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Molecular data show that *Bryoria fremontii* and *B. tortuosa* (*Parmeliaceae*) are conspecific

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Abstract: *Bryoria fremontii* and *B. tortuosa* are the only species in the lichenized ascomycete genus *Bryoria* known to contain the pulvinic acid derivative vulpinic acid. In *B. fremontii* this yellow pigment is restricted to the soralia and apothecia, while in *B. tortuosa* it can occur throughout the thallus. The actual amount of vulpinic acid produced by *B. tortuosa* is rather variable, however, with intermediate specimens bearing both white and yellow pseudocyphellae. We studied the relationship between the two species with parsimony analysis using four DNA regions: 1) the internal transcribed spacers of the nuclear rDNA including the 5.8S region (ITS), 2) partial sequences from the intergenic spacer of the nuclear rDNA (IGS), 3) partial sequences from the small subunit of the mitochondrial rDNA (mtSSU), and 4) partial sequences from the protein-coding glyceraldehyde-3-phosphate dehydrogenase gene (GAPDH). Our phylogenetic analysis revealed that *B. fremontii* and *B. tortuosa* must be regarded as conspecific, but allowing for some genetic differentiation between European and North American populations. *Bryoria tortuosa* is therefore synonymized with *B. fremontii*.

Key words: *Bryoria*, molecular systematics, lichen, secondary chemistry, taxonomy

Introduction

Bryoria fremontii (Tuck.) Brodo & D. Hawksw. and *B. tortuosa* (G. Merr.) Brodo & D. Hawksw. constitute a chemically unique species group in the genus *Bryoria* Brodo & D. Hawksw. (*Parmeliaceae*, *Lecanorales*, Ascomycota) because of their production of a pulvinic acid derivative, vulpinic acid. Vulpinic acid is a bright yellow pigment that in *B. fremontii* appears only in soralia and apothecia but in *B. tortuosa* is present throughout the thallus, including the pseudocyphellae. In their monograph of the genus *Bryoria* in North America, Brodo & Hawksworth (1977) classified the two species in the section *Tortuosae* (Bystr.) Brodo & D. Hawksw. Both taxa occur mainly in western North America and Northern Europe. *Bryoria fremontii* was described from North America in 1858, but it was collected much earlier in Europe, for example, by G. Wahlenberg (Ahlner 1948), and the lectotype of the rejected name *Lichen jubatus* L. (Hawksworth & Sherwood 1981) most probably represents *B. fremontii*. *Bryoria tortuosa* was for a long time known only from western North America. Motyka (1958) was the first author to report *B. tortuosa* in Europe but the report has never been checked. Holien (1986) reported the species from northern Europe for the first time (see also Hermansson & Thor 2004; Myllys *et al.* 2006).

In addition to having a unique chemistry, *Bryoria fremontii* and *B. tortuosa* are morphologically very distinct within the genus, and hence easily recognized in the field. Both are large species (often 20–50 cm in length) and both have twisted, foveolate and often partly flattened main branches and finer, rather terete secondary branches. The colour of the thallus is typically reddish or yellowish brown

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depending on the concentration of vulpinic acid. Typical specimens of *B. fremontii* and *B. tortuosa* are also rather easily distinguished from one another; the main distinguishing features being the surface of the cortex (mostly shiny in the first versus usually dull or matt in the other) and the frequency and colour of the pseudocyphellae (sparse and whitish versus usually abundant and yellow, respectively). Soralia production is more variable, ranging from rather common and widespread in European *B. fremontii* (Ahlner 1948), to distinctly localized in North America. In *B. tortuosa*, by contrast, soralia are unknown from Northern Europe (Holien 1986; Hermansson & Thor 2004; Myllys *et al.* 2006), and very rare in North America (Brodo & Hawksworth 1977). Apothecia have been found in North America in both taxa (Brodo & Hawksworth 1977; this study), but in northern Europe only *B. fremontii* is known to produce sexual fruiting structures.

The habitat requirements of *Bryoria fremontii* and *B. tortuosa* seem to be fairly similar. In northern Europe both species prefer conifers growing in humid, shaded *Picea abies* forests as well as in dry, open *Pinus sylvestris* forests, the latter being especially favoured by *B. fremontii* (Ahlner 1948; Holien 1986; Gjerlaug 1987; Hermansson & Thor 2004; Myllys *et al.* 2006). In North America both taxa grow most copiously on conifer branches in open, usually late seral or old-growth forests in summer-dry intermontane regions at middle to upper elevations (Brodo & Hawksworth 1977; Goward 1999). Here it can also be noted that *B. tortuosa* in North America can be locally dominant in drier coastal and lowland forests.

Ever since its description roughly a century ago as "*Alectoria tortuosa*" (Merrill 1909), most authorities have accepted this taxon as a distinct species (e.g., Gyelnik 1934, 1935; Fink 1935; Motyka 1958; Hale 1979; McCune & Geiser 1997; Brodo *et al.* 2001; Spribille 2002; Derr *et al.* 2003). Goward & Ahti (1992) and Goward (1999), however, challenged this view, pointing out that the key features distinguishing *Bryoria tortuosa* from *B. fremontii* combine in varying degrees in some British Columbian populations. They suggested that *B. tortuosa* might more appropriately be considered as a variety or chemical strain of *B. fremontii*. Holien (1986) considered the species to be distinct, but found some differences in the chemical and morphological characters as well as in the ecology between the Norwegian and North American specimens of *B. tortuosa*. However, he concluded that those differences are of minor importance considering the overall morphological similarity of the specimens.

