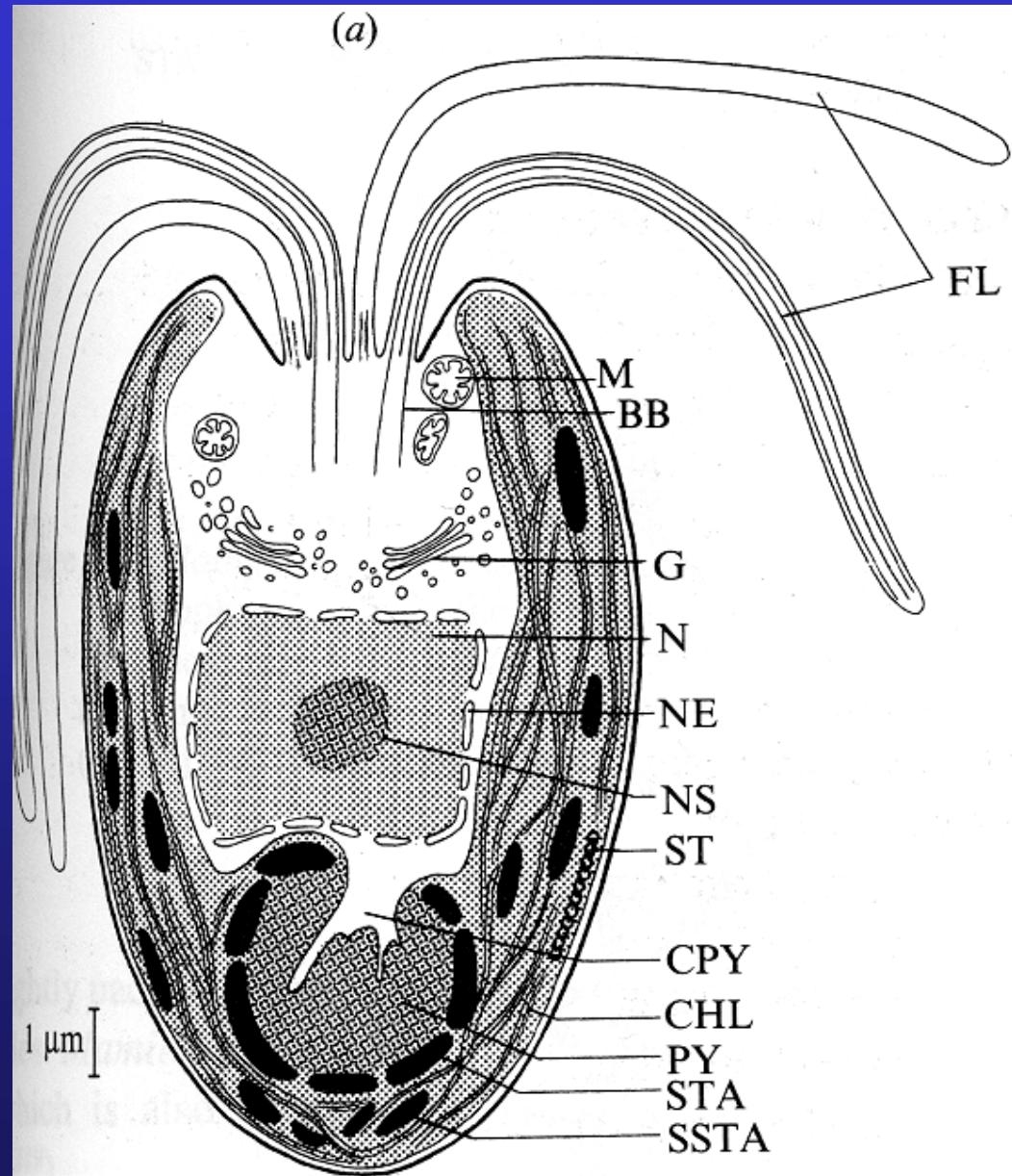


Tetraselmis

Chlorodendrophyceae

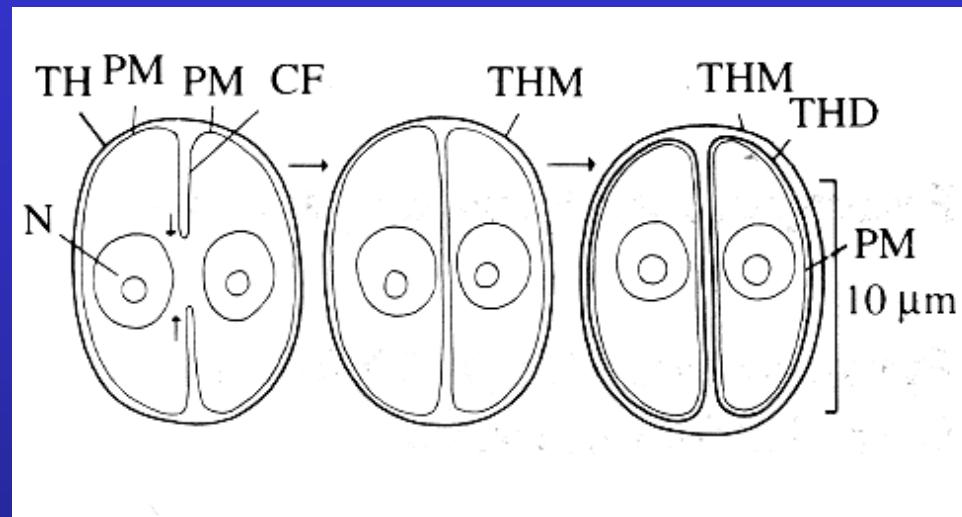
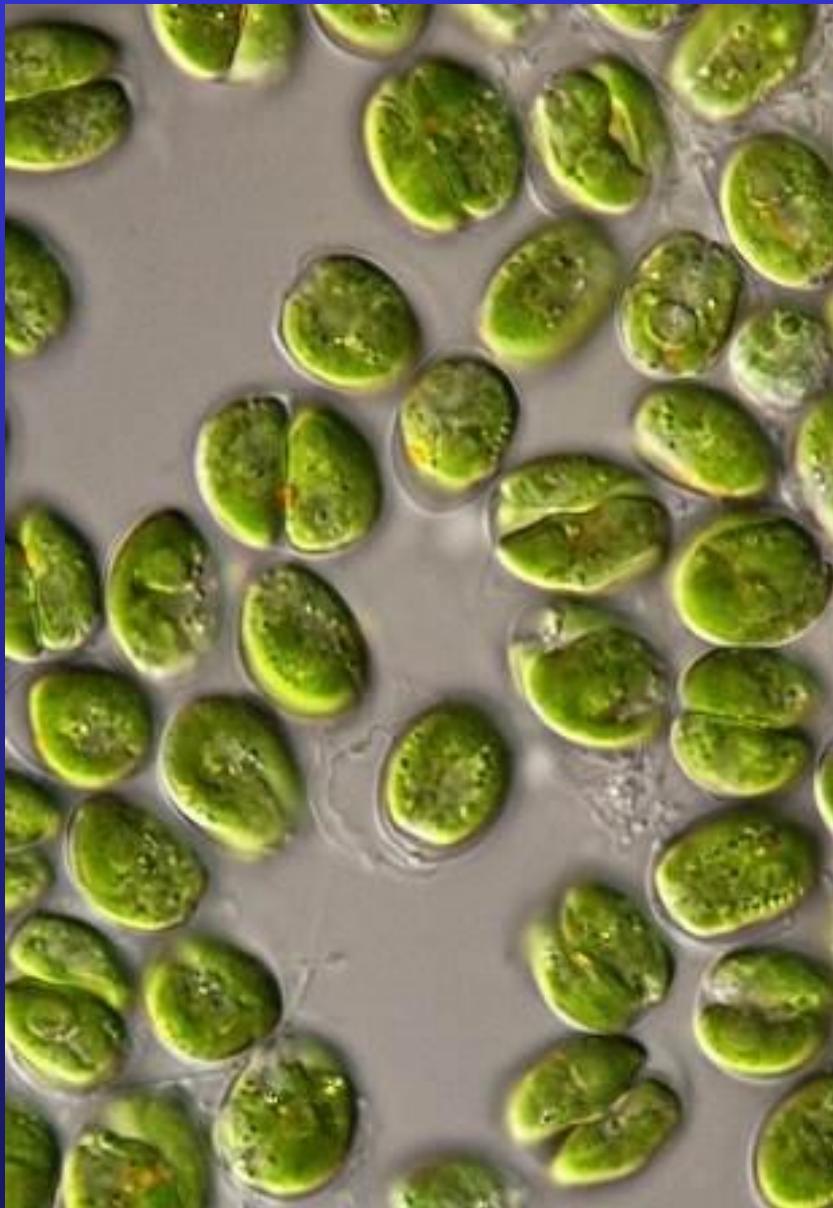


Tetraselmis suecica



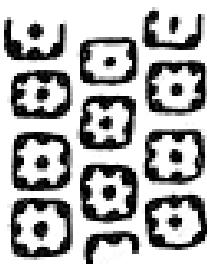
Mitóza a cytokineze

- na povrchu kryté thékou

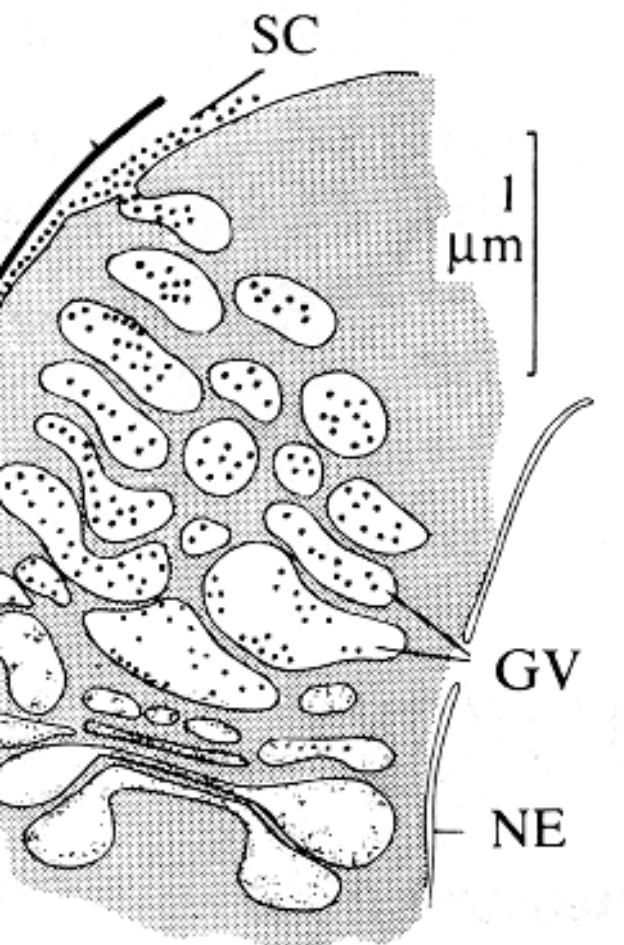


Tetraselmis suecica

shed flagella - division within the mother theca



0.1 μm

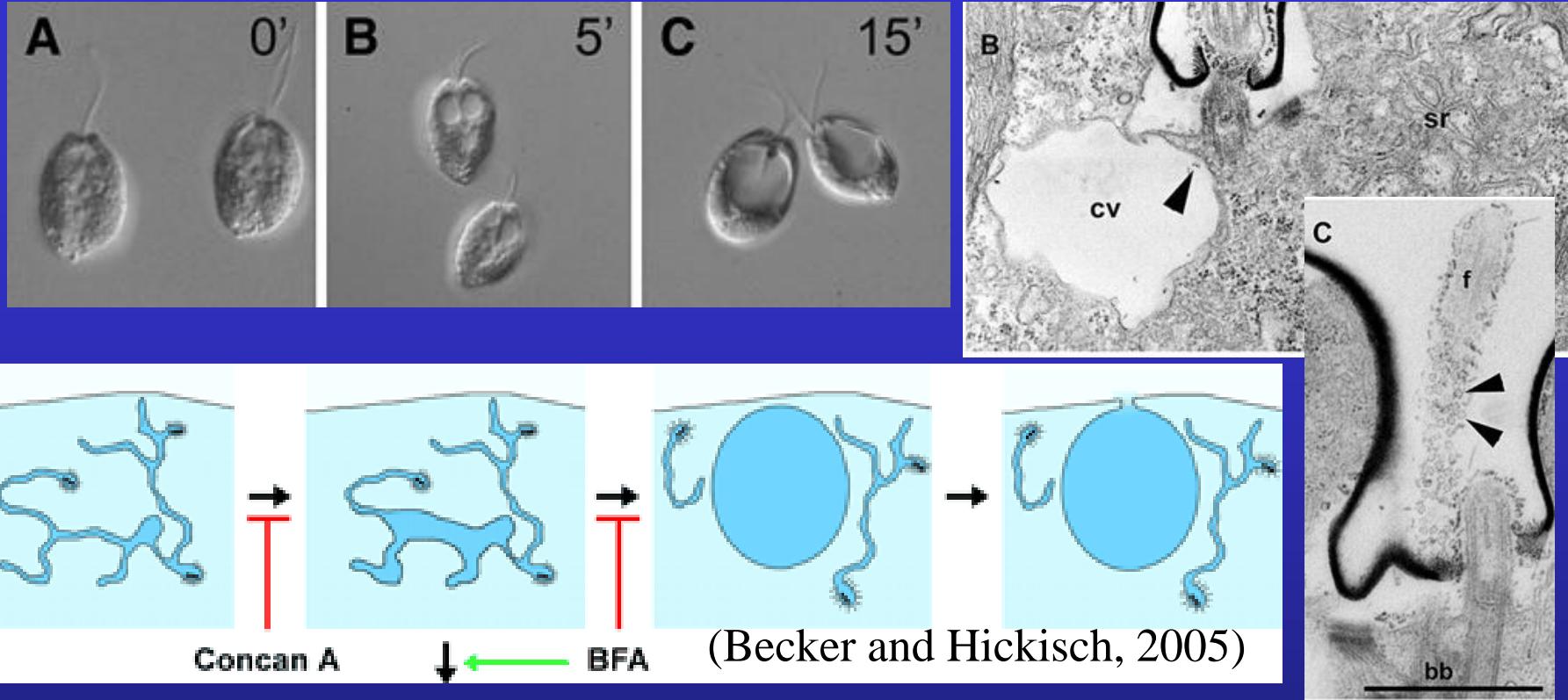


Scales fuse to form organic theca Chlorodendophyceae



Tetraselmis

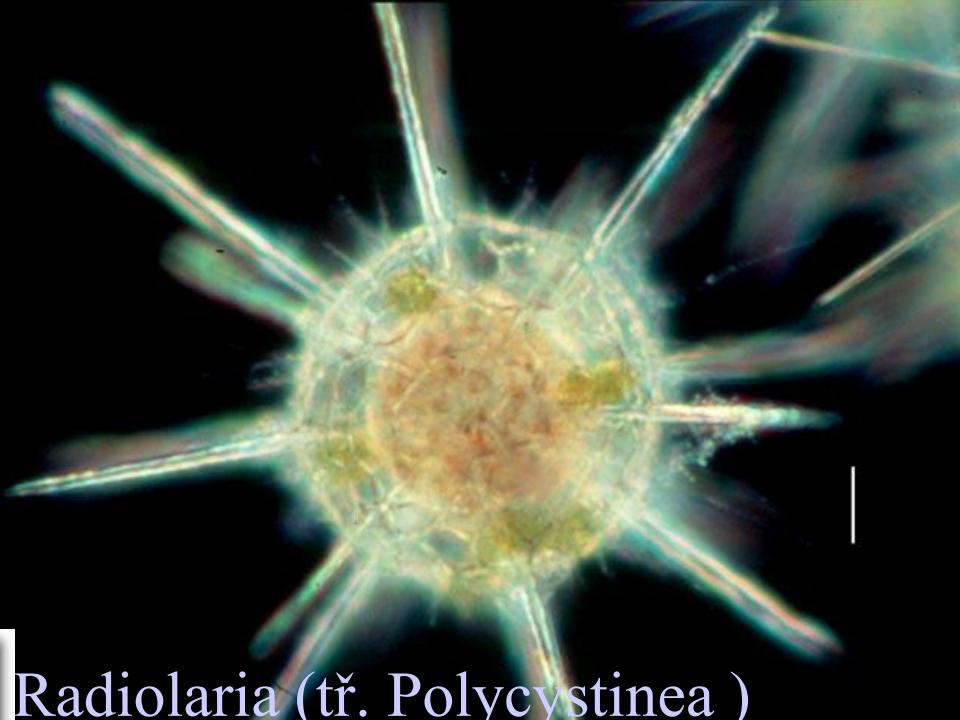
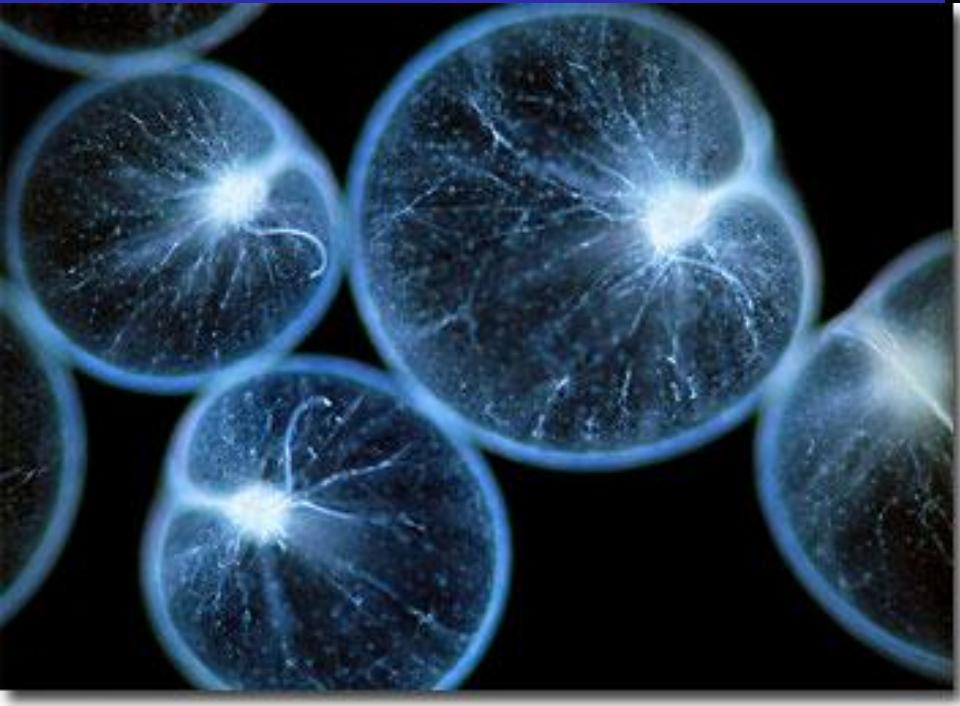
Contractile vacuoles



CV – *Scherffelia* - Osmoregulatory organelle – regularly removes water from the cell. Brefeldin – inhibits secretion, Golgi apparatus function, and the function of contractile vacuoles (blocks the fusion of the CV membrane with the plasma membrane). After Brefeldin application, a large central vacuole forms (representing the diastole phase) – the process is reversible; if the cell is placed in a hypertonic medium, the CV shrinks. An experimental model for studying diastole, essentially slowing down the entire process (otherwise 20ms)

Ekology

symbionts of different
heterotrophic organisms

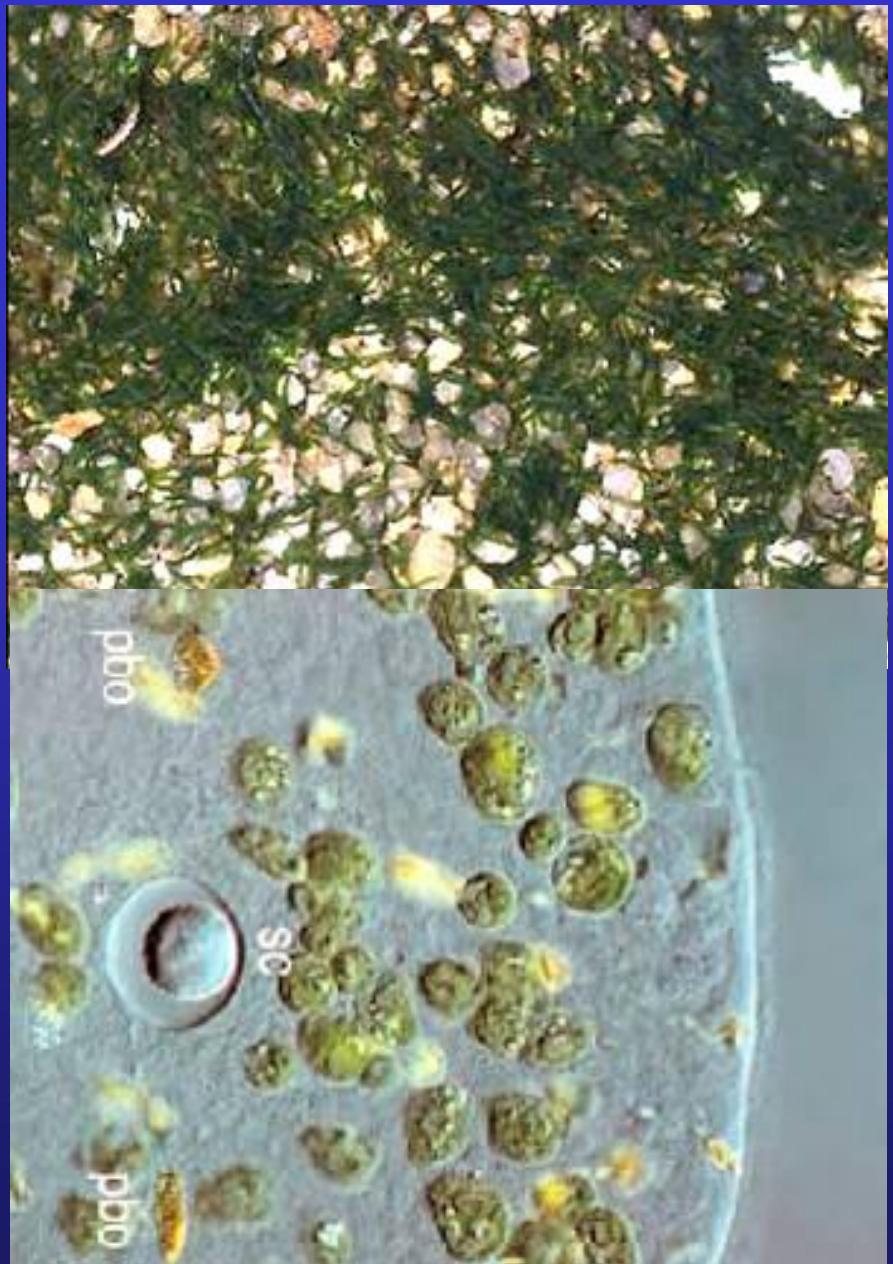


Radiolaria (tř. Polycystinea)

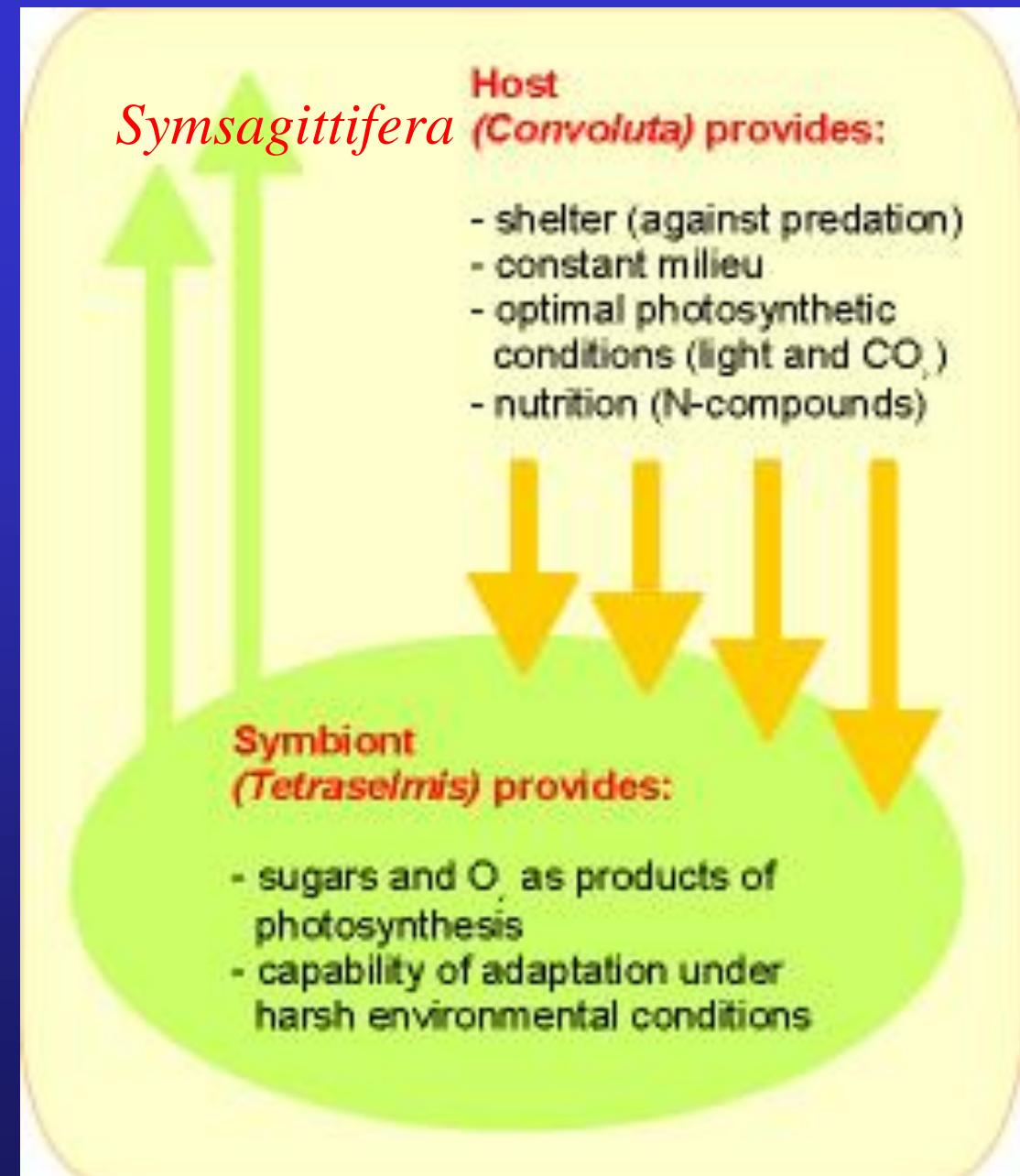
Noctiluca

Ekology

symbionts of different
heterotrophic organisms



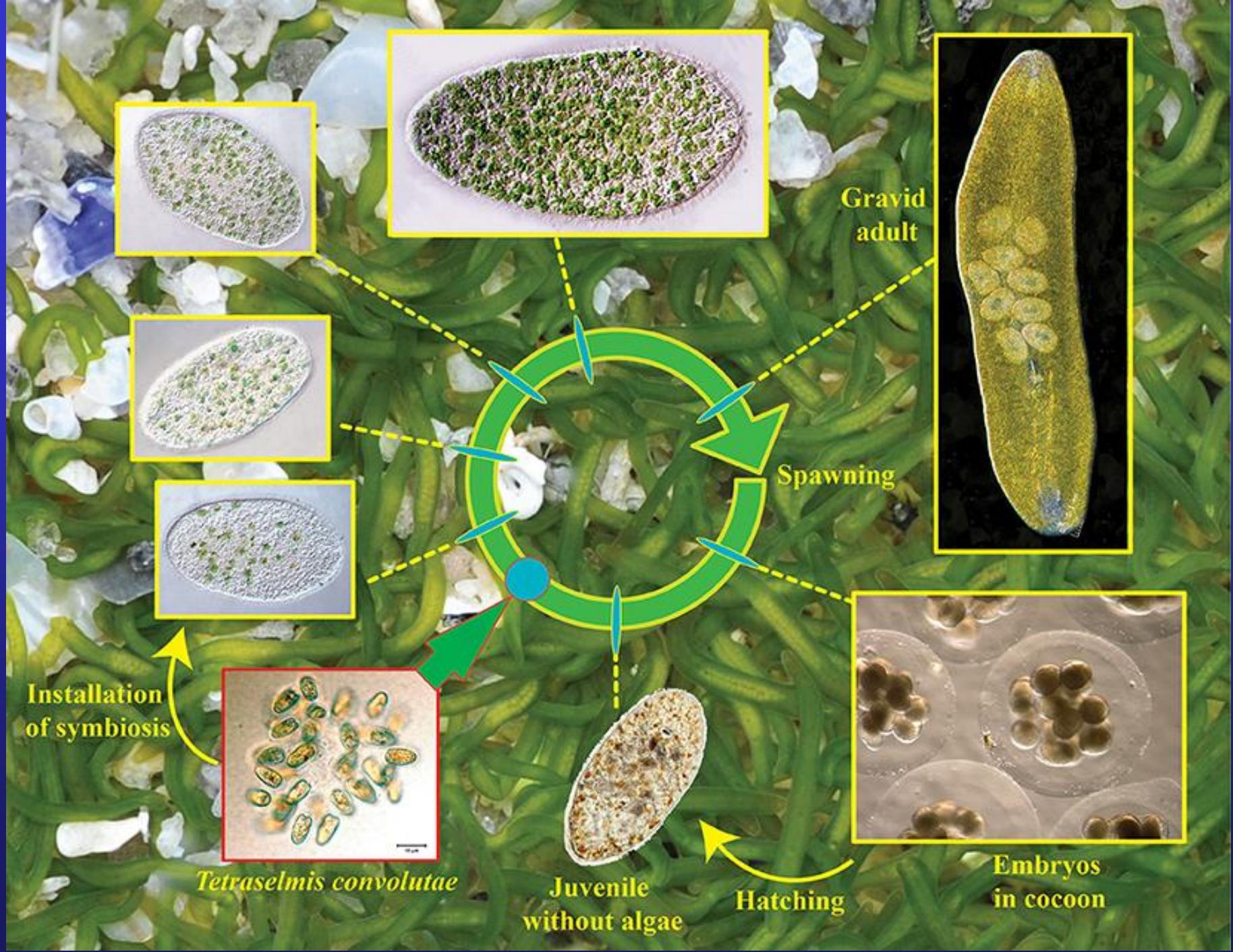
(*Convoluta*) *Symsagittifera roscoffensis* - flatworms Acoela—no mouth