During our phylogenetic studies on the genus *Bryoria* using both molecular data and intensive investigation of thallus morphology and chemistry, it became obvious that the relationship of *B. fremontii* and *B. tortuosa* required further clarification. These two taxa, especially in North America, often grow entangled with each other and at the same time exhibit a continuum of variation ranging from shiny and reddish or dark brown thalli deficient in vulpinic acid, to dull yellowish thalli rich in vulpinic acid. Moreover, the colour of the pseudocyphellae can vary from whitish to yellow within the same specimen, indicating a low concentration and uneven distribution of vulpinic acid.

In this paper we have reconstructed a molecular phylogeny using four DNA regions to examine whether *Bryoria fremontii* and *B. tortuosa* are distinct species or whether the differences in their chemical and morphological characters represent a range of variation across a single species. Furthermore, we have used both North American (Canadian) and European material to test the hypothesis that specimens from different continents form two monophyletic groups.

Materials and Methods

The material

The material used in this study was selected to represent as broad a range of morphological and chemical variation as possible. Altogether nine specimens of *Bryoria fremontii* and 27 specimens of *B. tortuosa* from both Europe and North America were included in the analyses (Table 1). Specimens were determined as *B. tortuosa* if there was any trace of vulpinic acid in the thallus. In a preliminary phylogenetic analysis using

TABLE 1. *Specimens used in the phylogenetic analyses with collection data and GenBank accession numbers.* Bryoria tortuosa *specimens were submitted to GenBank as* B. fremontii *(see discussion).*

Taxon	Voucher specimen	GenBank acc. no.			
		ITS	IGS	GAPDH	mtSSU
Ingroup					
Bryoria fremontii	Finland 2002 *Haikonen* 21909 (H)	FJ668495	FJ668457	FJ668401	FJ668429
B. fremontii	Finland 2005 *Myllys* 481 (H)	FJ668497	FJ668459	FJ668403	FJ668431
B. fremontii	Finland 2005 *Velmala* 13b, *Halonen & Myllys* (H)	FJ668498	FJ668460	FJ668404	FJ668432
B. fremontii	Finland 2005 *Velmala* 35b, *Halonen & Myllys* (H)	FJ668499	FJ668461	FJ668405	FJ668433
B. fremontii	Russia, Karelia 2002 *Uotila* 43962 (H)	FJ668500	FJ668462	FJ668406	FJ668434
B. fremontii	Canada, B. C. 2004 *Crawford* 7 (H)	FJ668502	FJ668464	–	–
B. fremontii	Canada, B. C. 2005 *Goward* 05-04 (UBC)	FJ668503	FJ668465	FJ668408	FJ668436
B. fremontii	Canada, B. C. 2005 *Goward* 05-16 (UBC)	FJ668504	FJ668466	FJ668409	FJ668437
B. fremontii	Canada, B. C. 2007 *Goward* 07020027 (UBC)	FJ668501	FJ668463	FJ668407	FJ668435
B. tortuosa	Finland 1998 *Kääntönen* 9/98 (H)	FJ668510	FJ668472	FJ668415	FJ668443
B. tortuosa	Finland 2000 *Kääntönen* 99/2000 (H)	FJ668511	FJ668473	FJ668416	FJ668444
B. tortuosa	Finland 2001 *Kääntönen* 10/01 (H)	FJ668496	FJ668458	FJ668402	FJ668430
B. tortuosa	Finland 2001 *Kääntönen* 229/01 (H)	FJ668509	FJ668471	FJ668414	FJ668442
B. tortuosa	Finland 2004 *Haikonen* 23658 (H)	FJ668508	FJ668470	FJ668413	FJ668441
B. tortuosa	Finland 2004 *Halonen & Hyvärinen* s.n. (OULU)	FJ668506	FJ668468	FJ668411	FJ668439
B. tortuosa	Finland 2005 *Halonen, Myllys & Velmala* 32 (H)	FJ668512	FJ668474	FJ668417	FJ668445
B. tortuosa	Finland 2005 *Myllys* 490 (H)	FJ668507	FJ668469	FJ668412	FJ668440
B. tortuosa	Sweden 2000 *Klintberg* 11347 (UPS)	FJ668505	FJ668467	FJ668410	FJ668438
B. tortuosa	Canada, B. C. 2004 *Crawford* 22 A (H)	FJ668522	FJ668484	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford* 22 B (H)	FJ668527	FJ668489	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford 25* A (H)	FJ668525	FJ668487	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford* 25 B (H)	FJ668528	FJ668490	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford* 25 C (H)	FJ668529	FJ668491	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford* 43 (H)	FJ668526	FJ668488	FJ668426	FJ668454
B. tortuosa	Canada, B. C. 2004 *Crawford* 52 (H)	FJ668521	FJ668483	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford* 56 (H)	FJ668524	FJ668486	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford 74* (H)	FJ668530	FJ668492	–	–
B. tortuosa	Canada, B. C. 2004 *Crawford 77* (H)	FJ668523	FJ668485	–	–
B. tortuosa	Canada, B. C. 2005 *Wright* 2005-16 (UBC)	FJ668513	FJ668475	FJ668418	FJ668446
B. tortuosa	Canada, B. C. 2007 *Goward* 07020001 (UBC)	FJ668514	FJ668476	FJ668419	FJ668447
B. tortuosa	Canada, B. C. 2007 *Goward* 07020003 (UBC)	FJ668515	FJ668477	FJ668420	FJ668448
B. tortuosa	Canada, B. C. 2007 *Goward* 07020013 (UBC)	FJ668516	FJ668478	FJ668421	FJ668449
B. tortuosa	Canada, B. C. 2007 *Goward* 07020016 (UBC)	FJ668517	FJ668479	FJ668422	FJ668450
B. tortuosa	Canada, B. C. 2007 *Goward* 07020018 (UBC)	FJ668518	FJ668480	FJ668423	FJ668451
B. tortuosa	Canada, B. C. 2007 *Goward* 07020025 A (UBC)	FJ668519	FJ668481	FJ668424	FJ668452
B. tortuosa	Canada, B. C. 2007 *Goward* 07020025 B (UBC)	FJ668520	FJ668482	FJ668425	FJ668453
Outgroup					
B. capillaris	Finland 2003 *Haikonen* 22228 (H)	FJ668493	FJ668455	FJ668399	FJ668427
B. glabra	Finland 2004 *Halonen* s.n. (OULU)	FJ668494	FJ668456	FJ668400	FJ668428