Akvitaine - France

Atlantic ocean shore







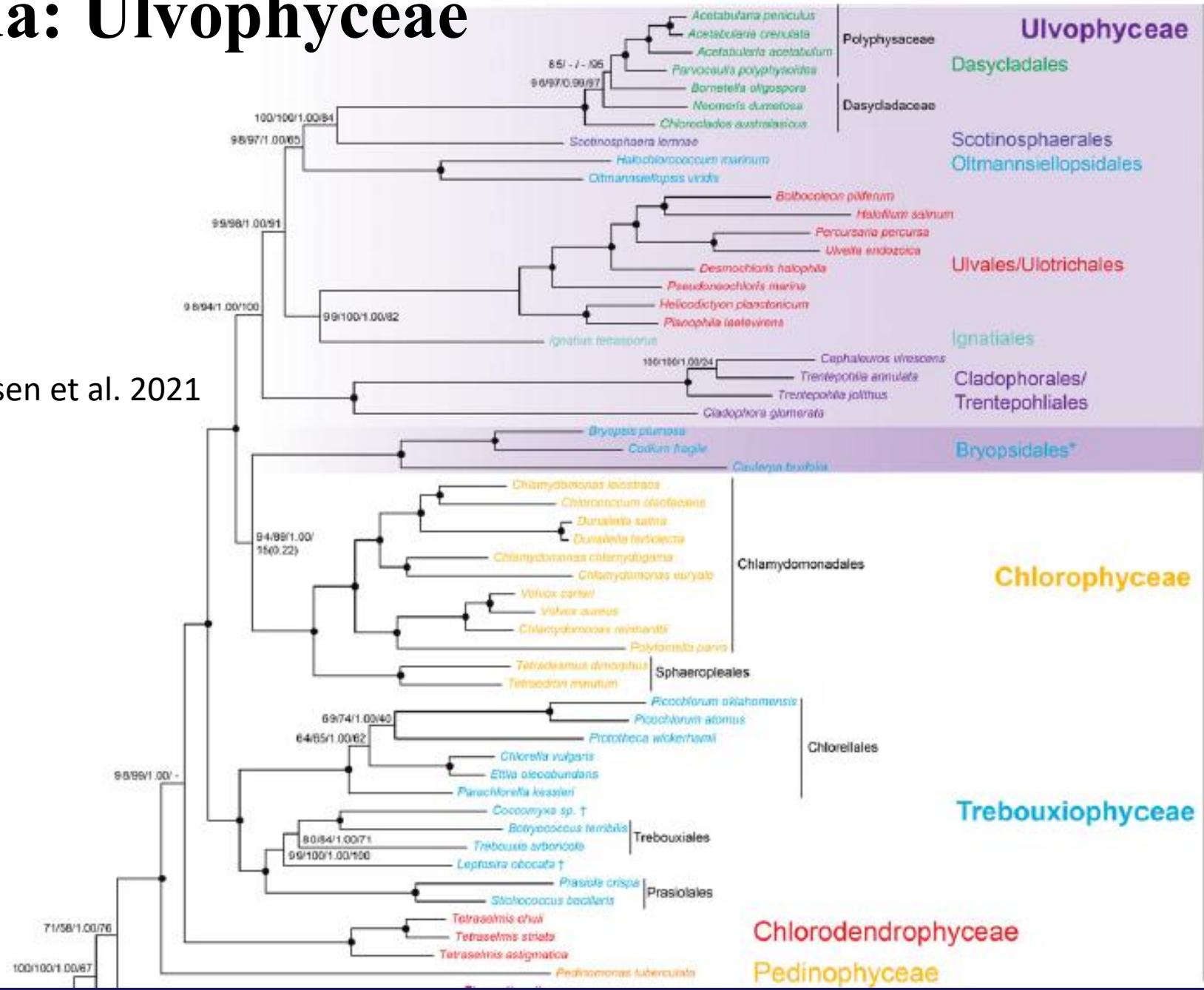
polyunsaturated
fatty acids

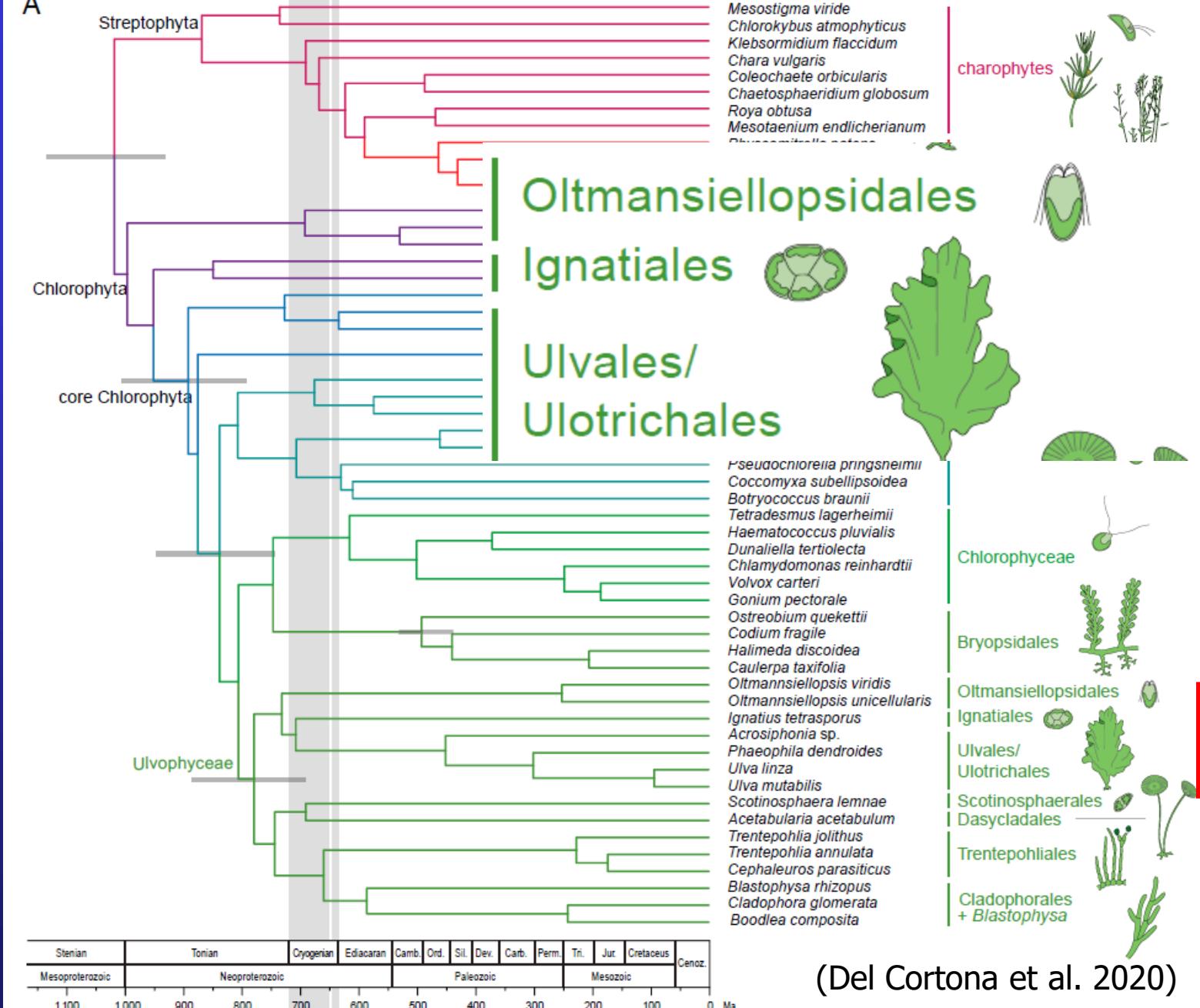


Live Foods for Feeding Aquarium Fish, Inverts & Corals

Třída: Ulvophyceae

Gulbrandsen et al. 2021

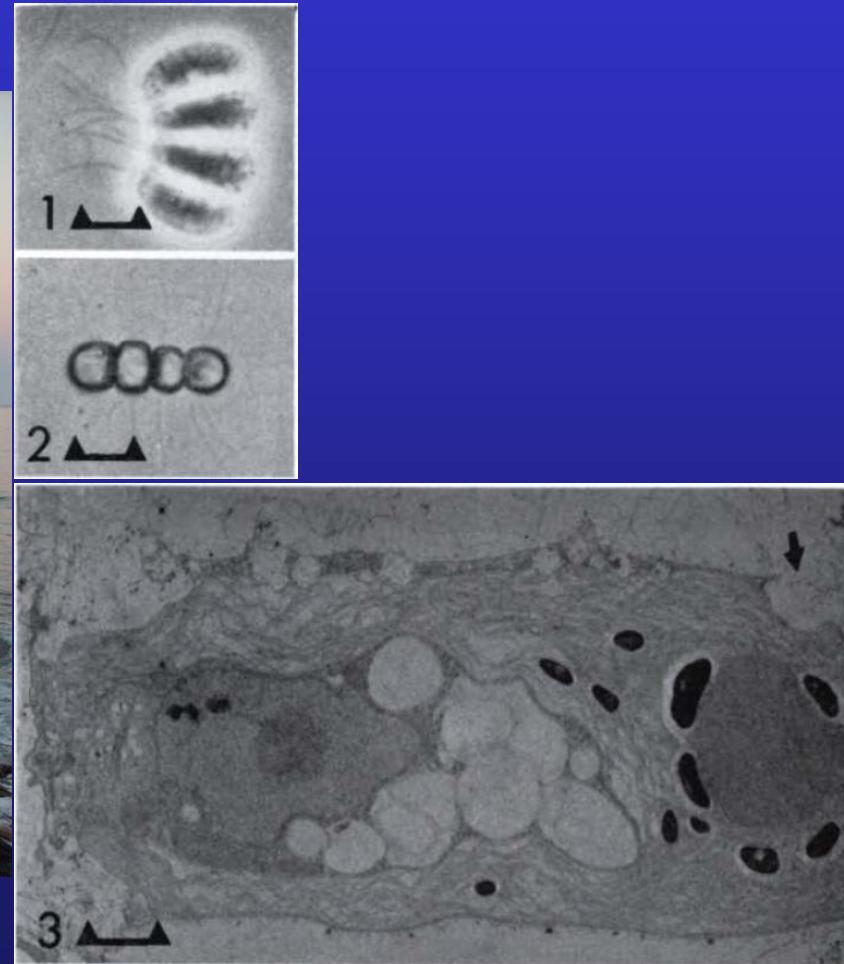
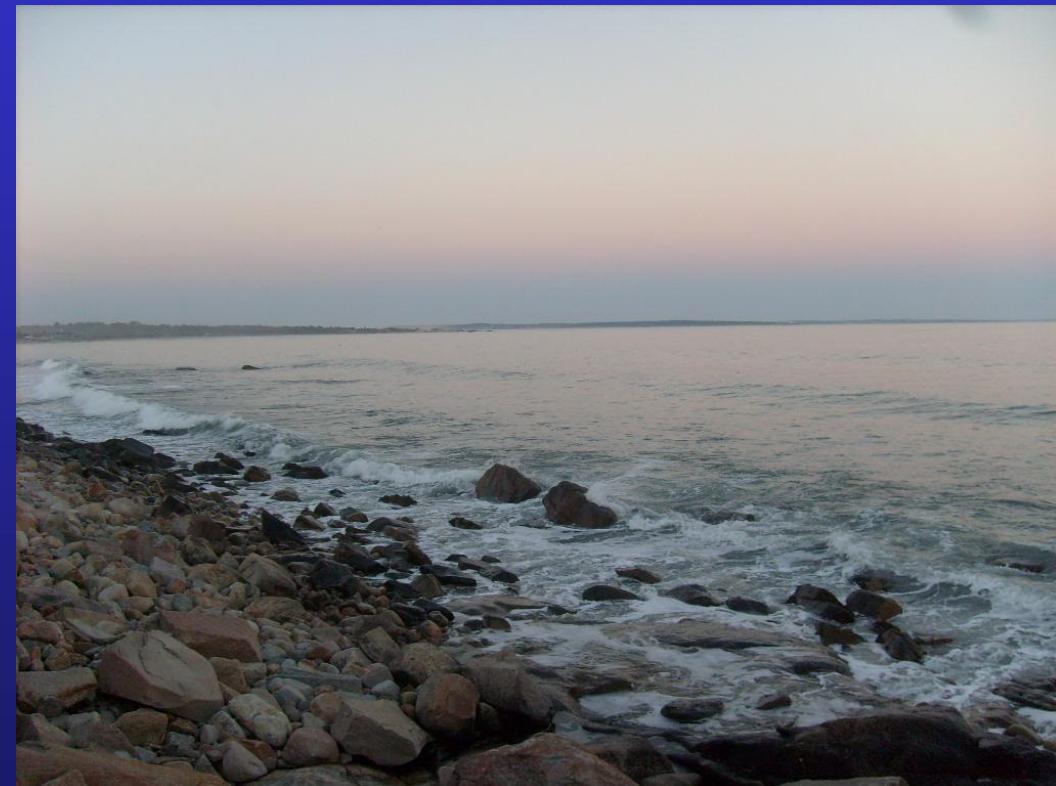




Time-calibrated phylogeny of the green algae. (A) The topology of the tree is based on the ML analysis inferred from a concatenated amino acid alignment of 539 nuclear genes

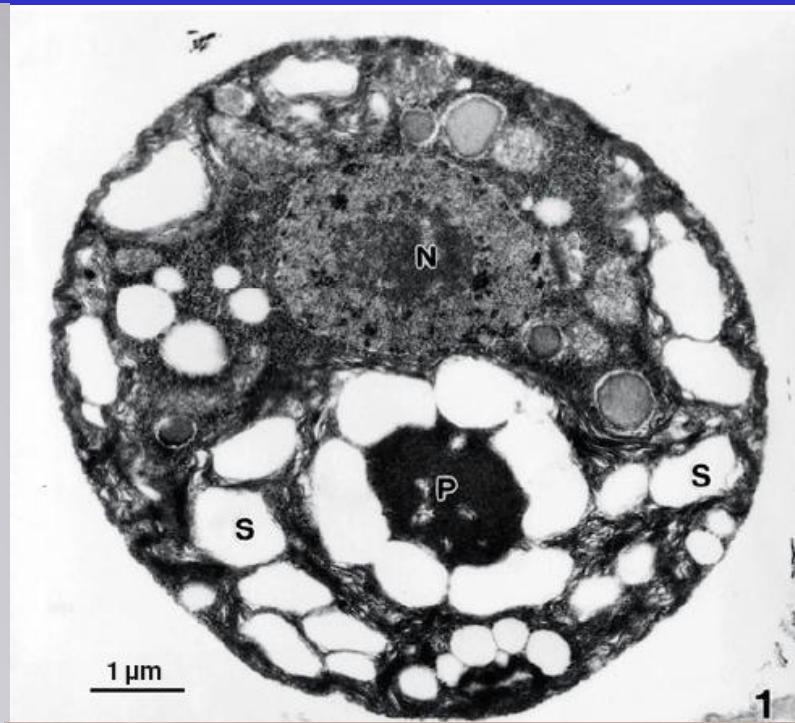
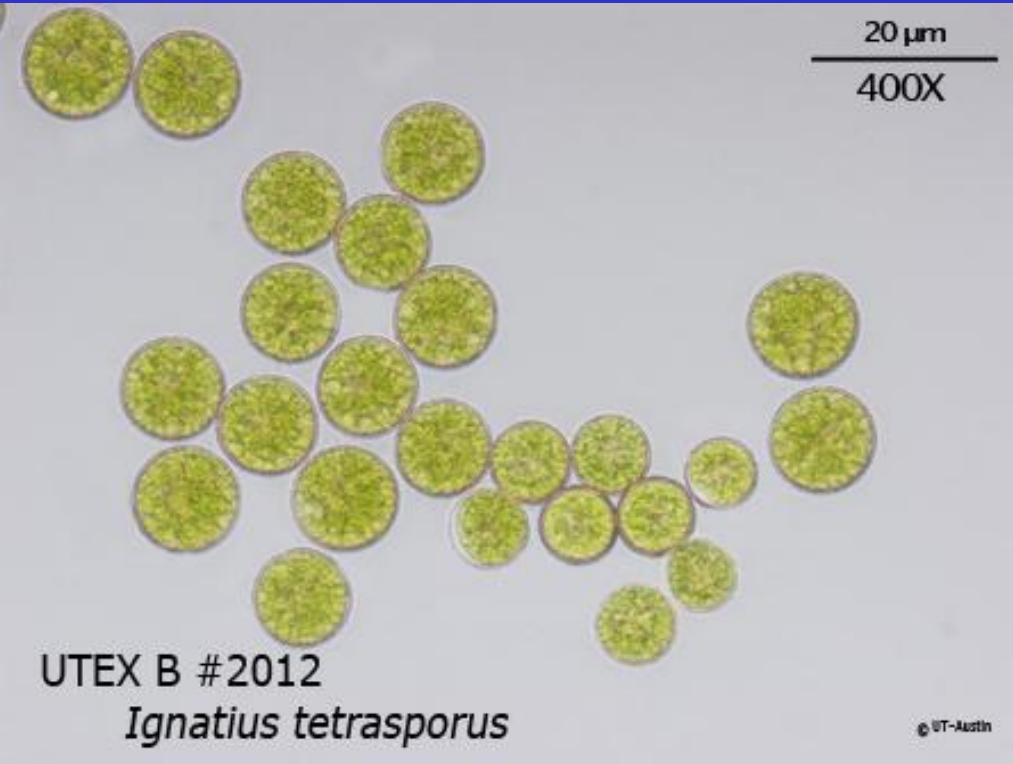
Oltmansiellopsidales

Oltmansiellopsis viridis -a quadriflagellate green alga is described from temperate coastal waters. It occurs primarily as four-celled colonies,



Narragansett Bay, Rhode Island,
USA

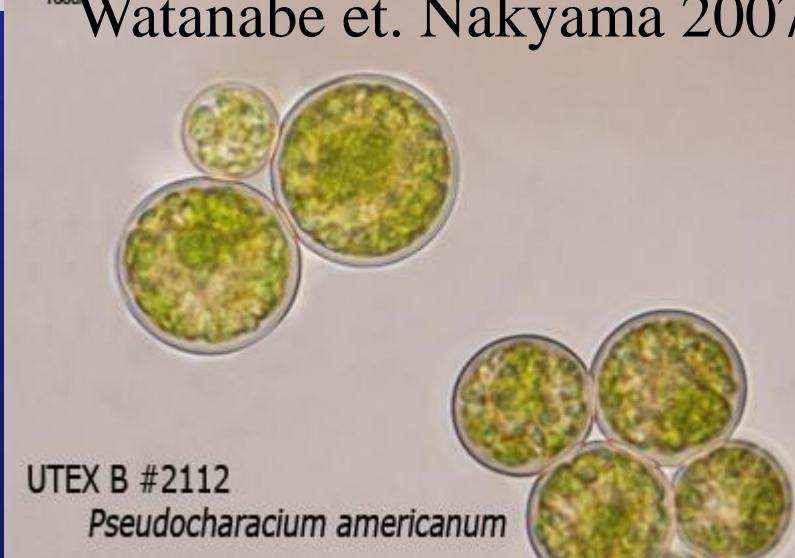
Igniales



Coccoids from damp terrestrial habitats

A young vegetative cell included a single nucleus and a cup-shaped chloroplast that contained a pyrenoid and stroma starch grains

Watanabe et. Nakyama 2007

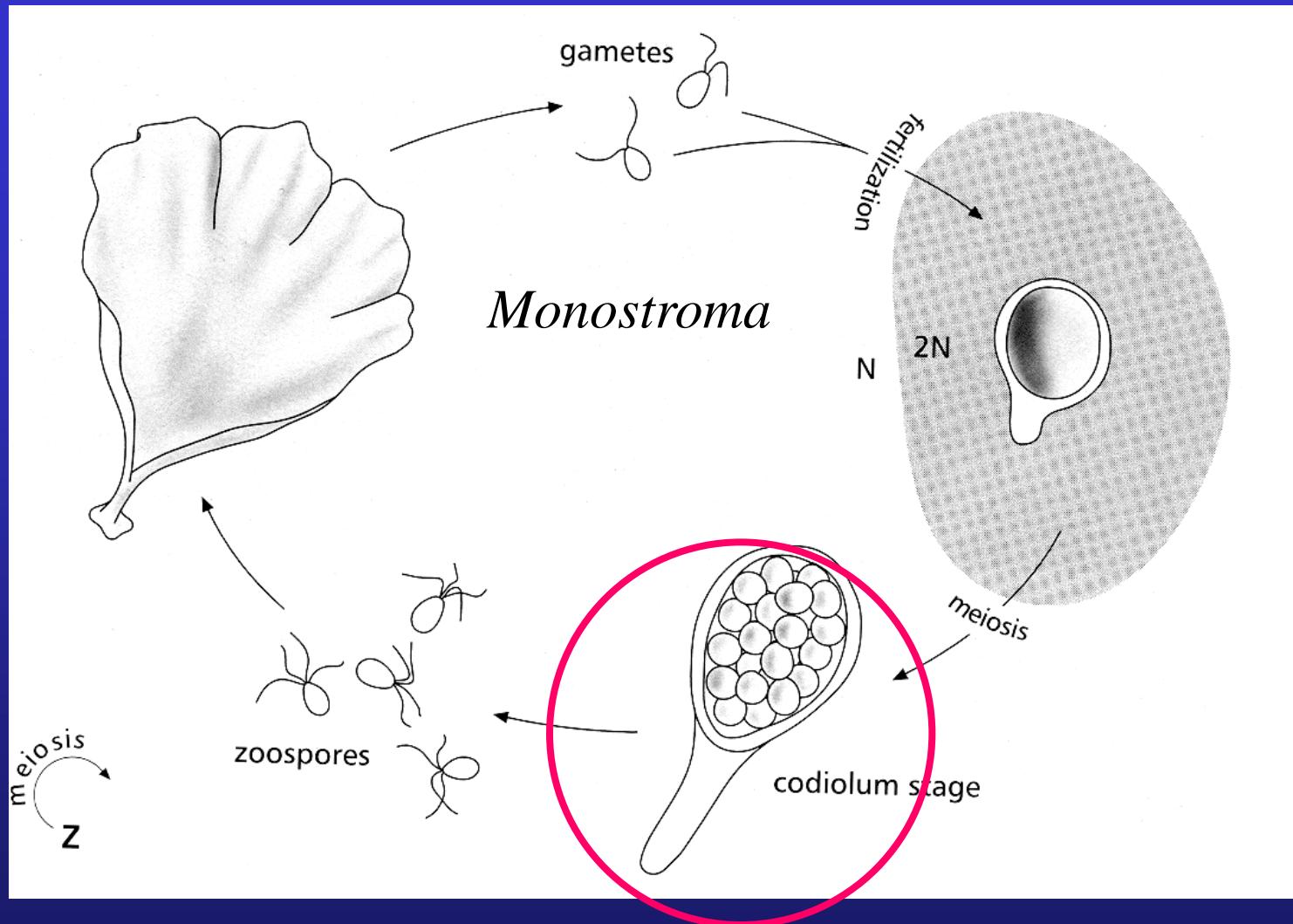


Ulvales/Ulotrichales

Basic Characteristics:

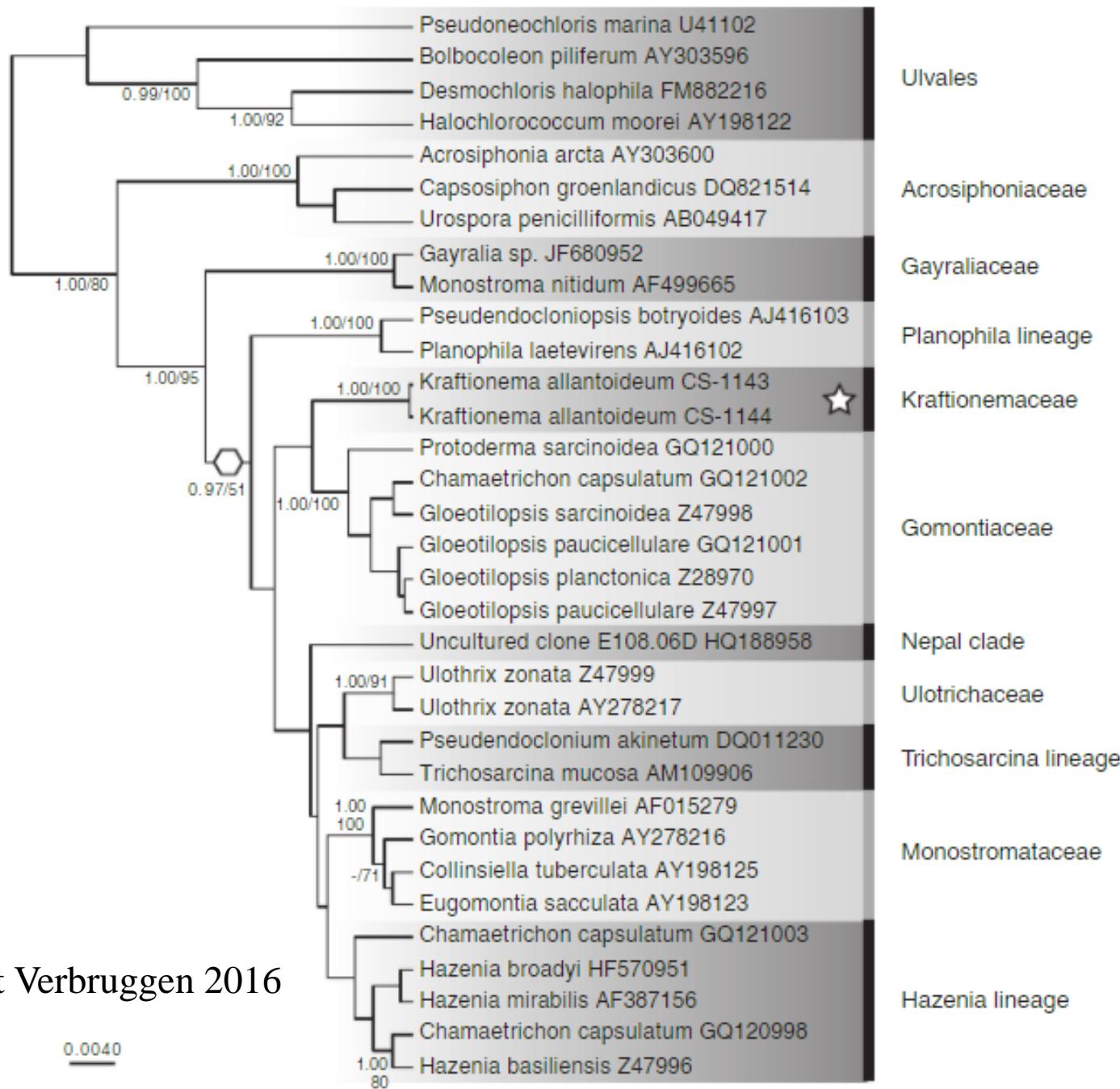
- Thallus can be coccoid, filamentous, siphonous, or parenchymatous.
- Zoids have 2 or 4 flagella (with a cruciate arrangement of flagellar roots and CCW orientation of basal bodies); some representatives of Ulotrichales have rhomboid scales covering the surface of zoids.
- Mitosis is closed, with a persistent telophase spindle (lacking phycoplast and fragmoplast).
- Each cell contains a parietal chloroplast (band-shaped or cup-shaped), sometimes perforated, with one or several pyrenoids.
- Haplontic or diplo-haplontic life cycle, with the formation of thick-walled hypnozygotes.
- Representatives of this class are almost exclusively marine organisms.

Řád: Ulotrichales (Codiolales)



- *Codiolum* stages can develop inside mussels or endophytically within the thalli of macroalgae encrusted with limestone, providing a protected and stable microclimate.
- Under favorable conditions, they germinate into four-flagellated zoospores (meiospores).

Phylogenetic tree of Ulotrichales inferred from 18S ribosomal RNA sequences



Wetherbee et Verbruggen 2016

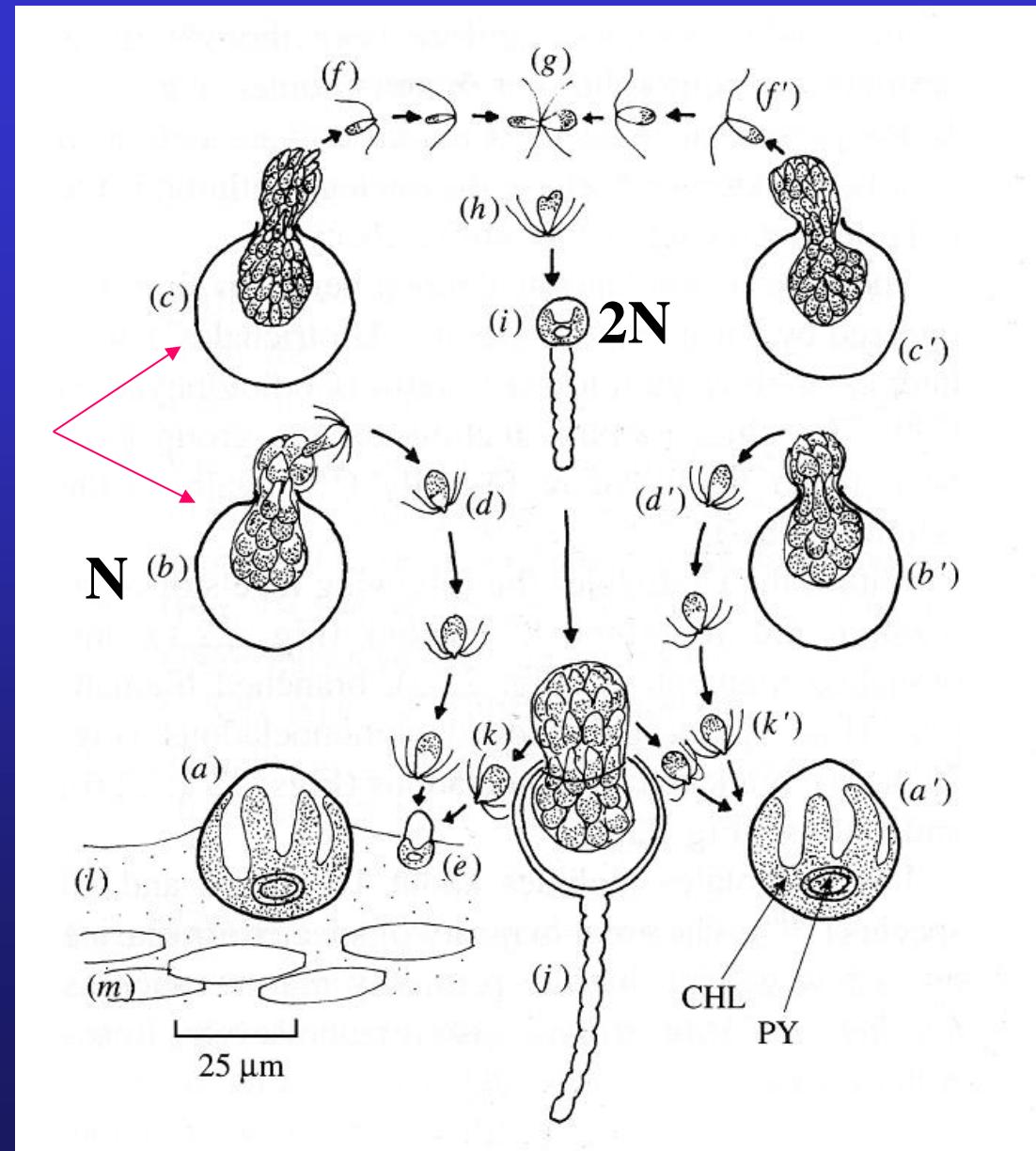
Chlorocystis

endophyte, haplontic life cycle, codiolus stage

anizogamy

heterothalism

evolutionary parallelism (*Trebouxia*, *Chlorococcum*)

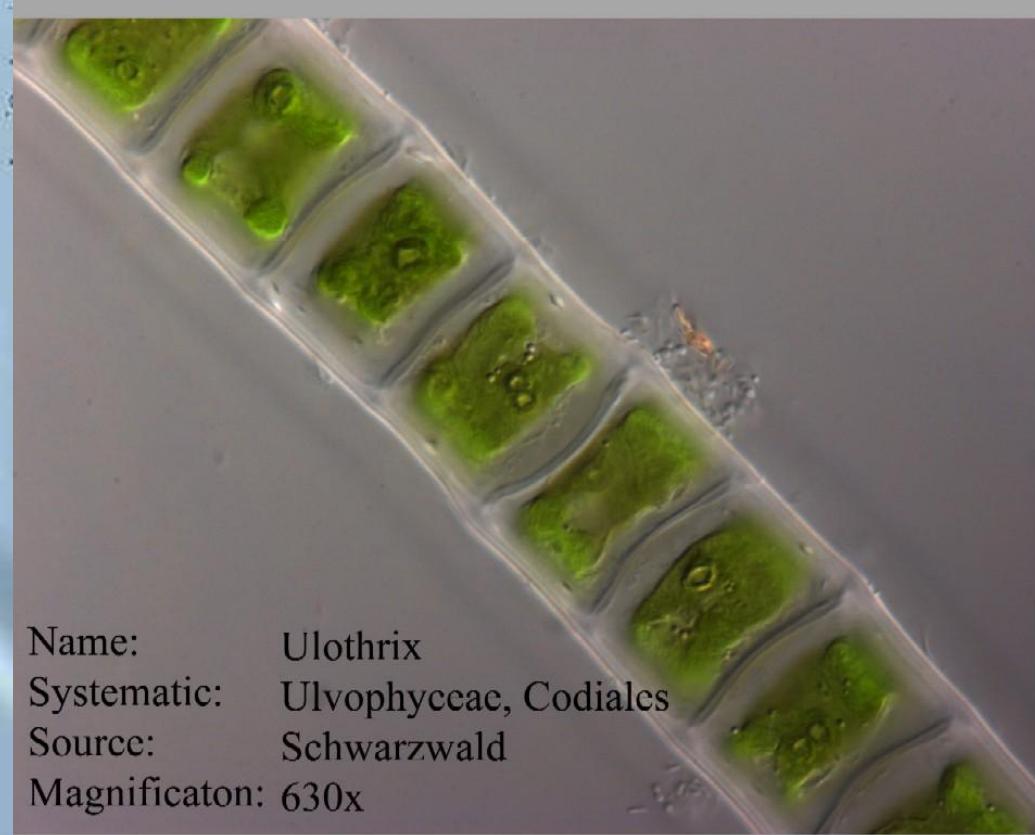




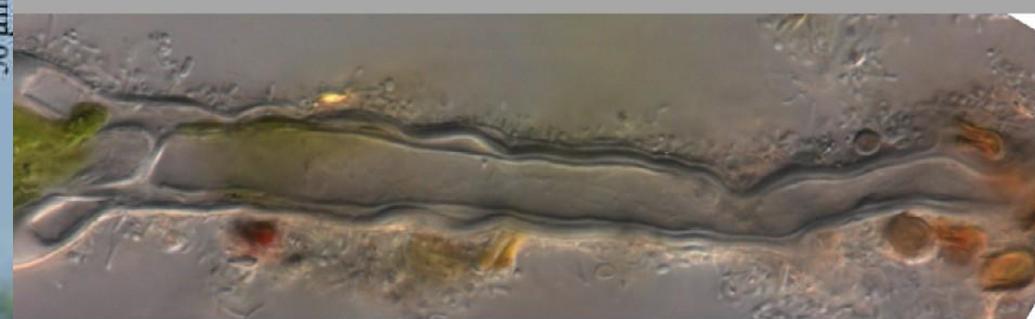
SINICE
&
ŘASY.cz



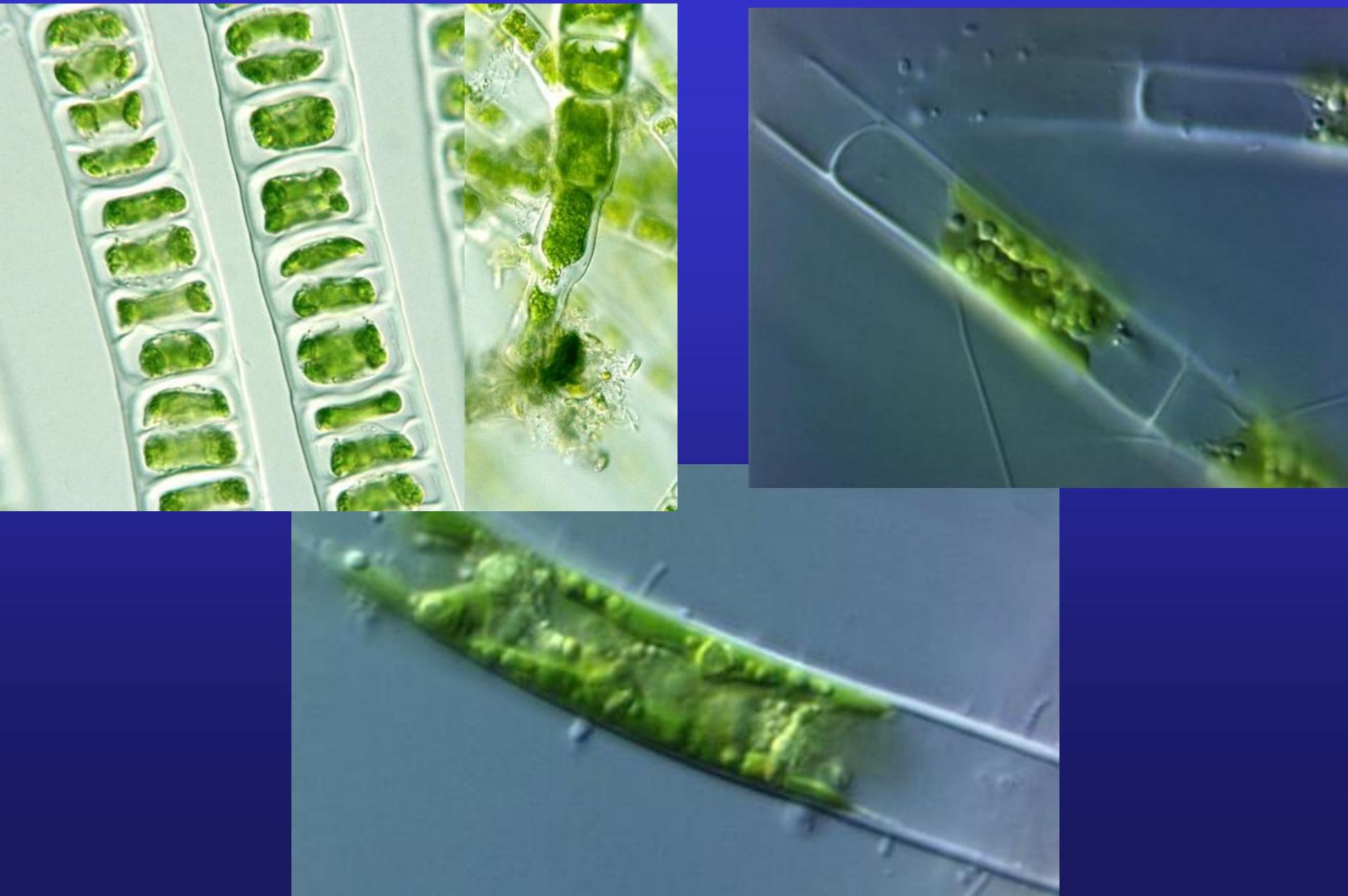
Ulothrix



Name: *Ulothrix*
Systematic: Ulvophyceae, Codiales
Source: Schwarzwald
Magnification: 630x