Platismatia glauca (L.) W. L. Culb. & C. F. Culb. as an outgroup, both the genus *Bryoria* and a clade including all *B. fremontii* and *B. tortuosa* specimens were monophyletic (tree not shown). In our final analysis we used *Bryoria capillaris* (Ach.) Brodo & D. Hawksw. and *Bryoria glabra* (Motyka) Brodo & D. Hawksw. as outgroup taxa in this study.

Secondary chemistry

The secondary compounds were examined using thin-layer chromatography (TLC) according to Orange *et al.* (2001). The acetone extracts were spotted with 75 mm / 75 µl Haematocrit capillaries (Hirschmann Laborgeräten) on 10 × 20 cm Merck silica gel 60 F-254 pre-coated glass plates and run in solvent systems A and B (B formulae from both Culberson 1972 and Mietzsch *et al.* 1994 according to Orange *et al.* 2001 were used).

Molecular techniques

Four DNA regions were used in this study: 1) the internal transcribed spacers of the nuclear rDNA including the 5.8S region (ITS); 2) partial sequences from the intergenic spacer of the nuclear rDNA (IGS); 3) partial sequences of the small subunit of the mitochondrial rDNA (mtSSU); and 4) partial sequences from the protein-coding glyceraldehyde-3-phosphate dehydrogenase gene (GAPDH).

DNA was extracted using either Qiagen's DNeasy Plant Mini Kit or DNeasy Blood & Tissue Kit following the manufacturer's protocol except that the liquid nitrogen phase was omitted. Instead, thallus fragments of approximately 0·5–3 cm long were ground with minipestles in 40 µl of the lysis buffer after which 140 µl of the buffer was added. The extracted DNA was eluted in 120 µl of the elution buffer included in the kits.

Primers used for PCR amplification were: a) for the ITS region: ITS1-F (Gardes & Bruns 1993) together with ITS4 (White *et al.* 1990) or ITS1-LM (Myllys *et al.* 1999) together with ITS2-KL (Lohtander *et al.* 1998); b) for the IGS region: IGS12B (Printzen & Ekman 2002) together with nu-SSU-0072-5′ (Gargas & Taylor 1992); c) for the mtSSU region: mtSSU1-KL together with mtSSU2-KL (Lohtander *et al.* 2002); and d) for the GAPDH region: Gpd1-LM together with Gpd2-LM (Myllys *et al.* 2002). PCR reactions were prepared using PuReTaq Ready-To-Go PCR beads (GE Healthcare). In addition, the 25 µl reaction volume contained 19 µl dH_2O, 0·4 µM each primer and 4 µl extracted DNA. The parameters for the PCR schedule were: initial denaturation for 5 min at 95°C followed by five cycles of 30 s at 95°C (denaturation), 30 s at 60°C or 58°C for ITS, 55°C or 50°C for IGS, 52°C for mtSSU and 56 °C for GAPDH (annealing), and 1 min at 72°C (extension); in the remaining 30 cycles the annealing temperature was decreased to 58°C or 56°C (ITS), 50°C (mtSSU) and 54°C (GAPDH); the PCR schedule ended to final extension for 7 min at 72°C.

The PCR products were visualized under UV light on 1 % agarose gels stained with ethidium bromide to check the size and quality of the DNA. The PCR products were purified according to the manufacturers' protocols with either Qiagen's QIAquick PCR Purification Kit (PCR products eluted in 30 µl elution buffer) or GE Healthcare illustra's GFX tm PCR DNA and Gel Band Purification Kit (PCR products eluted in 50 µl dH_2O).

The sequencing reactions were prepared using BigDye Terminator Cycle Sequencing Reaction Kit version 1.1 (Applied Biosystems). 10 µl reaction samples contained 3 µl dH_2O, 2 µl BigDye, 2 µl sequencing buffer, 0·25 µM each primer and 2 µl purified PCR product. The above listed PCR primers were used also for sequencing except for the ITS region for which the primer ITS5 (White *et al.* 1990) was used together with ITS2-KL. The sequencing reactions were run with the following parameters: initial denaturation for 1 min at 96°C or no initial denaturation, followed by 30 cycles of 30 s at 96°C, 15 s at 50°C and 4 min at 60°C.

The post-reaction purification of the samples for ABI PRISM™ DNA Sequencer (Applied Biosystems) was made following the protocol described in Högnabba (2006) and for MegaBACE 1000 DNA Analysis System (GE Healthcare) using Montage SEQ_{96} Cleanup Kit and MultiScreen SEQ_{384} Filter Plates (Millipore) according to the manufacturer's protocol. In some cases the cleaned PCR products were sequenced by Macrogen Inc., South Korea (www.macrogen.com). The DNA strands were assembled and manually corrected with SeqMan II 4.00 (DNASTAR).