Ulothrix

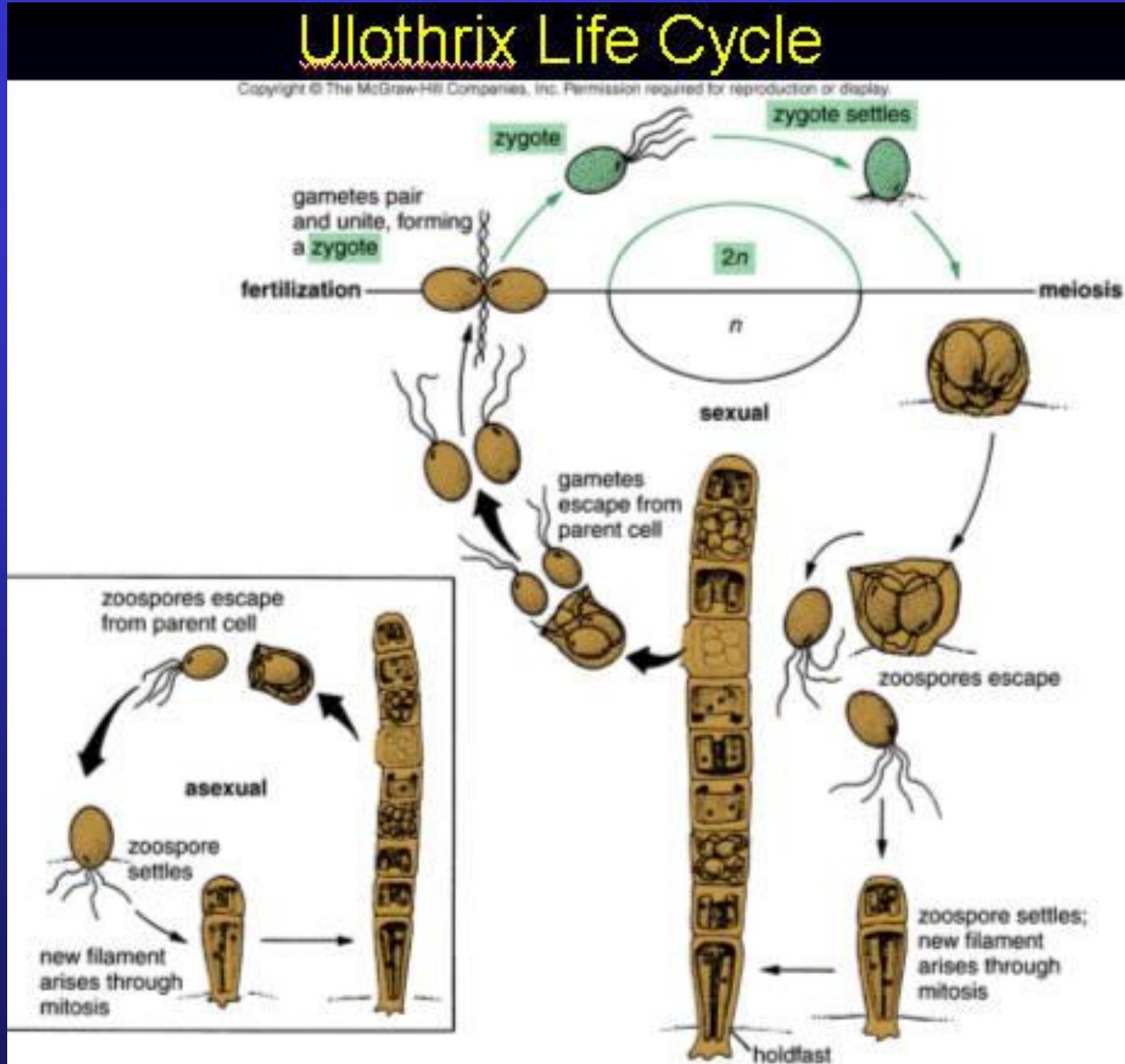


Ulothrix

haplontic cycle

izogamy

heterothalism

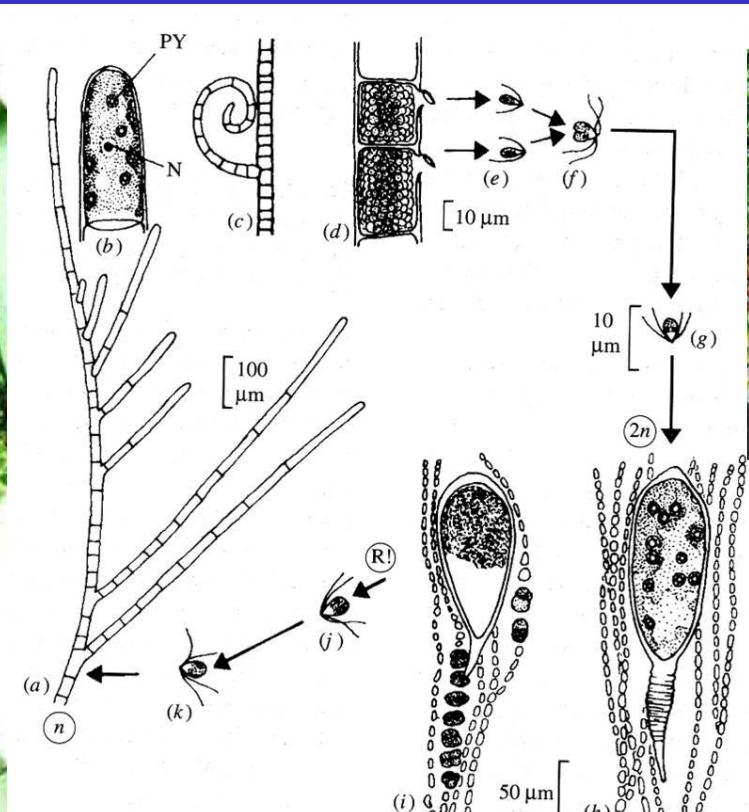
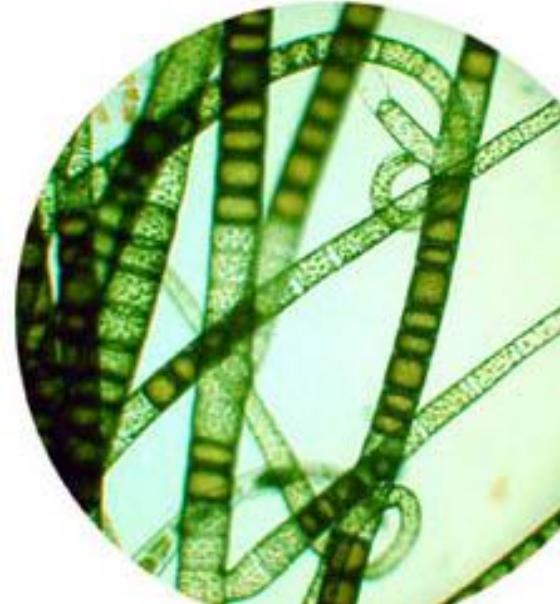
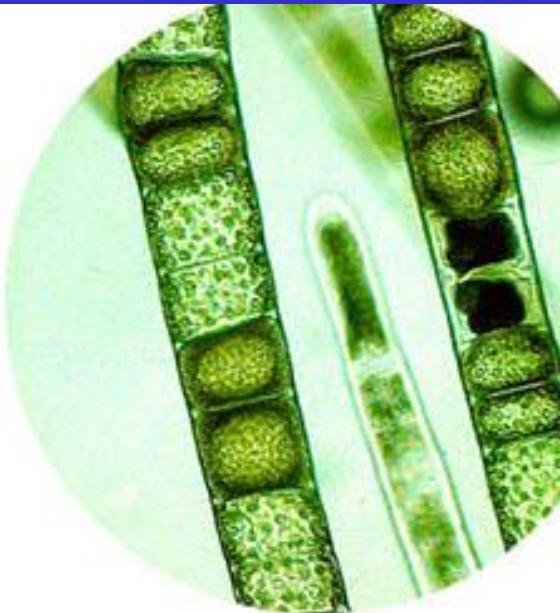


Ulothrix zonata

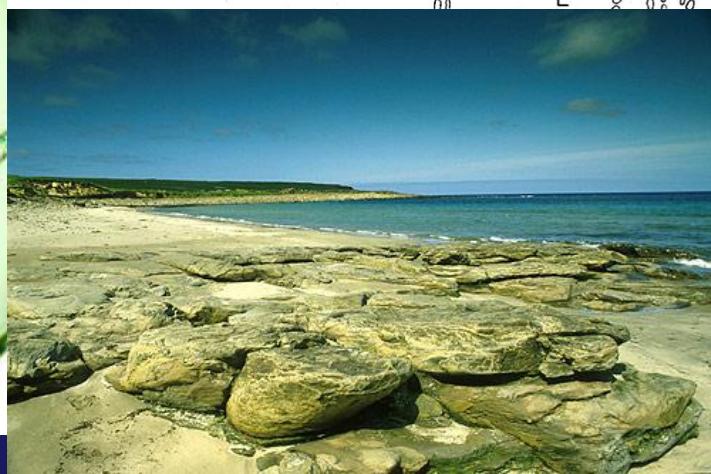


Spongomorpha

izogamy;
apical growth



uninuclear cells

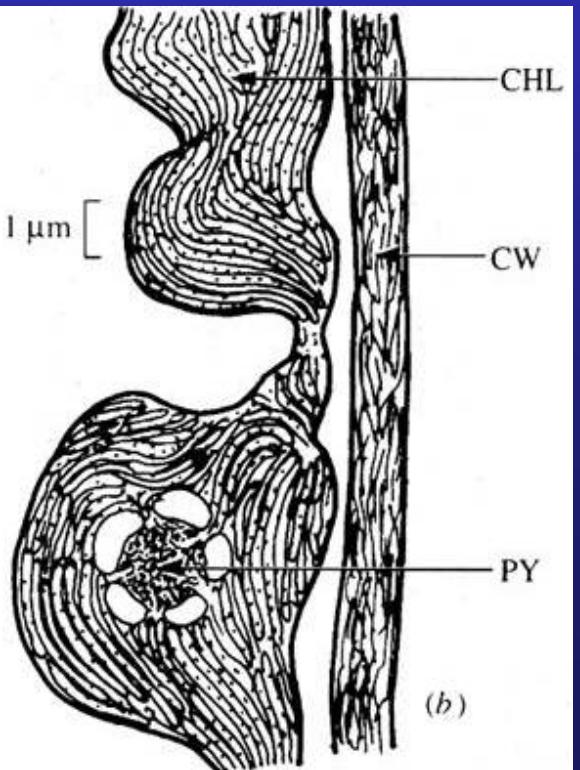


codiolar stage resembles
Petrocelis (red alga)

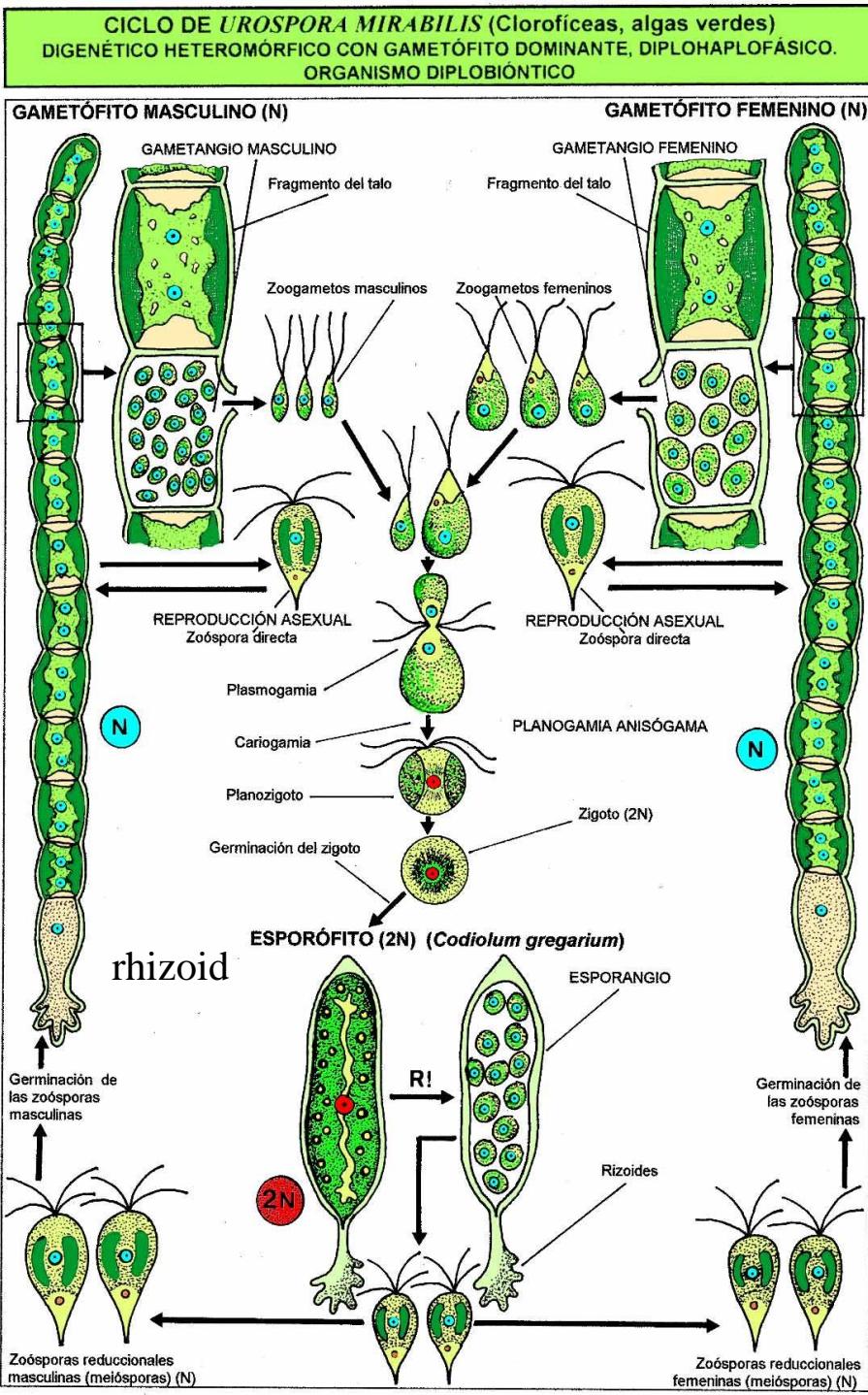
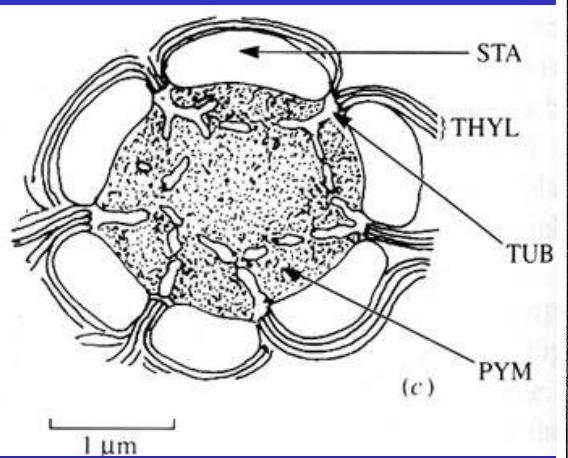
eulitoral of North
Altantic

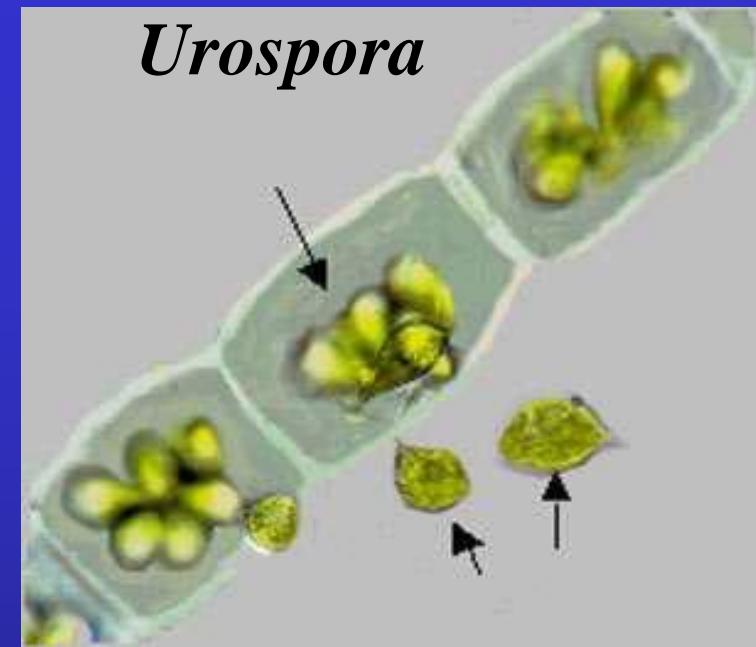
Urospora

anizogamy
heterothalism



multinucleate
cells

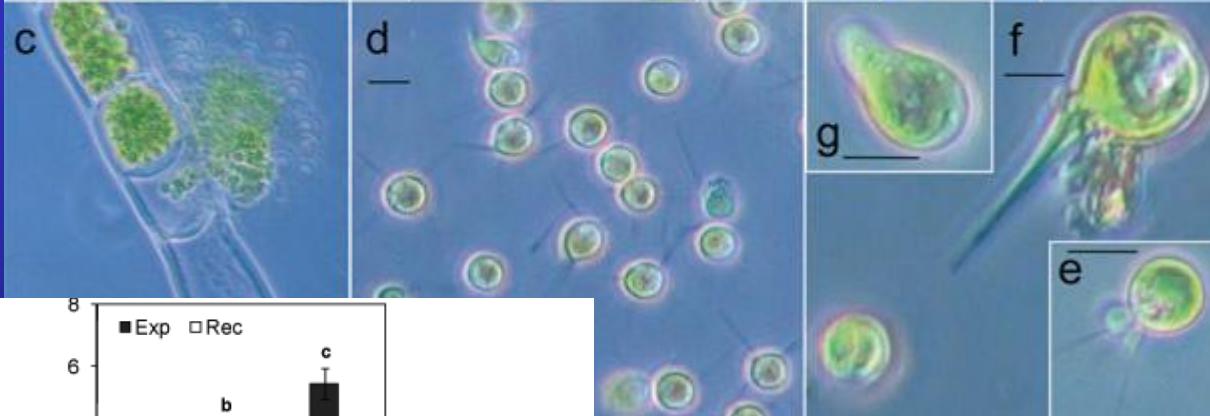
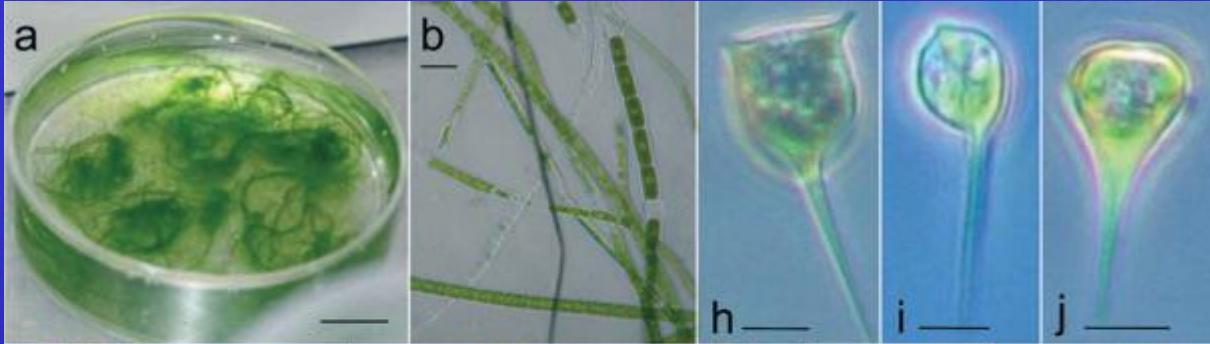