Sequence alignment and phylogenetic analyses

Each DNA region was aligned separately using ClustalX 1.87 (Thompson *et al.* 1997) with default parameters. The four aligned sequence matrices were manually edited and combined with MacClade 4.08 (Maddison & Maddison 2005) for simultaneous parsimony analysis. The cladograms were obtained using PAUP* 4.0b10 (Swofford 2002) with the following settings: heuristic search, random addition sequence with 500 replicates and TBR branch swapping. No more than 40 trees were saved for each replicate to save computation time. Gaps were treated as fifth character states and, for comparison, as missing data in a second set of analyses. Support for each node was estimated using bootstrapping (1000 repetitions) as implemented in PAUP*. We used equally weighted maximum parsimony as our optimality criterion, which according to Grant & Kluge (2003) maximizes congruence over all data by minimizing the total number of transformations. To assess character congruence between the four DNA regions, a partition-homogeneity test was performed on each pair of partitions using PAUP*. The test included 1000 homogeneity replicates each employing a heuristic search with 500 addition-sequence replicates. Uninformative characters were excluded (Cunningham 1997).

Unfortunately we could not obtain mtSSU and GAPDH sequences from all of the specimens. Consequently, in order to minimize the amount of missing data, we performed two analyses with different data sets. First, ITS + IGS data set, which included all 36 ingroup

specimens. Second, ITS + IGS + mtSSU + GAPDH data set, which included 26 specimens from which all DNA regions were successfully sequenced.

Results

Secondary chemistry

In addition to vulpinic acid, some of the *Bryoria tortuosa* specimens analysed contained norstictic and barbatolic acids. The TLC procedure was repeated for those specimens. In some cases the results were not identical in the two runs (e.g., no barbatolic acid was detected in the second run) most probably because the material for the second run was obtained from different branches. Many of the specimens also contained some unidentified lichen compounds, with slightly different R_f -classes from those recorded by Holien (1986).

Phylogenetic analyses

One hundred and thirty-two new sequences were generated for this study, including 38 ITS, 38 IGS, 28 mtSSU and 28 GAPDH sequences (Table 1). The sequences showed very little variation in length except at the 5′ and 3′ ends of each region due to poor sequence quality. The ITS + IGS data set contained 998 characters of which 45 (4·5 %) were parsimony informative. The PAUP* analysis resulted in two equally most parsimonious trees of 94 steps, with consistency index (CI) of 0·989 and retention index (RI) of 0·989. The ITS + IGS + mtSSU + GAPDH data set contained 3056 characters of which 195 (6·4 %) were parsimony informative. The PAUP* analysis resulted in 468 equally most parsimonious trees of 342 steps, CI of 0·956 and RI of 0·944. According to the partition-homogeneity test, two out of seven pairs of partitions were significantly incongruent, the IGS-GAPDH pair (P = 0·042) and the ITS-GAPDH pair (P = 0·001).

In the strict consensus trees of both analyses (Figs 1 & 2), the ingroup was monophyletic with a 100 % bootstrap support value. The relationships in the ingroup were mostly unresolved. In the ITS + IGS analysis, two clades with only European specimens appeared with bootstrap support values of 61 % and 98 % as well as two clades with only North American *Bryoria tortuosa* specimens with bootstrap support values of 87 % and 62 %. In the IGS + ITS + mtSSU + GAPDH analysis only two clades appeared both including merely North American specimens (support values 69 % and 100 %). The analyses with gaps treated as missing data revealed almost identical tree topologies (trees not shown).

In the ITS + IGS data set the IGS region was slightly more informative than the ITS (5·2 % vs. 4·0 %). In the ITS + IGS + mtSSU + GAPDH data set the mtSSU was the most informative region (9·3 %), followed by IGS (5·4 %), GAPDH (5·1 %) and ITS (4·0 %). Without the outgroup taxa, however, the information contents of the four DNA regions were considerably lower. In the ITS + IGS data set the regions were almost equally informative (ITS 1·1 % vs. IGS 0·9 %). In the ITS + IGS + mtSSU + GAPDH data set the mtSSU was the most informative region (7·3 %) followed by IGS and ITS (both 0·9 %) and GAPDH (0·6 %).

Discussion

Neither *Bryoria fremontii* or *B. tortuosa* was monophyletic in the combined analyses. Furthermore, the few monophyletic groups found in the strict consensus trees did not correlate with the current taxonomic delimitation. Although some of the clades included only *B. tortuosa* specimens, the remaining ones contained both *B. fremontii* and *B. tortuosa* specimens (Figs 1 & 2). Interestingly, in spite of low resolution all clades in the strict consensus trees included either North American or European specimens, hence suggesting some level of geographical population differentiation, although analysis of more collections is needed before reaching definite conclusions. We interpret these results as consistent with the hypothesis that the chemical and morphological variation of the sample material fall within the range of

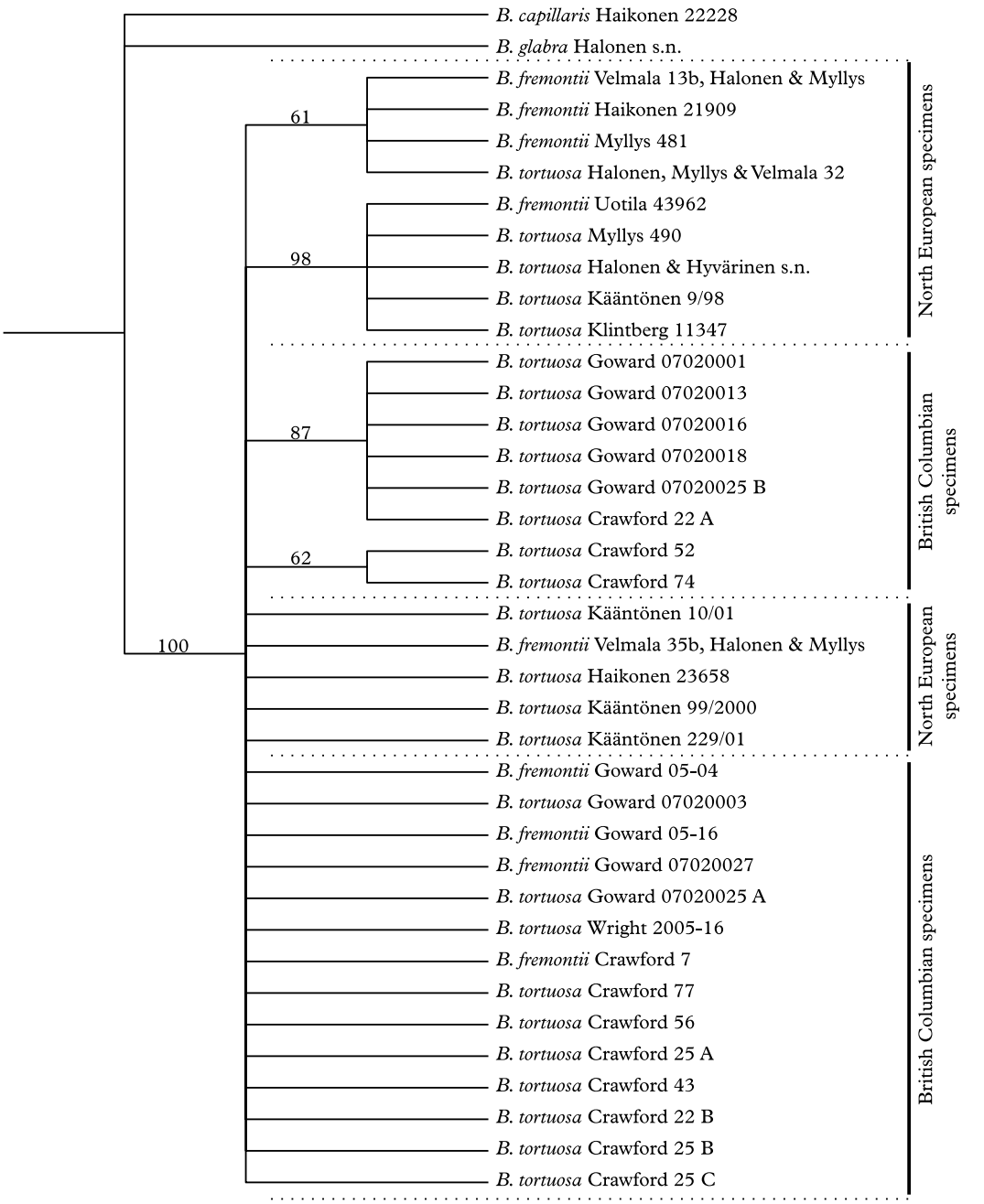

FIG. 1. Molecular phylogeny of *Bryoria fremontii* and *B. tortuosa*. A strict consensus tree of two trees based on ITS and IGS data. Bootstrap support values are shown above the nodes.

variation of a single species. *Bryoria tortuosa* can thus be considered as a synonym of *B. fremontii*; see below for a detailed synonymy. Specimens of *B. fremontii* in which vulpinic acid is present in abundance will henceforth be referred to as '*B. tortuosa*'.