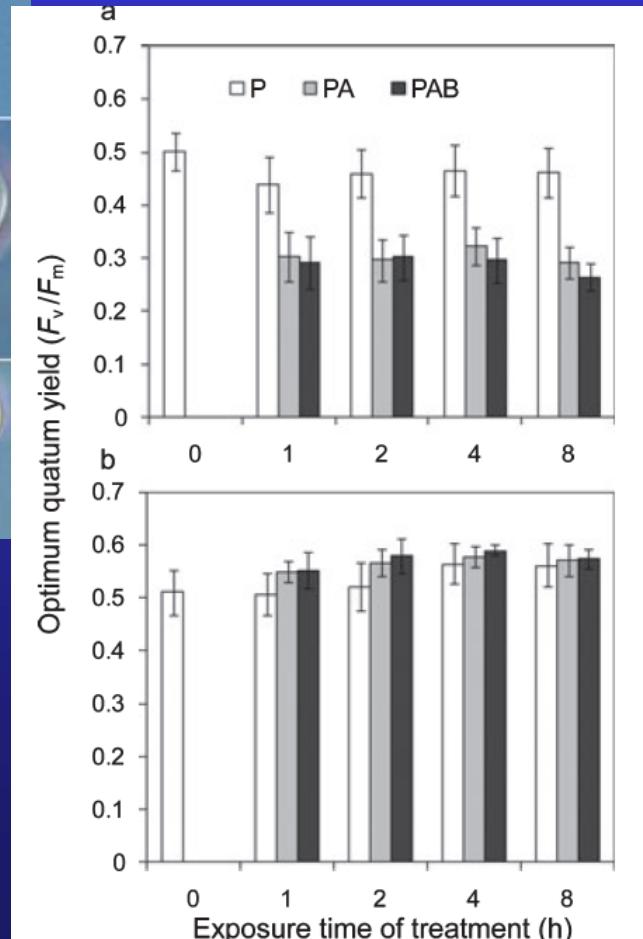
Arctic and Antarctic sea



Sensitivity of Antarctic Urospora to UV (UVA, UVB) radiation

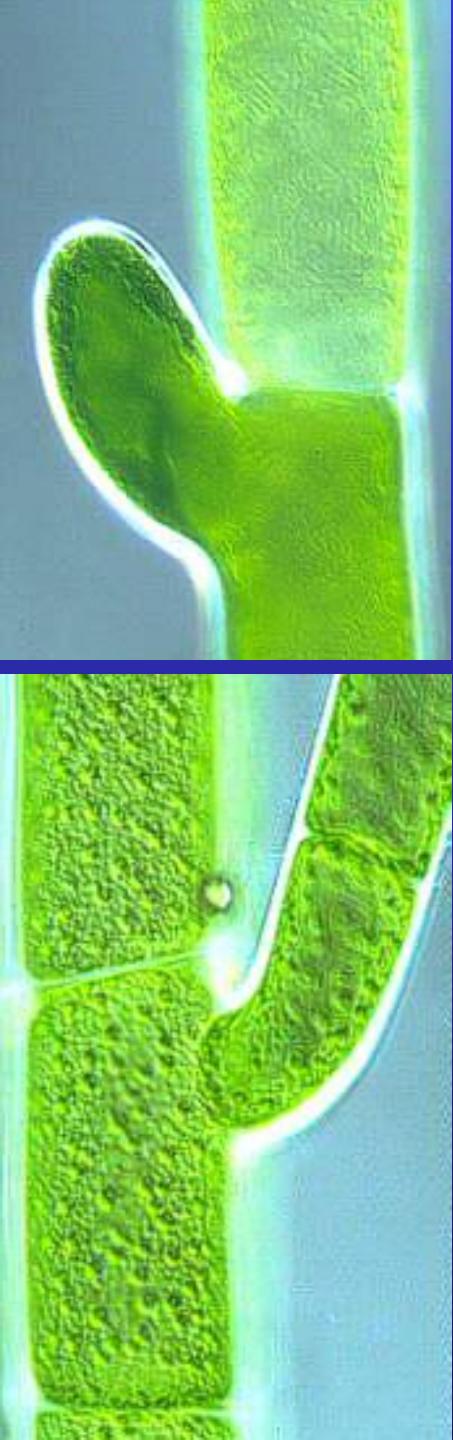


Gametophytic filaments
and propagules
(zoospores and gametes)



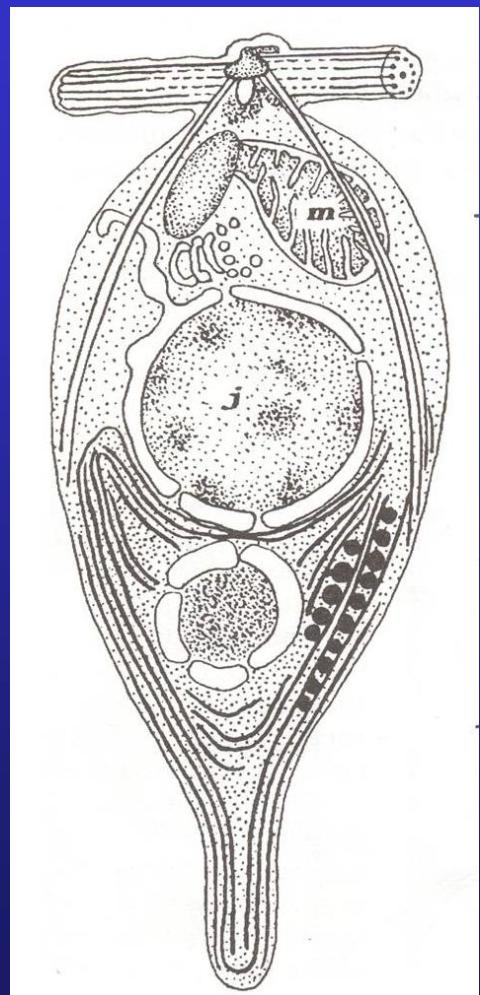
propagules slightly more sensitive, fast recovery

filaments with thick cell walls



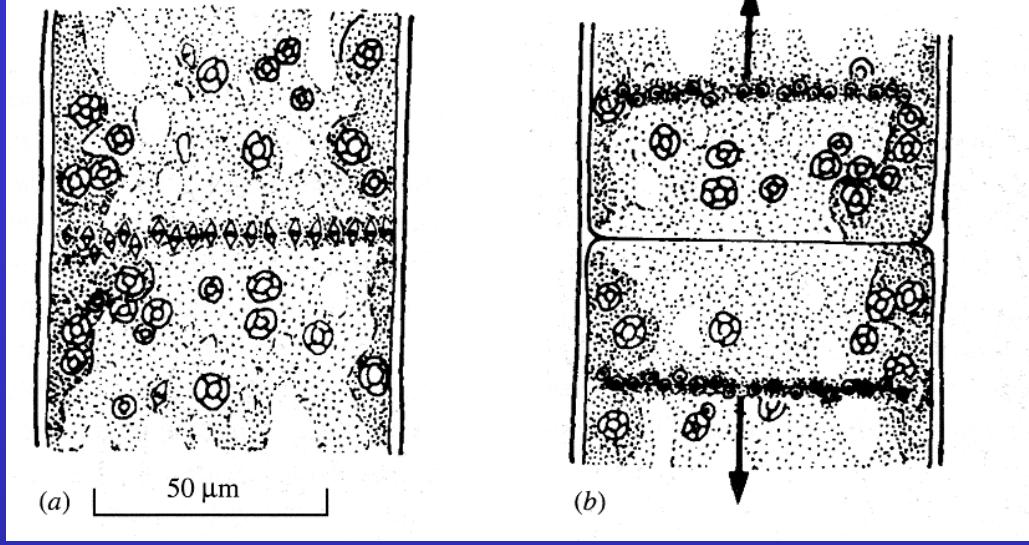
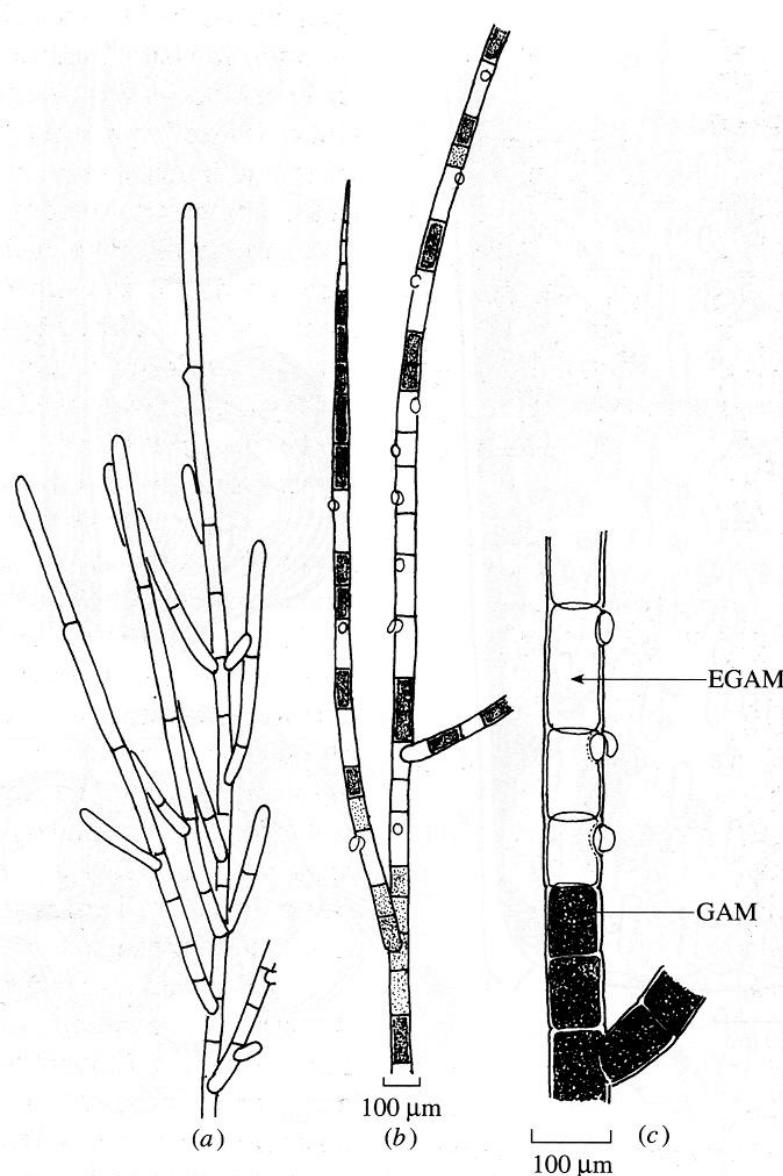
Acrosiphonia