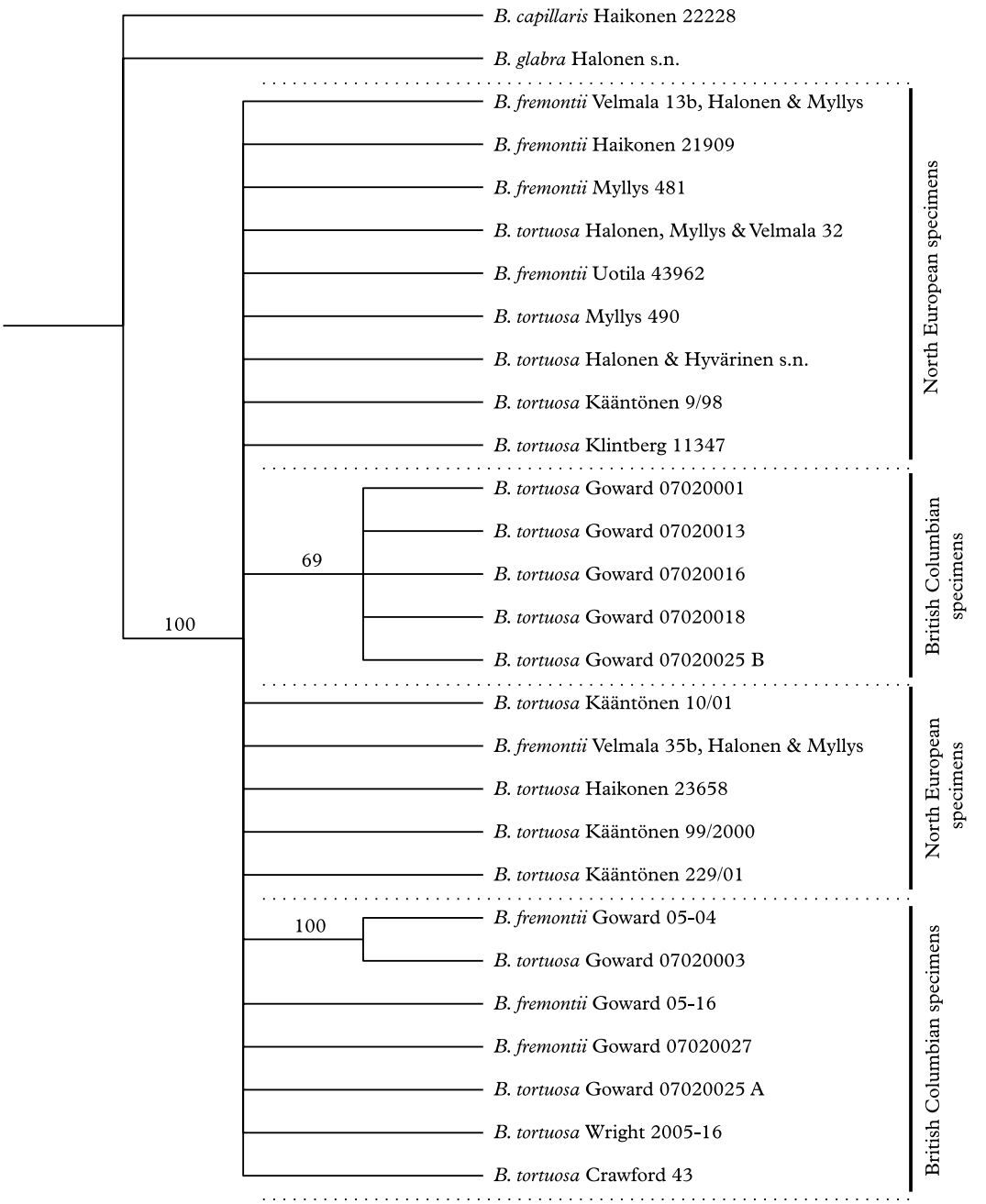

FIG. 2. Molecular phylogeny of *Bryoria fremontii* and *B. tortuosa*. A strict consensus tree of 468 trees based on ITS, IGS, mtSSU and GAPDH data. Bootstrap support values are shown above the nodes.

The amount of homoplasy was extremely low in both analyses as indicated by high CI and RI values. This suggests that the low resolution in the strict consensus trees (Figs 1 & 2) reflects a lack of information (i.e., almost identical ingroup sequences) rather

than conflicting characters between different gene regions. The partition-homogeneity test detected significant incongruence between GAPDH and IGS regions as well as between GAPDH and ITS regions. Because of the low amount of homoplasy, the incongruence between these regions might be explained by, for example, recombination. This implies that usually asexually reproducing *Bryoria fremontii* may have a capability for sexual reproduction.

Recent studies have successfully used the ITS, IGS, mtSSU and GAPDH regions in several intrageneric and population studies of lichenized fungi (e.g., Myllys *et al.* 2002; Printzen *et al.* 2003; Argüello *et al.* 2007; Lohtander *et al.* 2008). IGS is usually more variable than ITS and can be expected to contain sufficient information to separate genetically isolated populations (Printzen & Ekman 2002; Lindblom & Ekman 2006, 2007). The mtSSU region usually contains highly variable portions, which are difficult to align and are thus sometimes removed from parsimony analyses (e.g., Mangold *et al.* 2008). In this study, the mtSSU was the most informative region. However, the relatively high information content resulted mostly from a long deletion at the end of the mtSSU present in two specimens. When the deletions were excluded, the ingroup sequences were almost identical.

As already discussed, we were unable to find any significant correlation between chemistry and phylogeny, as both vulpinic acid-rich specimens ('*Bryoria tortuosa*') and vulpinic acid-deficient specimens (*B. fremontii*) appeared in the same clade. The factors regulating the production or distribution of vulpinic acid are unknown. According to Stephenson & Rundel (1979) and Golojuch & Lawrey (1988), light intensity and substratum have no effect on the concentration of vulpinic (and pinastric) acid in either *Letharia vulpina* (L.) Hue or *Vulpicida pinastri* (Scop.) J.-E. Mattsson. These results may be species-specific, but according to Hermansson & Thor (2004), Crawford (2007) and our own observations, *B. fremontii* and '*B. tortuosa*' can grow in mixed populations under the same environmental conditions. This fact could indicate that the variation in vulpinic acid concentration is not connected to the possible ecological differences (Holien 1986) between *B. fremontii* and '*B. tortuosa*'. On the other hand, extensive field experience by one of us (TG) in western North America strongly suggests that '*B. tortuosa*' is much more frequent here in summer-dry regions than in more humid regions. Crawford (2007), in his western Canadian study of *B. fremontii* and *B. tortuosa*, showed that: 1) vulpinic acid-rich specimens usually grow intermixed with vulpinic acid-deficient specimens, while the vulpinic acid-deficient specimens, for their part, often form non-mixed populations; and 2) vulpinic acid-rich specimens tend to be localized within certain areas and even on certain trees within those areas. These observations are consistent with the hypothesis that '*B. tortuosa*' may arise from time to time in populations of *B. fremontii*, the enhanced production of vulpinic acid being triggered by some environmental factor or factors as yet unknown. Here it can be noted that a somewhat similar mechanism may operate between *Calicium glaucellum* Ach., which lacks vulpinic acid, and *C. trabinellum* (Ach.) Ach. which contains it. Here again the presence of vulpinic acid correlates with summer-dry forests (T. Goward, pers. obs.). Interestingly, it appears that European '*B. tortuosa*' is never as rich in vulpinic acid as its North American counterparts, i.e., the production of vulpinic acid seems to be more abundant in North America.

The Species

The citations below include some corrections to earlier data.

Bryoria fremontii (Tuck.) Brodo & D. Hawksw.

Opera Bot. **42:** 136 (1977).—*Alectoria fremontii* Tuck., *Am. J. Arts Sci.* ser. 2, **25:** 422 (1858); type: USA, California, Sierra Nevada, 'camp of Dec. 5–6', 1845, *J. C. Frémont* (FH-Tuck. 498—holotype!).

New synonym: *Bryoria tortuosa* (G. Merr.) Brodo & D. Hawksw., *Opera Bot.* **42:** 138 (1977).—*Alectoria tortuosa* G. Merr., *Bryologist* **12:** 5 (1909); type: Canada, British

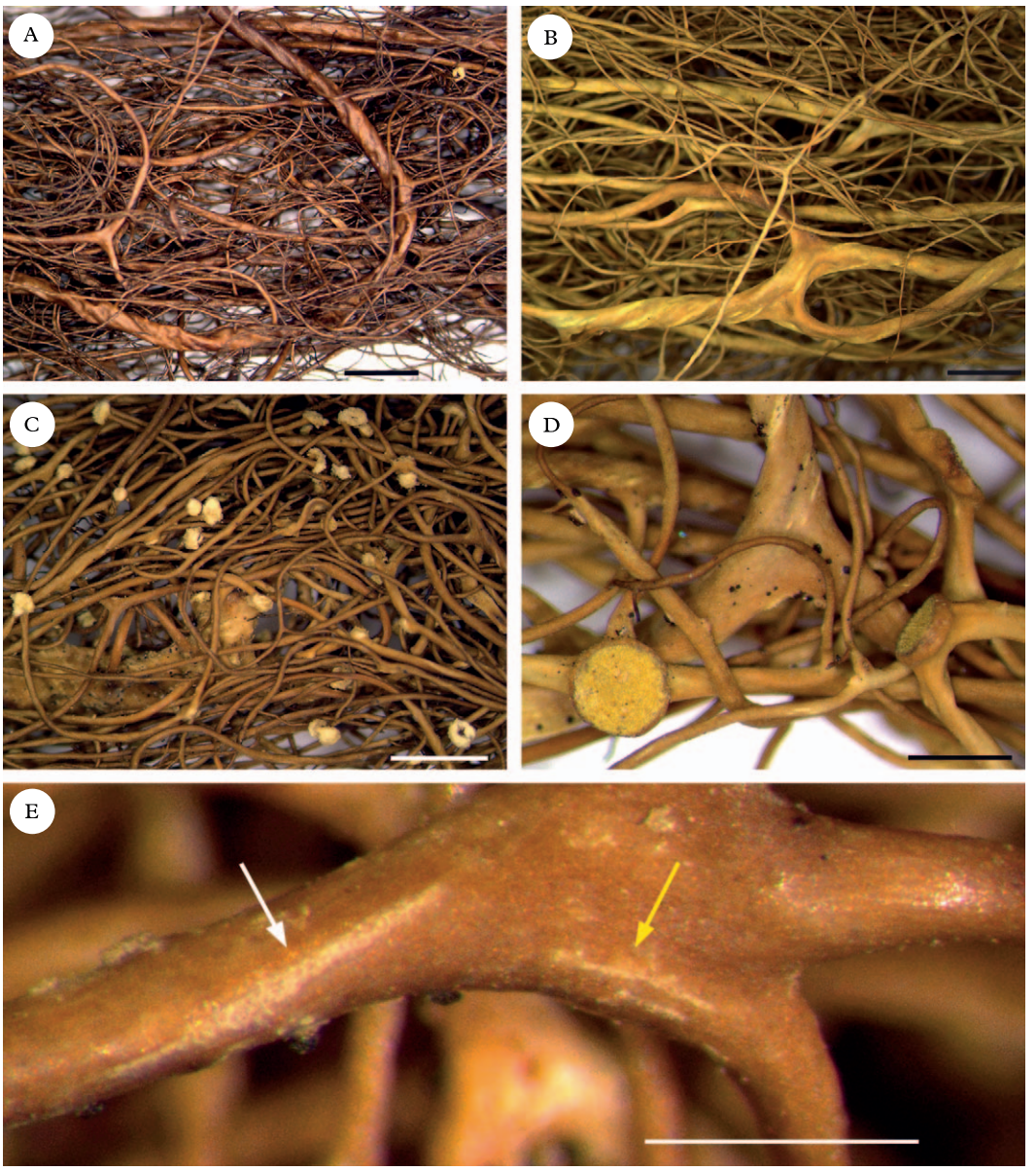

FIG. 3. *Bryoria fremontii*. A, typical vulpinic acid-deficient chemotype showing reddish brown hue and foveolate branches (*Velmala* 13b *et al.*); B, typical vulpinic acid-containing chemotype showing yellow-brown hue and yellow pseudocyphellae (*Goward* 07020025); C, thallus with yellow soralia and pseudocyphellae (*Crawford* 52); D, thallus with yellow apothecia and pseudocyphellae (*Crawford* 32); E, branch with both yellow (yellow arrow) and white (white arrow) pseudocyphellae as seen in fresh material (*Crawford* 52). Scales: A – C = 2 mm; D = 1 mm; E = 0·5 mm.