2-flagellate gamete

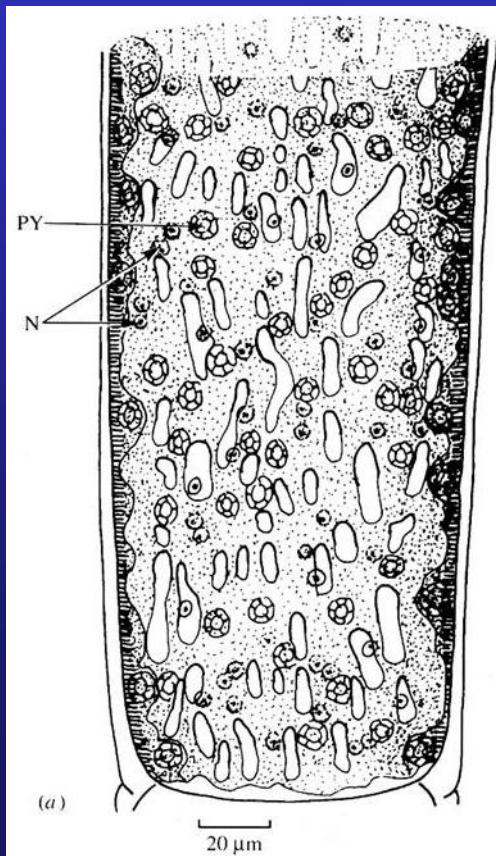


Branched filament, multinucleate cells. Epilithic in temperate and polar regions

Acrosiphonia

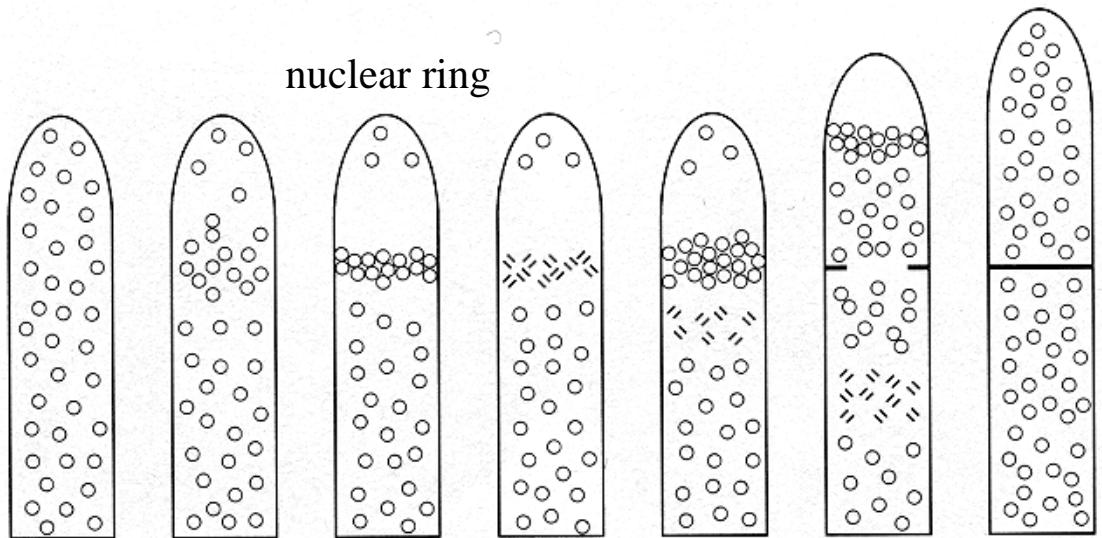


synchronous mitosis

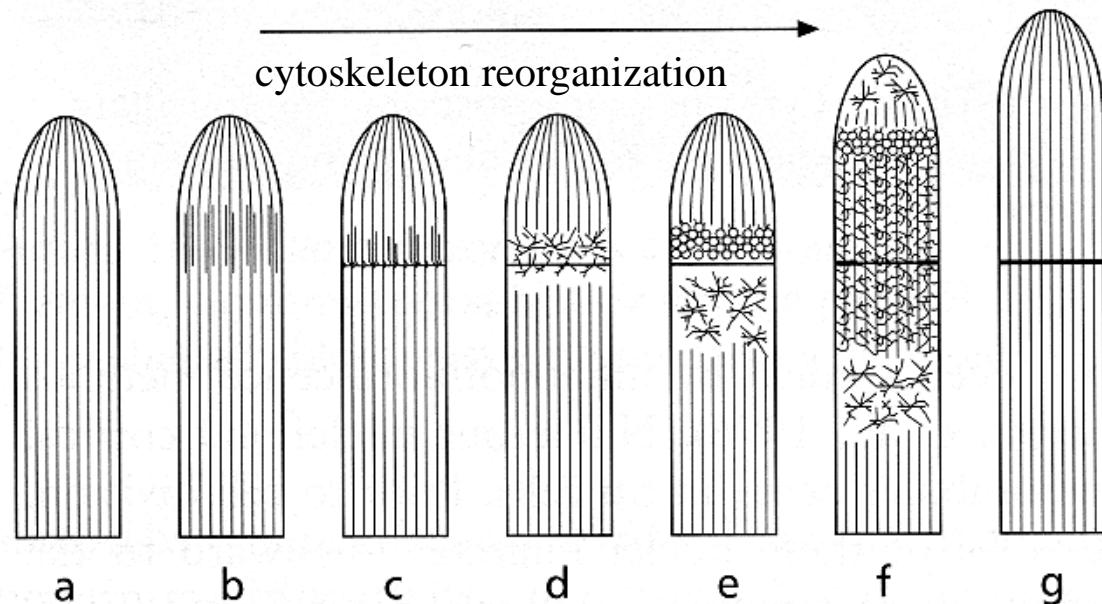


perforated chloroplast

Acrosiphonia

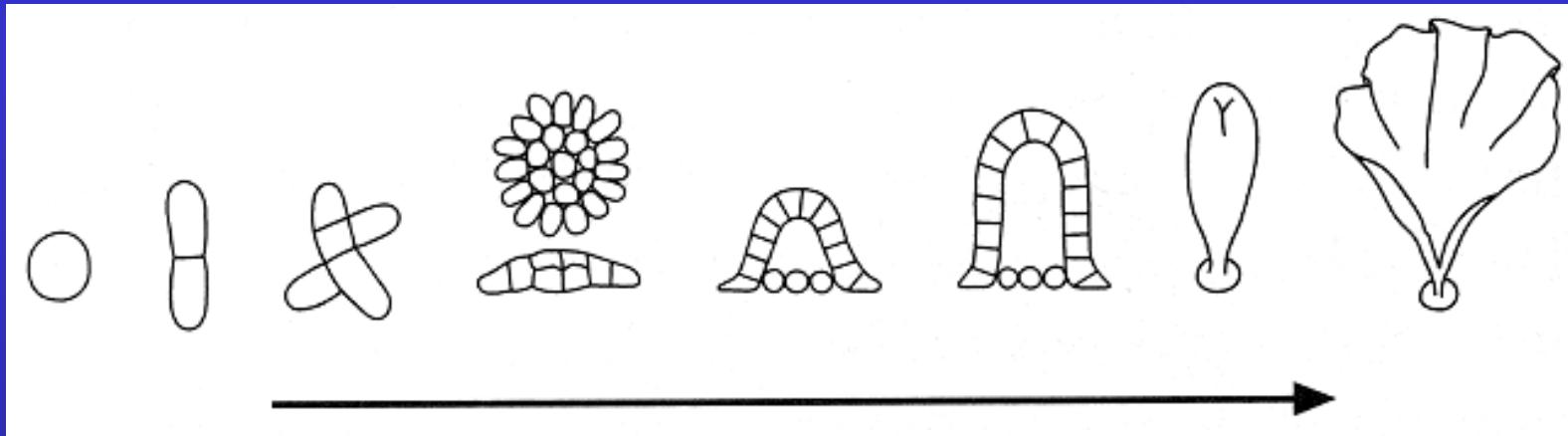


cytoskeleton reorganization



behaviour of nuclei and
microtubuli during mitosis

Monostroma



thallus formation from
the short filaments

monostromatic
thallus



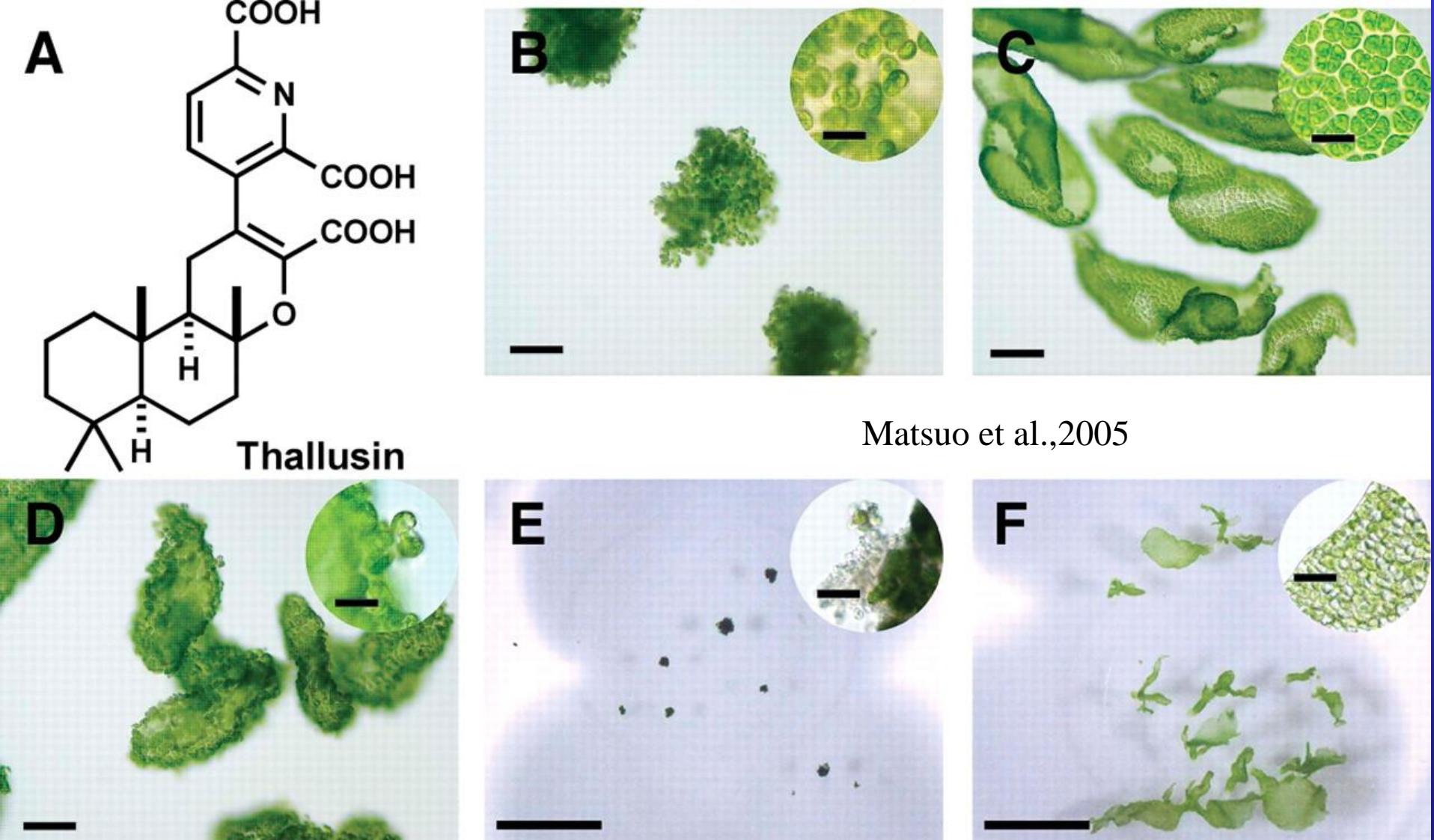


Fig. 1. (A) The structure of thallusin. (B) Morphology of axenic *M. oxyspermum*. (C) The effect of thallusin on *M. oxyspermum* (an initial concentration of 1 ng ml⁻¹). (D) Dedifferentiation given a lack of thallusin (an initial concentration of 1 pg ml⁻¹). Inset: Protruding cells from disintegrating thalli. (E) Small callus-like morphology of *U. pertusa*, after 2 months of cultivation under aseptic conditions. Inset: Colorless protrusions from the lateral cell walls. (F) The effect of thallusin (1 ng ml⁻¹) on *U. pertusa*. Inset: A foliaceous distromatic blade.

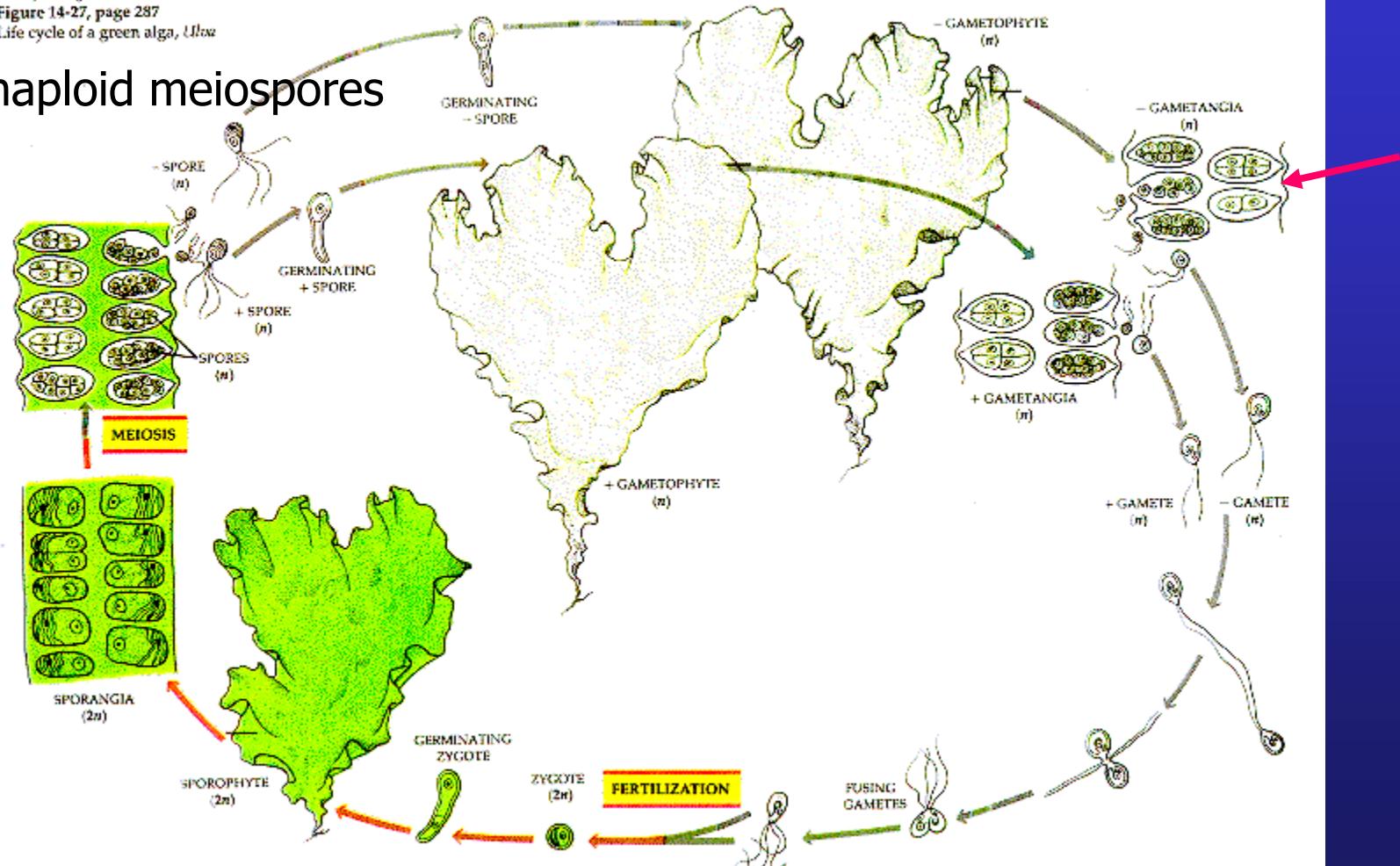
Řád: Ulvales

From the transparencies to accompany Peter H. Raven, Ray F. Evert, and Susan E. Eichhorn,
Biology of Plants, 5th edition. Worth Publishers, New York, 1992. Reproduced with permission.

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Transparency 49
Figure 14-27, page 287
Life cycle of a green alga, *Ulva*

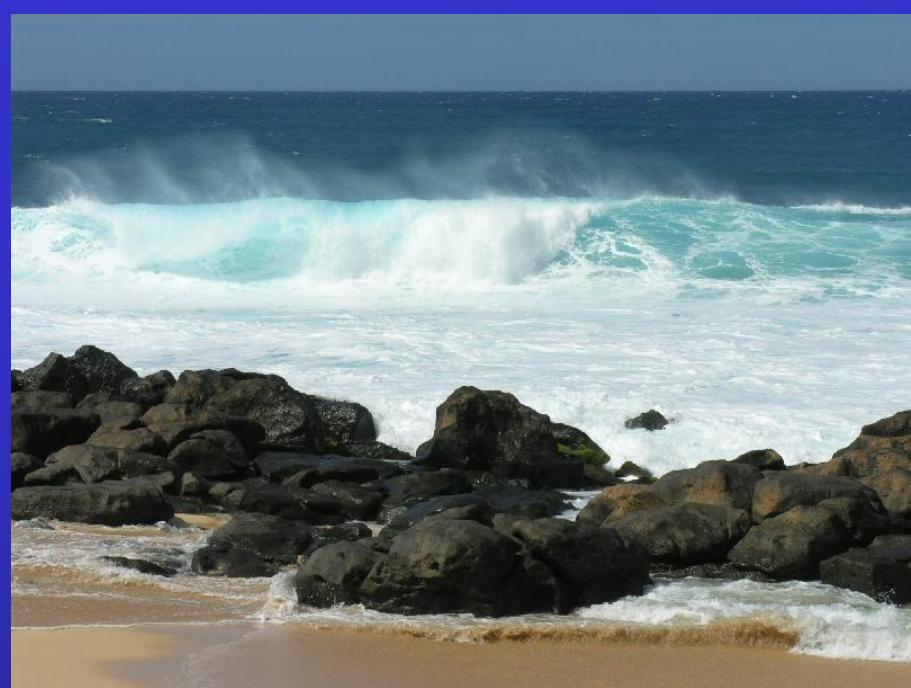
haploid meiospores



Ulva

Ulvaria

monostromatic thallus (single layer of cells)



temperate regions of the Northern hemisphere
North Atlantic, North Pacific



Enteromorpha=*Ulva*

Hayden et al. 2003 synonimize both genera



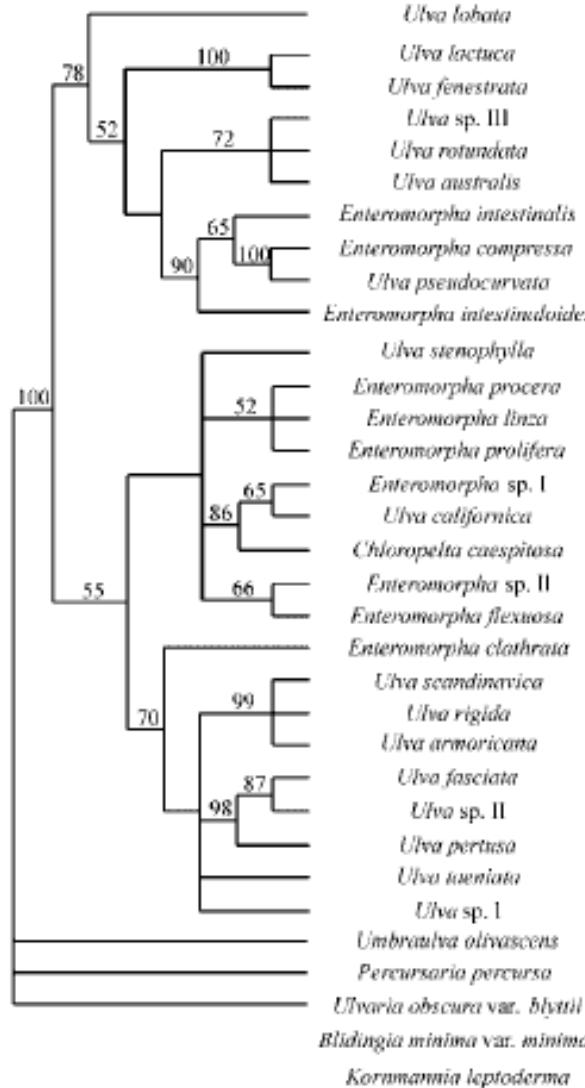
All after Entwistle et al. (1997)



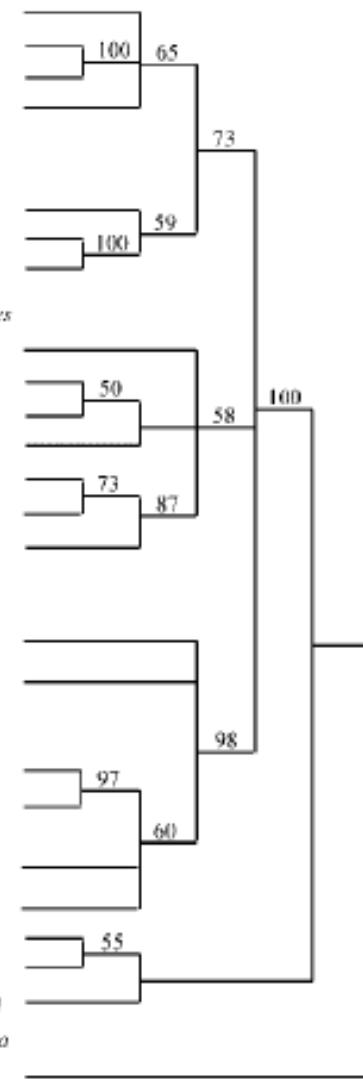
Table 3. Characters used to delimit species of *Ulva* and *Enteromorpha* based on Koeman and van den Hoek (1981) and Bliding (1963, 1968). Characters noted with (E) and (U) are used only in *Enteromorpha* and *Ulva*, respectively.

Character
Gross morphology, including colour and texture of mature plant
Structure of plant base
Arrangement and shape of cells in surface view
Structure of branch tips (E)
Number of pyrenoids per cell
Shape of chloroplast in surface view
Cell size at base, middle and apex of thallus
Height-to-width ratio of cells in cross section (U)
Thallus thickness (U)
Morphology of young germling
Mode of reproduction
Ecology

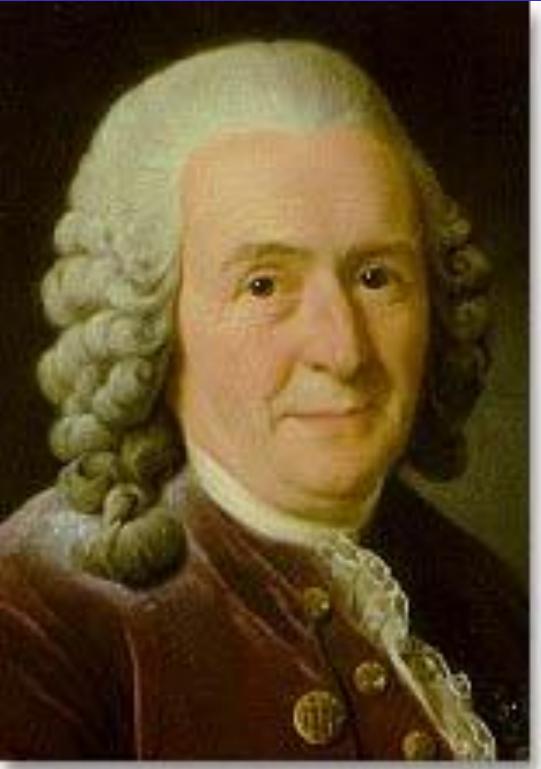
a. ITS nrDNA



b. *rbcL*



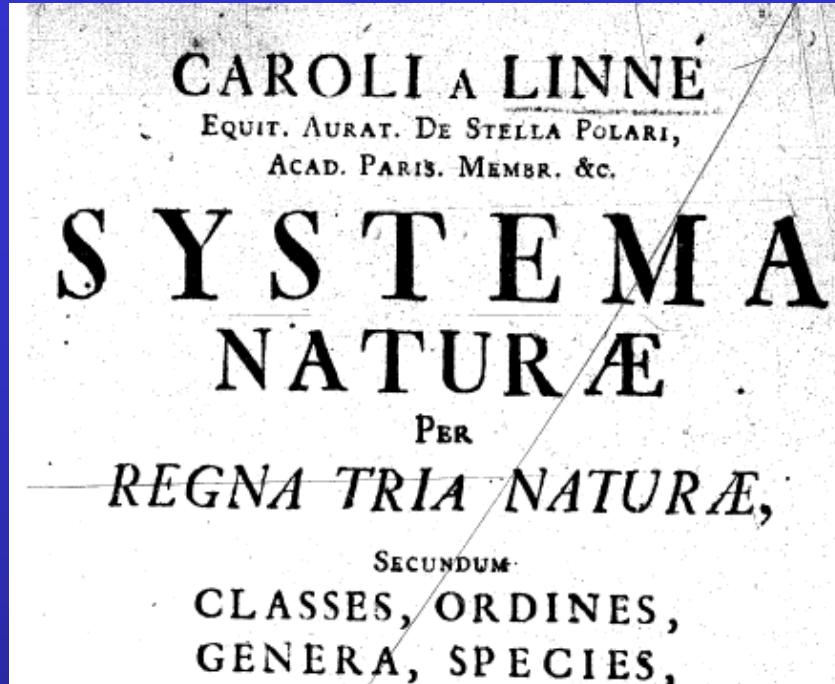
Hayden et al. 2003. Linnaeus was right all along: *Ulva* and *Enteromorpha* are not distinct genera.
European Journal of Phycology. 38: 277-294.