Columbia, near New Westminster, July 1904, *A. J. Hill* (FH-hb. G. K. Merrill—holotype!).

Alectoria olivacea Räsänen, *Meddeland. Soc. Fauna Fl. Fenn.* **43:** 4 (1917).—*Alectoria fremontii* subsp. *olivacea* (Räsänen) Räsänen, *Luonnon Ystävä* **23:** 10 (1919); type: Finland, Ostrobottnia ultima, Simo, 6 Jun 1916, *V. Räsänen* (H—lectotype!, designated here; CANL, H, NY, OULU, TUR, UPS—isolectotypes!).

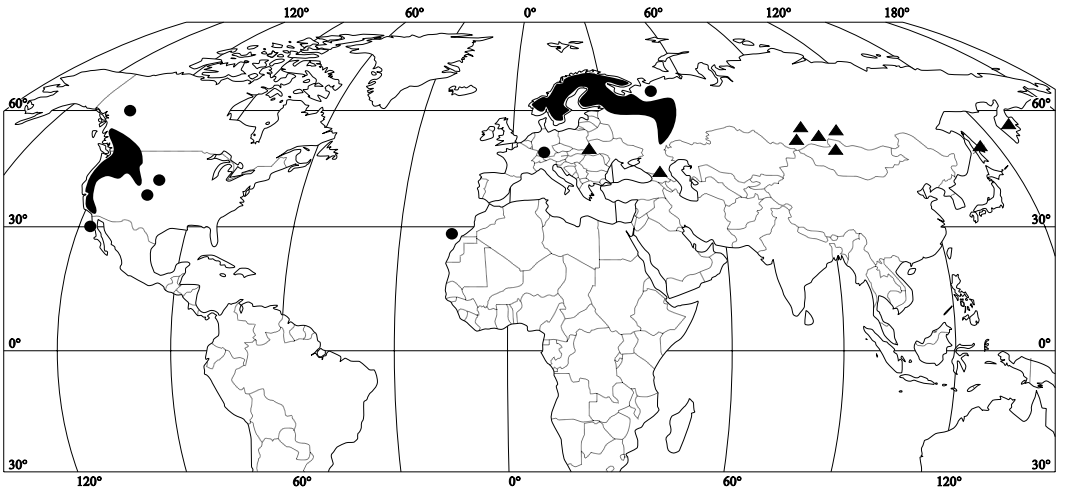

FIG. 4. The global distribution of *Bryoria fremontii* based on Ahlner (1948) and Brodo & Hawksworth (1977), with later additions from the literature as well as herbarium material. The record from the Canary Islands is by Hawksworth (1982). The records accepted from Russia include those in the following papers: Hermansson & Kudryavtseva 1997: 218, Pystina & Hermansson 1998 (map), Petrova 2000 (map), Kravchenko 2003, Urbanavichus & Urbanavichene 2004 (in part), Chkobadze 2004 (map), Kuznetsova *et al*. 2007, Fadeeva & Kravchenko 2007 (map), Fadeeva *et al*. 2008 and Urbanavichus *et al*. 2008. Records from the Altay, Sayan and other mountains in the south of Siberia, Mongolia, the Carpathians, the Caucasus, the Kamchatka Peninsula and Sakhalin (e.g., Sedel'nikova 1985, 1990, 1998; Byazrov *et al*. 1989; Mikulin 1990; Oxner 1993) could not be confirmed and are mapped as doubtful records (indicated with triangles).

Alectoria corneliae Gyeln., *Ann. Crypt. Exot*. **4:** 171 (1931); type: Oregon, Linn Co., near Brownsville, 25 May 1930, *L. L. Haskin* (US—holotype!).

For further synonyms see Brodo & Hawksworth (1977: 136 & 138).

The synonymous name *Alectoria olivacea*, given to a non-sorediate morph of *Bryoria fremontii*, was lectotypified in H by Brodo & Hawksworth (1977). However, there are several specimens with identical labels, and they did not annotate any of them. Therefore it was necessary to make a second-step lectotypification and restrict the type designation to one specimen. Isolectotypes exist in several other herbaria.

(Figs 3 & 4)

Thallus pendent, up to 60 cm long, dull yellow to reddish, pale or dark brown, colour varying within the same specimen (i.e., depending on the concentration of brown pigment and vulpinic acid), cortex dull or shiny; branching mainly anisotomic dichotomous, angles between the dichotomies acute, sometimes with perpendicular side branches, branches uneven in diameter, the broadest ones often twisted and foveolate, sometimes flattened; soralia absent or sparse, sometimes abundant, tuberculate, bright to pale yellow; pseudocyphellae abundant or sparse to almost absent, when present usually conspicuous, elongate or fusiform, white to bright yellow, colour sometimes varying within the same specimen; true lateral spinules and isidia absent; apothecia rare.

For a detailed description of vegetative tissues, ascoma and spore characters, see Brodo and Hawksworth (1977).

Chemistry. Contains vulpinic acid in the thallus, soralia and apothecia, though often absent from the thallus *per se*. Barbatolic and norstictic acids rarely present.

Distribution and habitat. *Bryoria fremontii* occurs predominantly in western North America (Canada, USA, Mexico) and northern Europe (Norway, Sweden, Finland, Russia), with outliers in Germany and the Canary Islands, mostly on conifers, but also on deciduous trees in open or shady forests. The global distribution of *Bryoria fremontii* is presented in Fig. 4.

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