Carl Linné

(1719-1772)

only 5 algal genera discerned
(Tremella, Fucus, Ulva,
Conferva, Corallina + Volvox
(=animal like plant)



(1767)

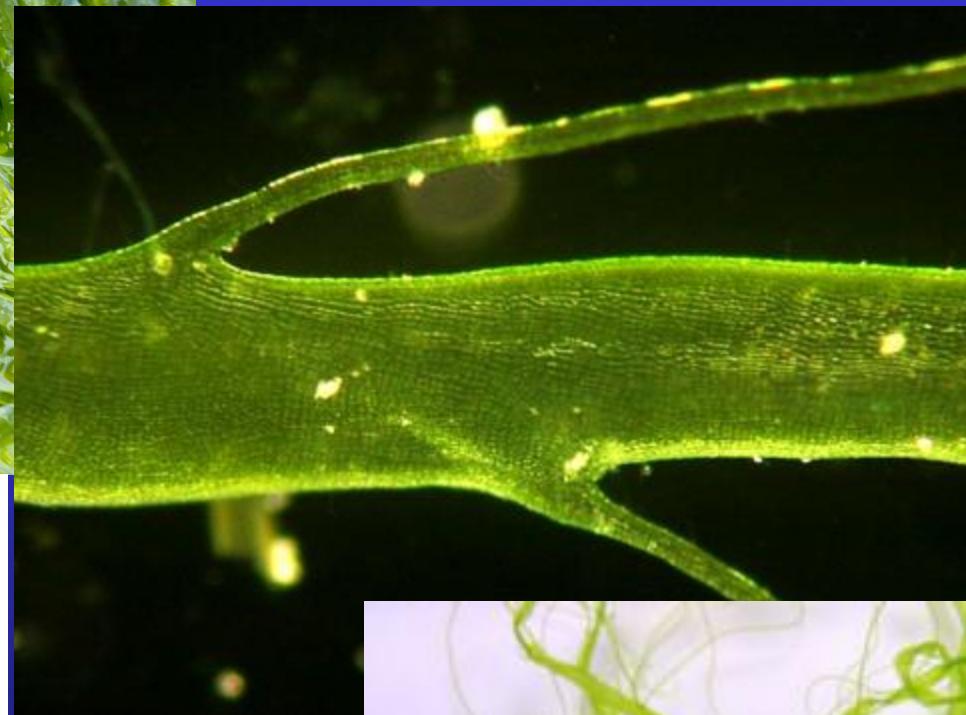
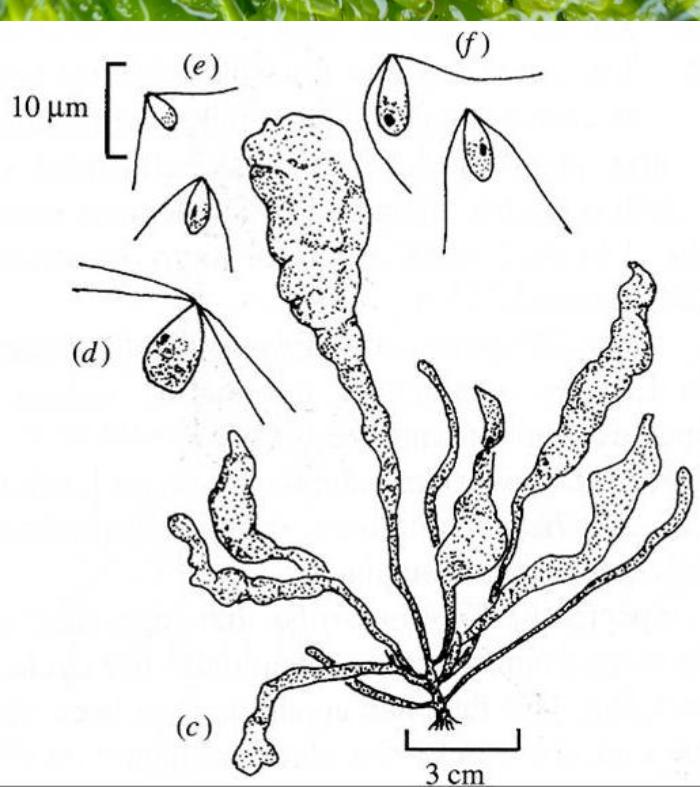
1204. TREMELLA. *Fru<ficationes* vix manifestæ,
1625. in corpore gelatinoso.

juniperin. 1. T. sessilis membranacea auriformis fulva.
Noftoc. 2. T. plicata undulata.
lichenoi- 4. T. erecta plana: margine crifo lacinulato. *Dill. muſc.*
des. t. 19. f. 32, 34, 35.
verruco- 5. T. tuberculosa folida rugosa. *Dill. muſc.* t. 10. f.
ja. 16.
difformis. 8. T. subrotunda sinuata difformis gelatinosa.
bemisph. 6. T. hemiſphericā ſparsa.
purpurea. 7. T. subglobofa ſeſſilis ſolitaria glabra.
adnata. 9. T. rotunda imbricata livida.

1205. FUCUS. MASC. *Veficulae* villis intertextæ.
1626. FEM. *Veficulae* adperfite granis im-
mersis apice prominulis. Sem. fo-
litaria.

ovarius. 36. F. caule filiformi ramoso, fol. conſertis ovatis for-
nicatis. *Habitat in O. Afſatico.* Folia conſerta
in racemum, retrorsum fornicata, membranacea,
purpurascens, parva. *Fru<ficationes* non vidi.

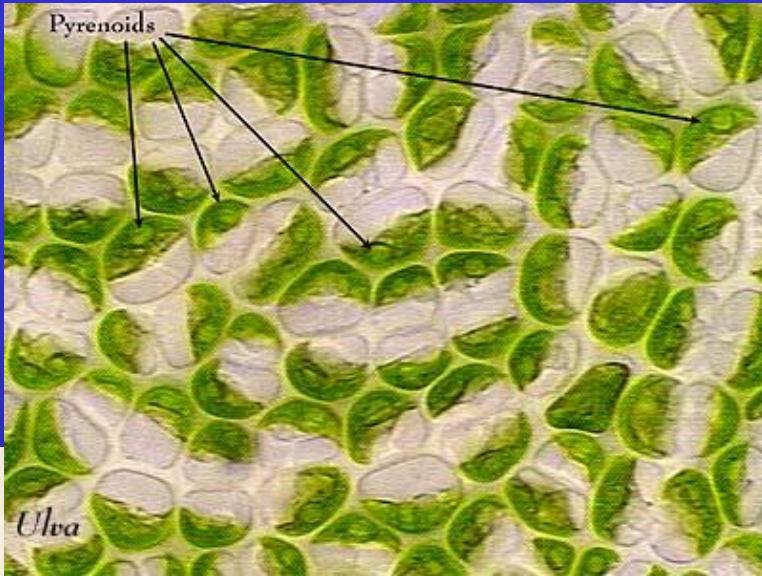
Enteromorpha=Ulva



release of
gametes
influenced by
lunar cycle



Ulva





Going Green: Beach Carpet



Ulva prolifera

(Qingdao on 18.7.2011)
South China





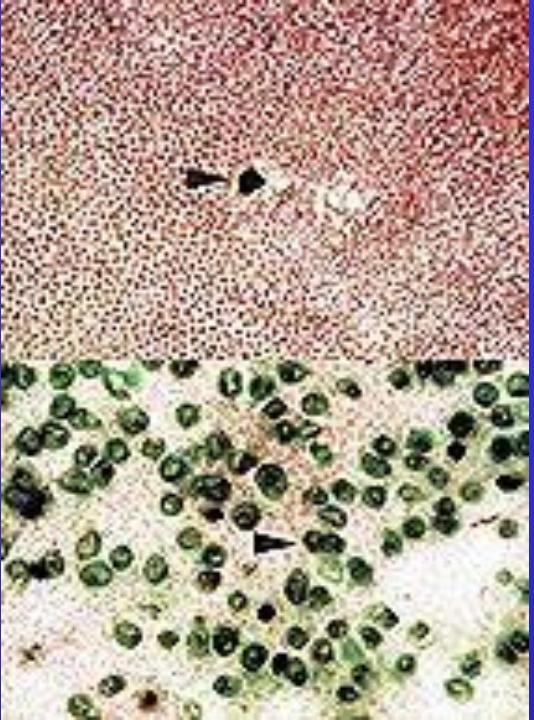


Photo by Viktor Kovalenko | YachtPals.com

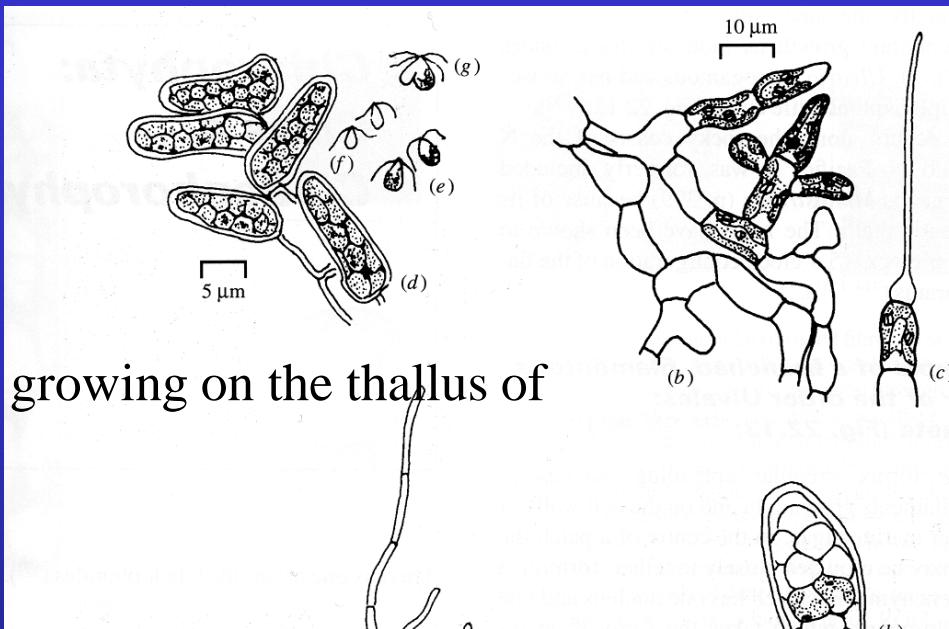
1. 7. 2008 - Sailing course for the Olympic Summer Games. Qingdao, China



Acrochaete - parasite

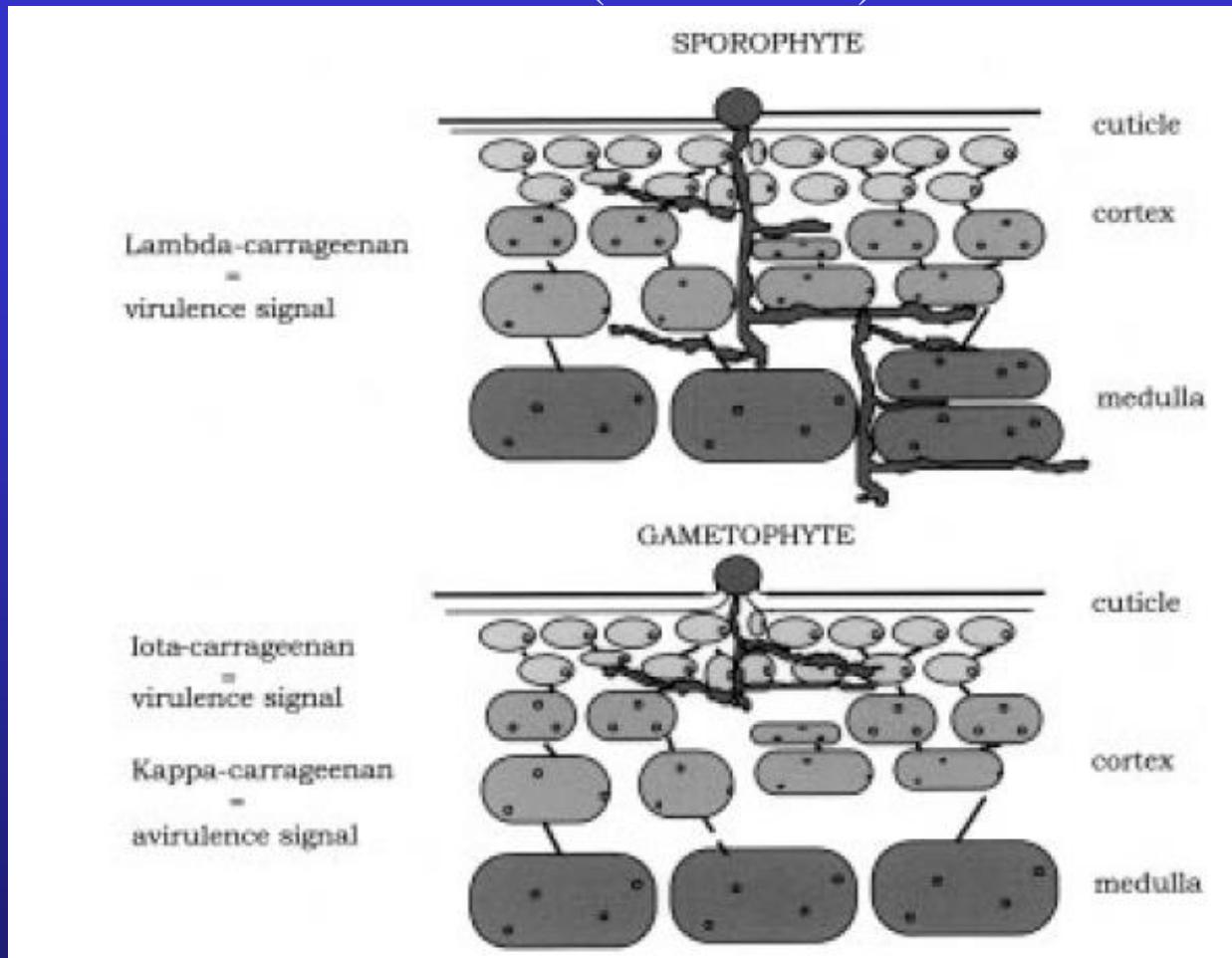


*Acrochaete
Chondrus*



growing on the thallus of

The virulence of *Acrochaete* mediated by CW composition of the host (*Chondrus*)



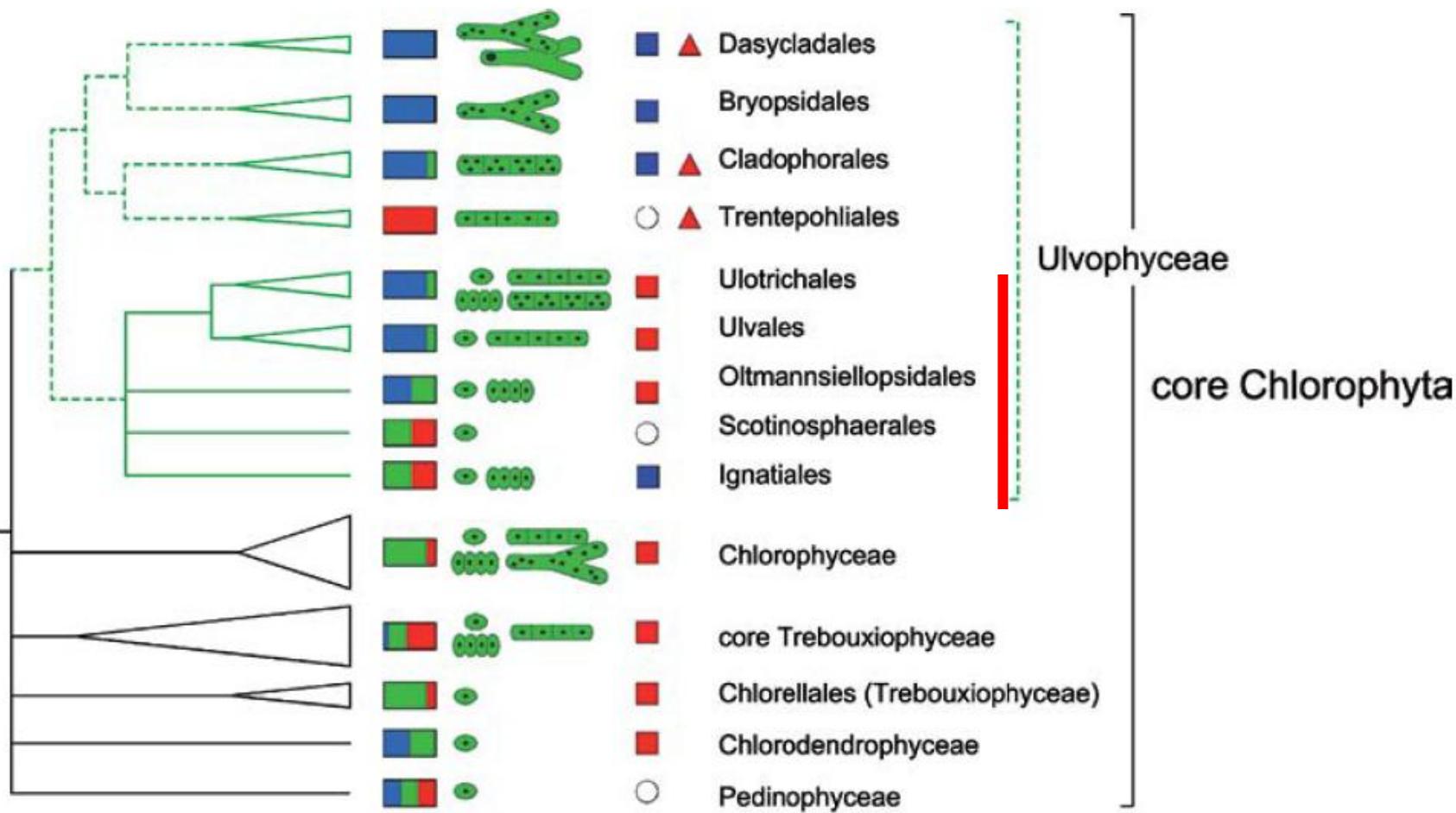
In this pathosystem, the host gametophytes are resistant to the pathogen, whereas the sporophytic generation is susceptible to infection. The virulence of the green algal pathogen is mediated by the recognition of carrageenan oligosaccharides released from its red algal host: kappa-carrageenan oligosaccharides inhibit *A. operculata* virulence while lambda-carrageenan oligosaccharides enhance its pathogenicity. Bouarab et al. 2004

Induced defence of red algae against pathogens



We investigate the alga/pathogen interaction in the system of the red alga *Chondrus crispus* (left on top) and the green algal pathogen *Acrochaete operculata* (below left), which is able to grow within the tissue of the red algal host. This process, called endophytism, can be inhibited during certain developmental phases of *C. crispus*. During the resistant, gametophytic, phase of the life cycle, *C. crispus* can recognise the attacker and kill it by an immediate release of hydrogen peroxide. We found that carrageenans from the red alga induce the release of asparagine from the green algal parasite. The free amino acid itself acts as a substrate for an amino acid oxidase of the host that releases micromolar amounts of hydrogen peroxide, sufficient to contain the attacker. The aim of a related collaborative project (European Union EPIFIGHT) is to understand the biological

carragenans induce the release of asparagine (Asn) from *Acrochaete*
Free Asn – substrate for amino acid oxidase of *Chondrus* – releas of
 H_2O_2 – enough to contain/kill the attacker



Habitat

- aero-terrestrial
- freshwater
- marine

Translational apparatus

- EFL
- EF-1 α
- unknown
- alternative nuclear genetic code

Cyto-morphological organisation

- Type 1a: flagellate or non-flagellate unicells, cells uninucleate
- Type 1b: flagellate or non-flagellate colonies, cells uninucleate
- Type 2: multicellular filaments or blades composed of uninucleate cells.
- Type 3: siphonocladous organisation: multicellular thalli, multinucleate cells
- Type 4: siphonous organisation: thallus a single tubular cell, multinucleate
- Type 4b: siphonous organisation: thallus a single tubular cell, single macronucleus